

Pharmaceutical Biology



ISSN: 1388-0209 (Print) 1744-5116 (Online) Journal homepage: informahealthcare.com/journals/iphb20

Stimulation of Autonomic Cholinoceptors in the Rat Uterus by a Crude Extract from Eleophorbia drupifera Leaves

A.E. Eno & E.H. Itam

To cite this article: A.E. Eno & E.H. Itam (1998) Stimulation of Autonomic Cholinoceptors in the Rat Uterus by a Crude Extract from Eleophorbia drupifera Leaves, Pharmaceutical Biology, 36:2, 97-102, DOI: 10.1076/phbi.36.2.97.4600

To link to this article: https://doi.org/10.1076/phbi.36.2.97.4600



STIMULATION OF AUTONOMIC CHOLINOCEPTORS IN THE RAT UTERUS BY A CRUDE EXTRACT FROM *Eleophorbia drupifera* Leaves

A.E. Eno and E.H. Itam

Department of Physiology, College of Medical Sciences, University of Calabar, PMB 1115, Calabar, Cross River State, Nigeria

ABSTRACT

The crude extract from E. drupifera was tested for its effects on isolated rat uterine preparations. The extract $(2.0{\text -}300~\mu\text{g/ml})$ produced graded increases in uterine contractions. The contractions were potentiated by neostigmine $(1.64\times 10^{-6}\text{M})$ and abolished by atropine $(4.4\times 10^{-4}\text{M})$. The extract-induced contractions were not prevented by previous addition to the bath of hexamethonium or 5-hydroxytryptamine and bradykinin antagonist. However, extract-induced increase in uterine contraction was abolished by either lidocaine $(4.2\times 10^{-5}\text{M})$ or tetrodotoxin $(5\times 10^{-8}\text{M})$. These data seem to indicate that the crude extract-induced increase in myometrial contractility is due to actions on postganlionic autonomic nerve endings, with acetylcholine release and stimulation of muscarinic receptors.

INTRODUCTION

Plants of the Euphorbiaceae are frequently used in the indigenous system of medicine. Little is known about the pharmacological actions of the species *Elaeophorbia drupifera* (Thonn.) Stapf, although it is listed among the plants that "heal" (Ampofo, 1977). The leave is used as a filaricide and for guinea worm infestation (Comley, 1990). It is also reported to contain hypoglycemic agent(s) (Eno & Itam, 1995). The fruit is succulent (Kinghorn & Evans, 1974) but the latex has skin irritant effects (Kinghorn & Evans, 1975).

Keywords: Cholinoceptors, Eleophorbia drupifera, rat uterus.

Address correspondence to: Dr. A.E. Eno, Department of Physiology, College of Medical Sciences, University of Calabar, P M B 1115, Calabar, Cross River State, Nigeria.

Apart from the use of *E. drupifera* leaves for the treatment of various ailments, traditional herbalists claim it is also effective in aiding or inducing labor or abortion. Ground leaves (paste) are dissolved in either water or soft drink (cola), and administered orally in doses determined by age.

Also, although the innervation of the mammalian uterus has been studied extensively, precise information regarding the intrinsic autonomic neural pathways and the relationship between the cholinergic and adrenergic components is lacking (Adham & Schenk, 1969; Garfield, 1986). Much work has been devoted to uterine sympathetic nerves with no corresponding interest concerning the cholinergic nerves, despite evidence for cholinergic innervation in the uterus of different mammalian species, particularly rat (Stjernquist & Owman, 1985; Hollingsworth, 1974; Adham & Schenk, 1969).

The main objective of this paper was to study the effects of *E. drupifera* leaf extract on isolated rat uterus in an attempt to elucidate the scientific basis for justifying the action of the herb as claimed by the herbalists, and possibly, speculate on its probable mode of action in the uterus. Some mediators of smooth muscle action outside the cholinergic and adrenergic systems, together with their specific antagonists, have also been considered.

MATERIALS AND METHODS

Preparation of the Crude Extract

Fresh leaves of *E. drupifera* were collected. The leaves were first washed free of soil and debris. Wash water was blotted off and the leaves ground to a paste. A quantity of the ground sample (50 g) was weighed and Soxhlet-extracted with 150 ml distilled water at 100°C for 8–10 h. Where larger ground samples (500–1000 g)

were used, extraction was done under reflux with an appropriate volume of water. The extract was slowly evaporated to dryness *in vacuo* at 40° C using a rotary evaporator. A starting sample of 50 g of fresh material gave a mean yield of 1.58 ± 0.47 g of extract (mean \pm SEM, n = 10) which was stored at -4° C until use. Weighed samples of the extract were then used to make up test solutions of the desired concentrations (mg/ml) (expressed in terms of the weight of extract).

Animal Preparation

Eighty non-pregnant female white Wistar rats weighing 180–250 g were treated with stilboestrol (1 mg/kg i.p.) 24 h prior to the experiments. The animals were killed by a single blow to the head. The abdomen was opened and the uterine horns were removed, freed of fat, and immersed in a modified Tyrode solution with the following composition in mmole/liter: NaCl 136.8; KCl 2.7; MgCl₂ 2.1; CaCl₂ 0.72; NaH₂PO₄ 0.4; NaHCO₃ 11.9; glucose 5.5 (pH 7.4).

One uterine horn segment measuring 3 cm was suspended in a 20 ml organ bath containing Tyrode solution, bubbled with atmospheric air and maintained at 37 \pm 1°C. The isometric recording of uterine musculature activity was done with a FT 03 transducer connected to a Washington recorder (model 400 D). The transducer was previously calibrated to establish a relationship between the force applied to the transducer and the gauge deflection (1 g = 25 mm). The preparations were allowed to equilibrate in the bath for 30 min before the experiments.

E. drupifera leaves extract, agonists, and antagonists drugs were directly added to the reservoir containing the uterus. The tissue preparation was washed once with the bathing solution (Tyrode solution) after addition of the extract or drugs, and allowed to rest for 2 min before the next addition. The effects of the extract were compared with those of the other substances utilized and the results were analyzed based on the amplitude of contractions (expressed as tension in grams) in the absence or presence of antagonists.

Statistical Analysis

Regression lines with confidence limits were calculated for the linear portions of log-concentration response curves. The log-concentration limits at 50% of the maximum response were used in the analysis of the significance of concentration differences as described by Birmingham et al. (1970). Maximum responses were compared by unpaired Student's *t*-test.

Drugs

Nicotine alkaloid (95–98%), atropine sulphate, hexamethonium bromide, tetrodotoxin, 5-hydroxytryptamine (5-HT), and acetylcholine chloride, were from Sigma (U.S.A.). Lidocaine hydrochloride (Cristalia, Brazil); neostigmine bromide (Roche, Brazil); bradykinin, methysergide and oxytocin were from Sandoz, Brazil. HOE 140:D-Arg [Hyp-3-di-5-D-Tic-7-Oc-8] bradykinin was from Hoechst, Japan.

RESULTS

Effects of Extract on Uterine Contractions

The rat uteri showed spontaneous and rhythmic contractions in Tyrode solution. Although the amplitudes of these contractions varied from one preparation to another, the mean amplitude was about 15.4 ± 3.8 mm (about 0.61 g tension), and this was regarded as the control tension.

Investigations have shown that the crude extract from $E.\ drupifera$ leaves (2–300 µg/ml) dose-dependently increased the amplitude of uterine contractions with an ED₅₀ value of 23.6 µg/ml (Fig. 3, control). High doses of the extract (above the ED₅₀ value) produced sustained contractions which were characterized by a rise in the basal tone of the tissue preparation. Based on this ED₅₀ value, test doses of 5 and 30 µg/ml were selected.

The latency of response was less than 2 s in all tissue preparations studied. When the tissue was exposed to the extract for about 30 min without wash, the effect of the extract (15 μ g/ml) on the tissue was maintained to about 58.5% at the end of 30 min exposure. However, a longer period (about 2 h) was required before the tissue could return to its resting tension. However, it was possible to terminate the action of the extract as desired, by washing the preparation with Tyrode solution.

Effect of Drugs on Extract-induced Uterine Contractions

Results are summarized in Table 1.

Atropine Sulphate and Neostigmine

Addition of neostigmine ($1.64\times10^{-6}\,\mathrm{M}$) to the bath did not evoke any response. However, neostigmine ($1.64\times10^{-6}\,\mathrm{M}$) in Tyrode solution, potentiated the effects of the extract ($5\,\mu\mathrm{g/ml}$) (Fig. 1b) and acetylcholine (ACh) ($1.1\times10^{-7}\,\mathrm{M}$) (Fig. 1c). When the extract or ACh was added to neostigmine ($1.64\times10^{-2}\,\mathrm{M}$) - treated uterus, sustained contractions were observed (Figs. 1b and c). However, atropine sulphate ($4.4\times10^{-4}\,\mathrm{M}$) abolished

Table 1. Influences of drugs on the contractions induced by the crude extract, acetylcholine, oxytocin, 5-HT and bradykinin on isolated rat uterus.

Treatment	N	Extract (30 µg/ml)	Acetylcholin $(1.1 \times 10^{-8} \text{M})$	Oxytocin $(1 \times 10^{-2} \text{UI})$	5-HT $(2 \times 10^{-5} \mathrm{M})$	Bradykinin $(3.8 \times 10^{-10} \mathrm{M})$
None	10	2.58 ± 1.85	2.44 ± 2.09	2.62 ± 2.31	2.67 ± 2.57	2.55 ± 3.06
TTX (5 × 10^{-8} M)	10	0	1.42 ± 1.76	_	_	1.35 ± 1.78
Lidocaine $(4.2 \times 10^{-5} \text{M})$	10	0	1.55 ± 2.45	1.69 ± 1.48	_	1.71 ± 2.59
Atropine $(4.4 \times 10^{-4} \text{M})$	12	0	0	2.41 ± 2.63	_	1.67 ± 1.75
Hexamethonium (2 \times 10 ⁻⁶ M)	8	2.64 ± 2.58	2.73 ± 1.93	1.72 ± 3.11	_	_
Methysergide $(1.1 \times 10^{-5} \text{M})$	5	2.23 ± 1.97	2.35 ± 1.55	_	0	1.78 ± 2.45
Hoe 140 (7.6 \times 10 ⁻⁸ M)	8	2.41 ± 3.32	1.75 ± 2.66	_	_	0

The isolated rat uterus was immersed in a 25 ml Tyrode solution at $37 \pm 1^{\circ}C$ and constant oxygenation.

the sustained contractions produced by the extract or ACh (Figs. 2a and b), but failed to abolish oxytocin-induced contractions (Fig. 2f).

Tetrodotoxin (TTX)

The addition of the crude extract (30 μ g/ml) to the bath solution caused sustained contractions of the uterus. This effect was abolished by TTX (5 \times 10⁻⁸ M) (Fig. 2d). After repeated washing, the uterus started to con-

tract again with the same pattern as observed in TTX-free preparations. TTX also abolished oxytocin-induced contractions (Fig. 2e).

Lidocaine

When lidocaine hydrochloride (4.2×10^{-5} M) was added to the bath, the contractile effect of the extract was abolished (Fig. 2c). However, the tissue recovered from this effect after repeated washing.

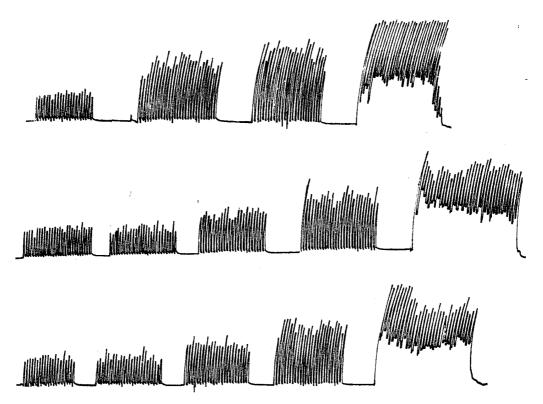


Fig. 1. Typical recordings showing the effects of the extract (5, 15 and 30 μ g/ml) on the isolated uterine muscle (a); the potentiation of extract's effect on the tissue by neostigmine (1.64 \times 10⁻⁶ M and 1.64 \times 10⁻² M) in Tyrode solution (neo-Tyr) (b); and of ACh-induced contractions (c). The tissue preparation was washed after each drug treatment.

N = Number of experiments. The blockade is expressed as 0, - indicates experiments were not performed.

The values for each group are the means \pm S.E.M. of the tension (grams).

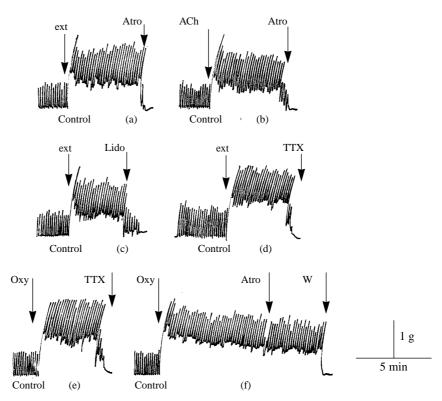


Fig. 2. Mechanical recordings showing the effects of atropine (4.4 \times 10⁻⁴ M) (a); lidocaine (4.2 \times 10⁻⁵ M) (c); TTX (5 \times 10⁻⁸ M) (d) on the contractions induced by the extract (30 µg/ml). Atropine also blocked ACh-induced contractions (b). Contractions caused by oxytocin (1 \times 10⁻² M) were blocked by TTX (5 \times 10⁻⁸ M) (e); but not by atropine (4.4 \times 10⁻⁴ M) (f). The tissue was washed (W) with Tyrode solution after each drug treatment.

Methysergide (5-HT Blocker)

5-Hydroxytryptamine (2 \times 10⁻⁵ M) produced increased uterine contractions which were prevented by methysergide (1.13 \times 10⁻⁵ M). This dose of methysergide failed to prevent the increase in uterine contractions produced by the extract (30 μ g/ml) (not shown).

Hexamethonium (Hex.)

The ganglion blocker, Hex. $(1.9 \times 10^{-6} \, \text{M})$ failed to abolish the contractions produced by the extract (30 µg/ml) or acetylcholine $(1.1 \times 10^{-7} \, \text{M})$ (not shown).

Nicotine

Nicotine (from 1×10^{-7} to 1×10^{-3} M) did not increase uterine contractions (not shown).

Bradykinin Blocker (HOE 140)

The addition of $7.6\times10^{-8}M$ HOE 140 to the bath was able to prevent the contraction induced by 3.8×10^{-10} M bradykinin, but had no effect on the contraction induced by 30 μ g/ml extract (not shown).

Agonist Action of the Extract on the Uterus

Concentration–response curves were established by graded increases in the concentration of the crude extract (2.0–300 µg/ml). The bathing solution of the tissue was then changed to atropinized-Tyrode solutions. Atropine concentrations in Tyrode were: $1\times 10^{-9}\,\text{M}, 1\times 10^{-6}\,\text{M}$ or $1\times 10^{-4}\,\text{M}$. From the results (Fig. 3) the presence of atropine (1 \times 10⁻⁹ – 1 \times 10⁻⁴ M) in the bathing fluid dose-dependently inhibited extractinduced increases of uterine contractions, and shifted the dose-response curve of the extract (control) to the right. Also, the atropine inhibition of contractions was surmounted by raising the concentration of the extract in the bathing fluid. The atropine antagonism is probably of a competitive type.

DISCUSSION

Clinically, drugs used to induce labor or abortion contract the uterine smooth muscle. Such drugs include

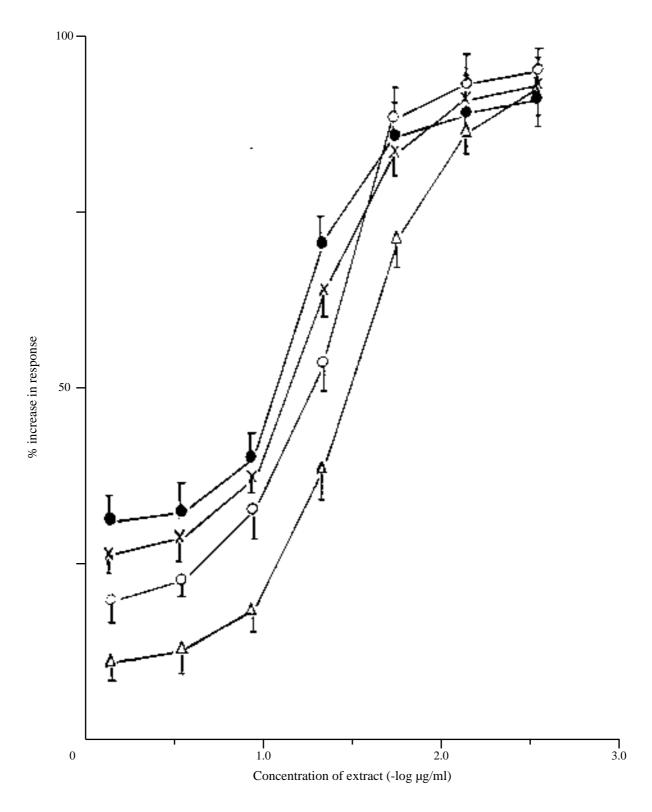


Fig. 3. Mean contraction of the isolated uterus to graded increases in the concentration of extract (2–300 µg/ml). Responses (% controls) in the absence (\bullet) or presence of 1×10^{-9} M (X), 1×10^{-6} M (\bigcirc), and 1×10^{-4} M (\triangle) atropine sulphate in Tyrode solution. Each point represents the mean value \pm S.E.M., n = 5, P < 0.05 compared with controls by unpaired Student's t-test.

oxytocin, ergometrine and quinine (Bowman & Rand, 1980). From the results of the present study, there is sufficient evidence to believe that the crude extract from *E*. drupifera leaves contains a uterus-contracting agent(s). Small doses of the extract increased the amplitude of spontaneous uterine contractions, and large doses produced sustained spasms of the uterus, which was evidenced by the rise in basal tension of the tissue preparations. These tissue responses to the extract are similar to that reported for oxytocin and other standard uterus-contracting agents (Bowman & Rand, 1980). It was observed that the uterus-contracting action of the extract was fast in onset and could be totally terminated by washing with extract-free Tyrode solution. This may indicate the low molecular weight compound present in the extract, which may penetrate rapidly to its site of action. However, the fast onset of action exibited by the extract could merely reflect the high concentration of the active compound present.

Additions of specific antagonists of 5-HT and bradykinin to the bath prevented uterine contractions induced by these mediators, but did not prevent the effect of the extract. Therefore, the data indicate that the uterus contraction elicited by the extract is not related to the activity of these mediators. As the effects of the extract are potentiated by neostigmine and prevented or abolished by atropine, it seems likely that the action of the extract depends on stimulation of cholinergic muscarinic receptors. The atropine antagonism could be competitive in nature, since it was surmounted by raising the concentration of the extract. The data (Table 1) showed that TTX did not prevent the contractions induced by ACh or bradykinin, but totally abolished the contractions elicited by the extract. These probably suggests the interaction of the extract with the sodium channels. Therefore, it is likely that the extract acts on the sodium channels, releasing ACh from postganglionic nerve fibers with subsequent stimulation of muscarinic cholinoceptors. These findings are consistent with reports on the autonomic innervation of the rat uterus (Hollingsworth, 1974; Garfield, 1986; Mendonca et al., 1995). Hexamethonium (Hex) abolishes nicotinic actions just as it abolishes the actions of nicotine itself (Bowman & Rand, 1980). This probably explains why this ganglion blocking drug (Hex) failed to abolish the contractions produced by the extract, suggesting that the uterine smooth muscle cells could be devoid of nicotinic receptors, or that the extract was acting post-synaptically.

However, it is premature to speculate on the actions of this extract on smooth muscle since it may contain more than one active compound in its crude stage. Further progress must await refinements in its separation techniques.

ACKNOWLEDGEMENTS

The authors are indebted to Mr. Daniel and Mr. D.D. Dakat, of the University of Jos, Nigeria; for their technical assistance and to the University of Calabar, Calabar, Nigeria, for the research grant.

REFERENCES

Adham N, Schenk EA (1969): Autonomic innervation of the rat vagina, cervic and uterus and its cyclic variation. *Am J Obstet Gynec* 15: 508–516.

Ampofo C (1977): *Plants that heal*. World Health, pp. 26–30. Birmingham AT, Paterson G, Wojiciki J (1970): Comparison of the sensitivities of innervated and denervated rat vasa deferentia to agonist drugs. *Br J Pharmacol* 30: 748–754.

Bowman WC, Rand MJ (1980): Autonomic nervous system-Neurochemical transmission in smooth muscle. In: *Text-book of Pharmacology*, 2nd ed., pp. 9.27–11.45. London, Blackwell Scientific Publ.

Comley JCW (1990): New macrofilaricidal leads from plants. *Trop Med Parasitol 43*: 1–9.

Eno AE, Itam EH (1995): Hypoglycemic agent(s) in leaves of *Elaeophorbia drupifera*. *Phytotherapy Res* 10: 680–682.

Garfield RE (1986): Structural Studies of innervation on nonpregnant rat uterus. *Am J Physiol* 251: C41–C54.

Hollingsworth M (1974): The innervation of the rat cervix and its pharmacology *in vitro* and *in vivo*. *Br J Pharmac* 52: 539–547.

Kinghorn AD, Evans FJ (1974): Occurrence of ingenol in *Elaeophorbia* species. *Planta Med 26*: 150–154.

Kinghorn AD, Evans FJ (1975): The succulent *Euphorbias* of Nigeria. *Lloydia 38*: 363–365.

Mendonca M, Profeta DL, Freire-mala L, Cunha-melo JR (1995): Effect of scorpion toxin from *Tityus serrulatus* on the contraction of the isolated rat uterus. *Toxicon 33*: 355–361

Stjernquist M, Owman C (1985): Cholinergic and adrenergic neural control of smooth muscle function in the non-pregnant rat uterine cervix. *Acta Physiol Scand 124:* 429–463.

Accepted: August 14, 1997