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ORIGINAL ARTICLE

Effect of graded hyperbaric atmospheric pressure on the quantity and composition of faecal flora

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Abstract

Objectives: The ecosystem of the intestinal flora is always dynamic and it is maintained even in the changing chemical and physical environment of the host. The effects of the physical environment such as atmospheric pressure on the composition of intestinal flora has not been studied previously. Considering this, an *in vitro* experiment was designed to explore the effects of hyperbaromatric pressure on the faecal flora of the rat. *Materials and methods:* Faecal samples from male albino rat were exposed to graded hyperbaric conditions (122 and 170 kilopascal, kPa) for different time periods (1, 3 and 5 h). Total aerobes, total anaerobes, *Escherichia coli, Bifidobacterium* spp. and *Clostridium perfringens* present in the faecal samples were quantified by specific culture-based methods. Variation in their number in comparison to control conditions was analysed statistically (ANOVA and Tukey t test). *Results:* The numbers of total aerobes and *E. coli* were increased with the increase in air pressure, whereas a reduction in numbers was recorded for total anaerobes, *Bifidobacterium* spp. and *C. perfringens*. Variations of these groups of bacteria in relation to dose and duration of hyperbaric treatment were also recorded. *Conclusion:* Air pressure is an important exogenous factor that strongly regulates the composition of the faecal flora.

Key words: barometric pressure, faecal flora, Clostridium perfringens, Bifidobacterium spp

Introduction

The ecology of microbes in the intestine is fascinating. Numerous interactions between host and microorganism are continually occurring there in a complex fashion (1,2). Bacteria are the predominant inhabitants of the alimentary tract of mammals and they are designated as indigenous microflora or popularly known as normal flora (3). They perform several beneficial roles for the host, such as the breakdown of undigested food, enhanced absorption of foodstuffs, metabolism of drugs, synthesis of vitamins, creation of resistance against pathogenic bacteria by colonization resistance, induction of host immunity and stimulation of intestinal maturation (4,5). The gastrointestinal microecosystem is always fluctuating. This condition arises due to the high sensitivity of microflora to numerous host-induced physio-chemical and environmental factors such as antimicrobial agents, disorders of peristalsis, inflammatory bowel diseases, cancer, stress, redox potential, drugs, temperature and nutrients (6,7). The microflora is also highly sensitive to oxygen tension (8,9) and that can be correlated with the atmospheric pressure.

During activities such as deep-sea diving, digging tunnels beneath a river or mining, individuals are exposed to hyperbaric atmosphere (an increase of 0.1 kilopascal (kPa) air pressure per 1 cm drop in sea level; with a sea level pressure of 101.3 kPa. This can create various kinds of health hazards (6). Among them, the major problems are indigestion and gas formation (flatulence). Although such problems are mainly associated with the alteration of the composition of gastrointestinal microflora (4), there is still no detailed record that can correlate the variations in composition of indigenous microflora with atmospheric pressure.

The bacterial community of the gastrointestinal tract is not well understood owing to the inadequacy of classic culture-dependent methods. Recently many advanced techniques have been developed but no single technique could give an overall view

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of the total bacteria (4). In the present study, selective oxygen-sensitive culturable bacteria like total aerobes and anaerobes, and some indicator bacteria like *Escherichia coli*, *Bifidobacterium* spp. and *Clostridium perfringens* of the faecal matter (normally found in the gastrointestinal tract of the rat) were selected and their quantitative variation was studied by exposing them to graded hyperbaric pressure.

Materials and methods

Animals and diet

Healthy male albino rats with an average body weight of 115 ± 7 g were used in the present study. They were housed in metal cages ($34 \times 28 \times 19$ cm). All animals had access to boiled rat feed (carbohydrates, 74.05%; proteins, 10.38%; fibre, 2.20%; iron, 56 ppm; calcium, 400 ppm and sodium, 500 ppm) and water *ad libitum*. The animals were maintained without interrupting their normal activity.

Sample collection and treatment

Rat faeces were collected just after dropping onto clean paper underlying the cage. Fresh faecal sample was suspended in sterilized phosphate-buffered saline (PBS; pH 7.0 and 9 g 1^{-1} NaCl) using a manual glass homogenizer for 5 min. The faecal homogenates were exposed to two different simulated hyperbaromatric pressures (122 and 170 kPa, these are 1.2 and 1.7 times higher than normal atmospheric pressure, respectively) for different time periods (1, 3 and 5 h). The suspension was then centrifuged (1000 g for 5 min) and clear supernatant was used for microbial analysis.

Analytical measurements

The total aerobic and anaerobic faecal bacteria were enumerated by standard pour-plate technique in single-strength trypticase soya agar (TSA, Himedia, India) and reduced Wilkins Chalgren agar (WCA, Micromaster, India), respectively. An anaerobic jar was employed for anaerobic culture, from which oxygen was removed catalytically and replaced with 10% of both CO_2 and H_2 gas (Micromaster, Mumbai). Enumeration of *E. coli* and *Bifidobacterium* spp. was carried out using selective media – Mac-Conkey and bifidobacterium agar (Himedia), respectively. Reduced perfringens agar base (Himedia) was used for selective enumeration of *C. perfringens*.

Statistical analysis

Collected data are presented as the arithmetic mean of three replicas (mean \pm SE). The variations in microbial count were examined by one-way ANOVA and the multiple comparisons of all possible pairs were done by Tukey *t* test (SPSS-10.0). The alteration in bacterial quantity at different air pressures (122 and 170 kPa) for each specific time period (1, 3 and 5 h) was tested by Fisher's *t* test. Significant variation was accepted at the level of 5%, i.e. p < 0.05.

Results

The effect of graded hyperbaric pressures (122 and 170 kPa) for different durations (1, 3 and 5 h) on the faecal flora was evaluated and is presented in Table I. In control conditions (normobaric), rat faeces (per gram) contained total aerobes 1.9×10^6 , total anaerobes 3.7×10^{11} , *E. coli* 9.5×10^5 , *Bifidobacterium* spp. 4.0×10^4 and *C. perfringens* 3.7×10^5 .

When the faecal matter was subjected to higher atmospheric pressure of 122 and 170 kPa for 1, 3 and 5 h, the quantity of total aerobic bacteria was increased up to 10-fold (3.2×10^7) polynomially $(R^2 = 0.9389)$, step by step (-0.05-fold, 0.25-fold and 0.42-fold); and about 100-fold (1.4×10^8)

Table I. Quantification of gastrointestinal flora after hyperbaric treatment for various time periods.

Type of bacteria	Normal count (cfu g^{-1})	Pressures applied (kPa)	Colony-forming units (cfu g^{-1}) at different durations		
			1 h	3 h	5 h
Total aerobes	$1.9\pm0.17\times10^{6}$	122	$1.8 \pm 0.19 imes 10^{6}$	$2.3 \pm 0.213 imes 10^{6}$	$3.2 \pm 0.29 \times 10^{7}$
		170	$9.0 \pm 1.12 \times 10^{6}$	$1.9 \pm 0.21 \times 10^7$	$1.4 \pm 0.13 \times 10^8$
Total anaerobes	$3.7 \pm 0.42 imes 10^{11}$	122	$3.5 \pm 0.33 \times 10^9$	$3.5\pm0.35\times10^{7}$	$1.7 \pm 0.18 imes 10^{6}$
		170	$1.5 \pm 0.12 imes 10^9$	$4.8 \pm 0.51 imes 10^{6}$	$1.0 \pm 0.21 imes 10^4$
E. coli	$9.5 \pm 1.23 imes 10^5$	122	$8.5 \pm 0.82 imes 10^4$	$0.6 \pm 0.11 imes 10^{6}$	$0.5\pm0.09\times10^7$
		170	$1.5 \pm 0.14 imes 10^5$	$0.5 \pm 0.08 imes 10^7$	$1.5 \pm 0.15 imes 10^8$
Bifidobacterium spp.	$4.0 \pm 0.07 \times 10^{4}$	122	$1.3 \pm 0.14 imes 10^4$	$2.7 \pm 0.32 imes 10^2$	$1.1\pm0.21\times10^2$
		170	$3.1 \pm 0.31 imes 10^4$	$1.7 \pm 0.17 imes 10^2$	$4.1\pm0.43\times10^2$
Clostridium perfringens	$3.7 \pm 0.21 imes 10^5$	122	$1.7 \pm 0.17 imes 10^4$	$0.9 \pm 0.13 \times 10^{3}$	$0.2 \pm 0.09 imes 10^2$
		170	$1.5 \pm 0.12 imes 10^4$	$0.14 \pm 0.05 imes 10^2$	Not detectable

Data are presented as mean \pm SE.

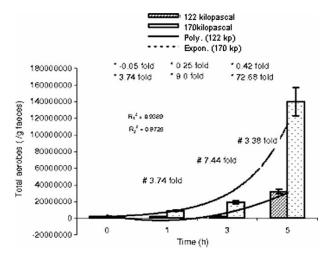


Figure 1. Comparative study of population size of total aerobes with time at 122 and 170 kPa. R_1^2 , regression coefficient (122 kPatime); R_2^2 , regression coefficient (170 kPa-time); *step-wise; –, reduction in population size; +, increase in population size; #, time-specific, \top , standard error of mean.

exponentially ($R^2 = 0.9728$), step by step (3.74-fold, 9.0-fold and 72.68-fold) (Figure 1 and Table I). The alterations in aerobic bacteria at both the atmospheric pressures were statistically significant (p < 0.05). Increasing the pressure by about 40% favoured the bacterial population to multiply by 3.74, 7.44 and 3.38 times at the incubation times of 1, 3 and 5 h, respectively ($t_{1h} = 426.34$, p < 0.05; $t_{3h} = 981.46$, p < 0.05 and $t_{5h} = 29090.35$, p < 0.05).

Interestingly, the count of *E. coli* mounted gradually and reached the level of the total aerobic population polynomially ($R_1^2 = 0.9794$ and $R_2^2 =$

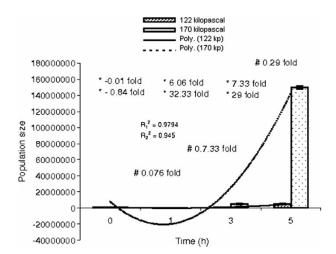


Figure 2. Comparative study of population size of *E. coli* with time at 122 and 170 kPa. R_{12}^2 regression coefficient (122 kPa-time); R_{22}^2 , regression coefficient (170 kPa-time); *step-wise; –, reduction in population size; +, increase in population size; #, time-specific; \top , standard error of mean.

0.945) after graded hyperbaric treatment (Figure 2 and Table I).

In rat faeces, total anaerobes, Bifidobacterium spp. and C. perfringens were normally present at a ratio of $1:1.08 \times 10^{-7}:1 \times 10^{-6}$ (Table I). The quantity of total anaerobes was reduced by 10⁵- and 10⁷fold at 122 and 170 kPa pressure, respectively, in comparison with their normal counts (p < 0.05 and p < 0.05, respectively) (Table I). In addition, the count of anaerobes also declined at above atmospheric pressure (120-170 kPa) for the three time periods tested by 57.14%, 89.09% and 99.41%, respectively $(t_{1h} = 54632.283, p < 0.05; t_{3h} = 786.64, p < 0.05 and$ $t_{5h} = 292.09, p < 0.05)$ (Figure 3). The counts of Bifidobacterium spp. and C. perfringens were abridged up to 10^2 -fold and 10^3 -fold, respectively (p < 0.05 and p < 0.05), at both the tested pressures (Table I). On increasing the pressure from 122 kPa and 170 kPa, the population of *Bifidobacterium* spp. declined by 24.4-, 5.11- and 2.73-fold at 1, 3 and 5 h, respectively $(t_{1h} = 101.17, p < 0.05; t_{3h} = 367.64, p < 0.05 and$ $t_{5h} = 63.48, p < 0.05$) (Figure 4), but a drastic reduction in number was observed in C. perfringens $(10.91\%, t_{1h} = 120.88, p < 0.05; 98.46\%, t_{3h} =$ 264.97, p < 0.05 and 100%, $t_{5h} = 52.08$, p < 0.05) (Figure 5).

Discussion

Microbial flora present in the microenvironment of the gastrointestinal tract performs several important and essential activities of the host (10). Diverse types of bacteria are associated in this ecosystem and among them a few have been well characterized (11). Their homeostasis can be disrupted by a wide variety of indigenous and exogenous factors (1). The

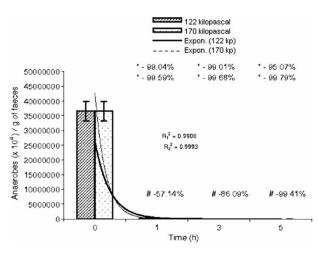


Figure 3. Comparative study of population size of total anaerobes with time at 122 and 170 kPa. R_1^2 , regression coefficient (122 kPatime); R_2^2 , regression coefficient (170 kPa-time); *step-wise; –, reduction in population size; +, increase in population size; #, time-specific; \top , standard error of mean.

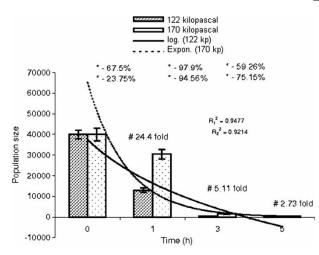


Figure 4. Comparative study of population size of *Bifidobacterium* spp. with time at 122 and 170 kPa. R_1^2 , regression coefficient (122 kPa-time); R_2^2 , regression coefficient (170 kPa-time); *stepwise; –, reduction in population size; +, increase in population size; #, time-specific; \top , standard error of mean.

effects of graded hyperbaric pressures on the microbes present in the faecal matter were examined. Two hyperbaric pressures, i.e. 122 and 170 kPa (equivalent to depths of 2.07 m and 6.87 m, respectively, from sea level) were chosen in this study, as humans generally encounter such air pressures during sea-diving and cleaning of underground municipal drainage, wells, tunnels, etc.

In the faeces of rat (normobaric), the aerobic, facultative (*E. coli*) and anaerobic bacteria were present in a ratio of $2:1:3.89 \times 10^5$. This ratio varies from species to species (12). Abundance of anaerobic bacteria in faeces is common in all animals due to their expanded large intestine. It has been found that the population of facultative bacteria is half of the total aerobes, whereas Samanta et al. (4) reported

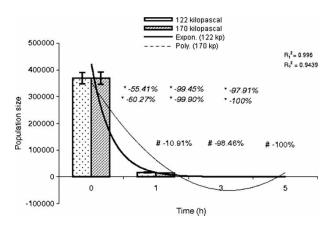


Figure 5. Comparative study of population size of *Clostridium* perfringens with time at 122 and 170 kPa. R_1^2 , regression coefficient (122 kPa-time); R_2^2 , regression coefficient (170 kPa-time); *stepwise; –, reduction in population size; +, increase in population size; #, time-specific; \top , standard error of mean.

that the quantity of *E. coli* in rat faeces is approximately one-third of the total aerobic microflora.

Faecal matter exposed to graded hyperbaric pressure for different time periods showed an increased number of total aerobes and *E. coli*. Normally bacteria are very sensitive to atmospheric oxygen and the limit of oxygen tolerance varies from species to species (13,14). In this experiment the reasons for elevation of aerobes and facultative bacteria at high atmospheric pressure is not clear but it may be due to accelerated oxidation of cellular metabolites and over-activation of different oxygensensitive rate-limiting enzymes of growth-related metabolism in the presence of high levels of molecular oxygen (15). Our observations are in agreement with the study of Gillmore et al. (16).

The anaerobic bacteria are the predominant inhabitants of the lower intestine; they have several beneficial effects and also maintain the normal health of the colon (3). In recent years, bifidobacteria and C. perfringens have been of great interest because of their nutritional and health impacts (17-19). In the present study, the quantity of the anaerobic group of organisms was drastically reduced at graded simulated air pressure in a duration-dependent manner (Table I). In excess oxygenation most of the anaerobic groups of bacteria were eradicated and this may be due to the production of reactive oxygen species (ROS). Normally this group of bacteria creates an anaerobiosis by reducing the action potential of its surroundings (20). In hyperbaric conditions, alteration of the potential creates an unfavourable environment for their survival.

Conclusion

It is clear from the study results that atmospheric pressure above the ambient level alters the composition of faecal flora. Although the selected groups of bacteria are very limited members of the overall microbial population in the gastrointestinal tract, the observed results indicate that atmospheric pressure has a significant impact on the colonization of such a group of microflora.

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Declaration of interest: The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

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