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BK_{Ca} channel is a molecular target of vitamin C to protect against ischemic brain stroke

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ABSTRACT

Epidemiological studies have demonstrated that vitamin C decreases the risk of stroke, which has generally been ascribed to its function as antioxidant and free radical scavenger. However, whether there is a defined molecular target for vitamin C on stroke is unknown. Utilizing middle cerebral artery occlusion (MCAO) in rats as a model for ischemic stroke, we demonstrated that long-term, low-dose administration of vitamin C prior to MCAO could exert significant neuroprotective effect on the brain damage. The long-term, low-dose vitamin C pretreated rats had decreased brain infarct size and decreased neurological deficit score compared with the vehicle or single high dose pretreated MCAO rats. Furthermore, electrophysiological experiments using patch clamp technique showed that vitamin C increased the whole-cell current of the large-conductance Ca^{2+} -activated K⁺ (BK_{Ca}) channel. Moreover, vitamin C increased the open probability of the channel without change its amplitude. Importantly, blockade of the BK_{Ca} channels abolished the neuroprotective effect of vitamin C on MCAO. Therefore, this study shows that long-term, low-dose pretreatment with vitamin C could reduce MCAO-induced brain damage through activation of the BK_{Ca} channels, suggesting that the BK_{Ca} channel is a molecular target of vitamin C on stroke.

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KEYWORDS

lschemic stroke; largeconductance Ca²⁺-activated K⁺ channel; middle cerebral artery occlusion; patch clamp; vitamin C

Introduction

Vitamin C (also known as Ascorbic acid) is best known for its role as an essential nutrient in humans. A number of epidemiologic studies have shown that dietary intake of vitamin C or high plasma vitamin C concentrations are associated with a lower risk of stroke or stroke-related mortality (Ascherio et al., 1999; Del Rio et al., 2011; Gale et al., 1995). It has been shown more than two decades ago that vitamin C concentration is strongly related to the risk of death from stroke but not from coronary heart disease in elderly people (Gale et al., 1995). This association has been confirmed in the subsequent studies. From a prospective study of 20,649 participants aged 40-79 years with an average follow-up of 9.5 years, it has been shown that the risk of stroke over 10 years of follow-up was 42% lower in subjects with baseline plasma vitamin C levels \geq 66 µmol.L⁻¹ compared to those with vitamin C levels $<41 \,\mu\text{mol.L}^{-1}$ (Myint et al., 2008). In addition, a meta-analysis of prospective studies has also demonstrated that both dietary vitamin C intake and circulating vitamin C are significantly inversely associated with the risk of stroke in a dose-response manner (Chen et al., 2013a). In consistence with the beneficial effect of vitamin C on stroke, multiple lines of evidence also support that a high consumption of fruits and vegetables is associated with a reduction in the risk of stroke (Dauchet et al., 2005; Joshipura et al., 1999; Johnsen et al., 2003; Lakkur & Judd, 2015; Padayatty & Levine, 2008).

On the other hand, several phase III randomized controlled trials have suggested that vitamin C supplementation has no effect on stroke prevention (Myung et al., 2013). Animal experiments also showed that vitamin C treatment was not associated with significant improvements in either infarct size or neurological function (Huang et al., 2001). These results are seemingly contradictory to the above epidemiologic studies. However, dietary intake represents long-term habitual exposure of vitamin C before the onset of stroke, whereas supplementation is generally characterized by shorter duration and higher dose. Thus, if

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vitamin C plays a preventive but not therapeutic role in stroke, the trials enrolling high-risk participants may not be able to detect any significant associations (Chen et al., 2013a). For example, it has been shown that administration of vitamin C (500 mg.day⁻¹, iv) to ischemic stroke patients for 10 days beginning on day 1 after ischemic stroke did not substantially improve the clinical and functional status of patients after 3 months (Lagowska-Lenard et al., 2010).

In the present study, to study whether vitamin C has protective but not therapeutic effect on ischemic stroke, we administrate vitamin C intraperitoneally once a day for 3 weeks to rats before middle cerebral artery occlusion (MCAO). Furthermore, we provided several lines of evidence to confirm whether there is a clear molecular target for the effect of vitamin C on ischemic stroke.

Experimental procedure

Cell culture and transfection

Chinese hamster ovary (CHO-K1) cells were cultured in Ham's F12 nutrient mixture supplemented with 10% FBS at 37 °C in a 5% CO₂ incubator. Cells were plated onto poly-L-lysine-coated coverslips in 35 mm dishes at 60–80% confluence. Then they were transiently cotransfected with the full length BK_{Ca} channel gene and GFP (Clontech, Palo Alto, CA, USA). Transfection was performed with LipofectAMINE Plus reagent (Invitrogen) following the manufacturer's instructions. Cells were used for electrophysiological recordings 1–2 days after the transfection.

Whole-cell recording

Whole-cell currents were measured using a conventional tight seal whole-cell recording technique at room temperature. Pipettes had resistance at a range of 2–5 M Ω in recording solution containing (in mM) 145 KCl, 10 EGTA, 10 HEPES (pH 7.3 with KOH). The intracellular free Ca^{2+} concentration was 1 μ M. The extracellular solution was Hanks' balanced salt solution (HBSS, Sigma, in mmol.L⁻¹): 1.3 CaCl₂, 0.8 MgSO₄, 5.4 KCl, 0.4 KH₂PO₄, 136.9 NaCl, 0.3 Na₂PO₄, 10 D-glucose and 4.2 NaHCO₃. Compensation for cell capacitance and series resistance was made automatically using EPC-10 patch-clamp amplifiers (HEKA, Lambrecht, Germany); only recordings with stable series resistance less than 25 M Ω were included in the study. Currents were sampled at 2-5 kHz and filtered at 1.5-2.9 kHz using a 4-pole low-pass Bessel filter. Program packages Patchmaster 2.1 and TAC 4.1

(HEKA) were used for data acquisition and analysis. The rundown of the whole cell current of the BK_{Ca} channels is apparent at first 10 min after formation of the whole-cell configuration. To overcome this problem, each of the chemicals was added to judge its effect on the current only after the current was stable for 3 min period, which usually takes around 15 min.

Single-channel recording

The single-channel recordings were carried out at inside-out patch-clamp configuration using an EPC-10 amplifier (HEKA Elektronic, Lambrecht, Germany) at room temperature. Data were acquired at 2-5 kHz and low-pass filtered at 1 kHz. Patch electrodes were pulled from thin walled borosilicate capillary tubes using a Sutter programable puller (model P-97, Sutter Instrument, Novato, CA, USA) and fire-polished to achieve an electrical resistance in a range of 3–7 M Ω . An Ag/AgCl wire bath electrode was used as ground electrode. The pipette solution was HBSS. The bath solution consisted of (in mmol.L⁻¹): 145 KCl, 10 EGTA, 2 Mg-ATP, 10 HEPES (pH 7.3 with KOH). The Po was simply calculated from the total time spent in the open state divided by the total time of recording for the patches containing a single channel. Continuous recordings of 2000-4000 ms were used to estimate Po. The Po in a patch with multiple channels was calculated by using TAC 4.1 (HEKA, Germany), based on the equation: $Po = (1 - P_C^{1/N})$, where P_c is the probability that all channels are in the closed state, N is the number of channels in the patch, which is estimated from the maximum number of channels observed.

Measurement of vitamin C concentration in serum and brain tissue

To obtain the serum sample, whole blood of each rat was sampled in a vacutainer tube. Serum was generated by centrifugation at 3000 g in 20 min at 4°C and immediately frozen to -80°C until further handled and analyzed. At the time of analysis, the serum sample (200 µl) was homogenized in a centrifuge tube with 500 µl vitamin C extracting solvent (0.35 mol.L⁻¹ perchloric acid, 0.27 mmol.L⁻¹ EDTA-Na and 0.1% dithiothreitol). After centrifugation at 1000 g in 5 min at 4°C, the supernate was filtrated by a 0.22 µm membrane filter. To obtain samples of the brain tissue, 100 mg brain tissue from each rat was homogenized in a mixture of 1.0 ml dithiothreitol (20 mg.ml⁻¹). After addition of 9.0 ml metaphosphoric acid (3%), the brain samples were eddied, sonicated for 10 min and centrifuged at 4000 g for 5 min after addition of 5 ml n-hexane. Then, the aqueous phase was filtrated by a 0.22 µm membrane filter. Vitamin C concentrations of the above samples were analyzed by an Ultra Performance Liquid Chromatography (UltiMate 3000, Dionex company, USA). Chromatographic separation was achieved by using an Agilent XDB-C18 column $(5 \,\mu\text{m}, 4.6 \times 150 \,\text{mm}, \text{USA})$. A $10 \,\mu\text{l}$ sample (in 5%) methanol) was injected into the column at a flow rate of 0.8 ml.min^{-1} . The compounds were eluted using 95% mobile phase A (a solution of acetic acid at PH3) and 5% mobile phase B (100% methanol). The detection wavelength was set at 254 nm and the time of equilibrium was 30 min. The vitamin C concentrations of the samples were determined from the standard curves generated from vitamin C standards.

Transient Middle cerebral artery occlusion

A total of 297 male Sprague-Dawley rats (weighing 200-250 g, Xiamen University animal experiment center) were used throughout the study. Among them 17 rats died during experiment, which were excluded from the study. Details on the number of the rats in each experiment were shown in the corresponding figures. The transient middle cerebral artery occlusion (MCAO) was induced by using intraluminal filament insertion technique as previously described (Chi et al., 2010). Briefly, the filament thread (with a blunt tip of 0.36 ± 0.02 mm in diameter, type 2636-A3, Beijing Cinontech, China) was inserted through the left common carotid artery and advanced into the internal carotid artery to occlude the origin of the middle cerebral artery (approximately 18-20 mm) for 24 hours. Body temperatures were monitored and controlled at 37±0.5°C with a homoeothermic blanket. The shamoperated rats were treated identically, except that the middle cerebral artery was not occluded after the neck incision. All the rats were given unrestricted access to food and water. The animals were sacrificed 24 hr post-MCAO. Brains were rapidly removed, sectioned coronally at 2 mm intervals, and stained by 2,3,5triphenyltetrazolium hydrochloride (TTC). The slices were photographed, and analyzed with Image-J software (NIH freeware) and presented with Photoshop software (Adobe Systems Inc., San Jose, California, USA). Infarct size in all slices was expressed as a percentage of the contralateral hemisphere after correcting for edema. All manipulations and analyses were performed by individuals blinded to all groups. Animal care and experiments were performed in accordance with procedures approved by the Animal Care and Use Committee of Xiamen University (accreditation number XMULAC20170250).

Drug administration

Vitamin C diluted in 0.9% saline was administrated intraperitoneally. Rats in the control group were given an equal volume of the vehicle (0.9% saline). Penitrem A (PA) and Paxilline (Pax) were dissolved in DMSO, diluted in 0.9% saline and then were applied intraperitoneally.

Neurological evaluation

Neurological deficit scores were evaluated after 24 hr of MCAO according to the Longa 5-point scale scoring system (Longa et al., 1989): grade 0, no deficit; grade 1, failure to fully extend the right forepaw; grade 2, spontaneous circling or walking to a contralateral side; grade 3, walking only when stimulated; grade 4, unresponsive to stimulation and a depressed level of consciousness; and grade 5, death. Grades of the neurological score were evaluated by an investigator blinded to the treatment protocol.

Statistics

The results were presented as mean \pm SEM. The Origin 7.0 (Originlab, Northampton, MA, USA) was used for statistical analyses and plotting graphs. Statistical comparisons were made by using one-way ANOVA, followed by Bonferroni's post hoc test. Differences at p < 0.05 were considered to be statistically significant.

Results

Preventive effect of long-term, low-dose vitamin C on MCAO rats

To investigate whether vitamin C has preventive effect on focal cerebral ischemia, we tested effect of long-term (once a day for 3 weeks) intraperitoneal administration of vitamin C on the rats prior to MCAO. At 24 hr post-MCAO, the brain slices were stained with TTC (Figure 1(a)), which stains healthy tissue red and leaves infarct tissue pale white. Statistical results showed that the percentage of infarct size was $22.77 \pm 0.61\%$ in the vehicle-treated MCAO rats, and the percentages of the infarct size were reduced to $20.83 \pm 1.15\%$, $19.58 \pm 0.89\%$, $20.46 \pm 0.67\%$, $17.85 \pm 1.93\%$ for the rats treated with 0.5, 5, 20 and 50 mg.kg⁻¹.day⁻¹ vitamin C, respectively (Figure 1(b)). In addition, the neurological deficit



Figure 1. Effect of long-term, low-dose vitamin C on MCAO induced infarct size and neurological deficit score. (a) Representative images of TTC stained brain slices obtained at 24 hr post-MCAO from MCAO rats pretreated with vitamin C at a dose of 0, 0.5, 5, 20 and 50 mg.kg⁻¹.day⁻¹ for 3 weeks. (b,c) Bars showing the relative infract size (b) or neurological deficit score (c) of MCAO rats pretreated with vitamin C at a dose of 0, 0.5, 5, 20 and 50 mg.kg⁻¹.day⁻¹ for 3 weeks. *p < 0.05; **p < 0.01; ***p < 0.005.

score (Figure 1(c)) was 2.03 ± 0.12 for the vehicletreated MCAO rats, and 1.87 ± 0.09 , 1.60 ± 0.10 , 1.69 ± 0.13 , 1.57 ± 0.20 for the rats treated with 0.5, 5, 20 and 50 mg.kg^{-1} .day⁻¹ vitamin C, respectively. As vitamin C at a dosage of 5 mg.kg^{-1} .day⁻¹ gave the significant protective effect on both infarct size and neurological deficit score, the dosage of 5 mg.kg^{-1} was used in most of the following studies. To test whether more frequently administration of vitamin C could have more beneficial effect on MCAO, we compared the effect of administration of 2.5 mg.kg^{-1} vitamin C twice a day with that of 5 mg.kg^{-1} once a day on MCAO. Representative images of TTC stained brain slices were present in Figure 2(a). Statistical results showed that even though administration of 2.5 mg.kg⁻¹ vitamin C twice a day had beneficial effect on infarct size (Figure 2(b)) and neurological deficit score (Figure 2(c)), there was no significant difference between the groups administrated with 2.5 mg.kg^{-1} twice a day and 5 mg.kg^{-1} once a day, suggesting that more frequently administration of vitamin C does not have more beneficial effect on MCAO. Next, we measured vitamin C concentration in the serum and brain tissue after administration of vitamin C for three weeks. Before MCAO (24 hr after the last administration of vitamin C), the concentration of vitamin C in both serum (Figure 3(a)) and brain tissue (Figure 3(b)) was significantly increased from the groups administrated with 5 mg.kg^{-1} vitamin C once a day or 2.5 mg.kg⁻¹ vitamin C twice a day. However, there was no significant difference between the groups administrated with 5 mg.kg^{-1} vitamin C once a day and the group administrated with 2.5 mg.kg^{-1} vitamin C twice a day. After 24 hr post-MCAO (i.e., 48 hr after the last administration of vitamin C), vitamin C concentration in the serum was significantly increased for both MCAO and vitamin C treated MCAO groups (vs. Sham operated group) (Figure 3(c)). Interestingly, the vitamin C concentration in the ipsilateral hemisphere was significant higher in the vitamin C treated MCAO group than that in the vehicle treated MCAO group (Figure 3(e)), even though there were no significant difference in that of the contralateral hemisphere (Figure 3(d)). The big differences of vitamin C levels in contralateral and ipsilateral brain might due to the influence of tissue damage. In addition, we found that administration of a single high dose (500 mg.kg⁻¹) vitamin C



Figure 2. Application of vitamin C twice a day does not have more beneficial effect on ischemic stroke than that applied once a day. (a) Representative images of brain slices obtained at 24 hr post-MCAO from MCAO rats pretreated with vehicle as well as pretreated with vitamin C at a dose of 5 mg.kg⁻¹ once a day or 2.5 mg.kg⁻¹ twice a day for 3 weeks. (b,c) Bars showing the relative infract size (b) and neurological deficit score (c) of MCAO rats pretreated with vehicle and vitamin C at a dose of 5 mg.kg⁻¹ once a day or 2.5 mg.kg⁻¹ twice a day for 3 weeks. (b,c) Bars showing the relative a day or 2.5 mg.kg⁻¹ twice a day or 2.5 mg.kg⁻¹ twice a day or 2.5 mg.kg⁻¹ twice a day (2*2.5 mg.kg⁻¹) for 3 weeks. *p < 0.05; **p < 0.01; ***p < 0.005.

has no effect on MCAO (Figure 4(a-c)) and did not significantly change the vitamin C concentration in the serum (Figure 4(d)) and brain tissue (Figure 4(e)) before MCAO (24 hr after the last administration of vitamin C). These results suggested that pretreament with long-term, low-dose vitamin C could reduce the MCAO induced brain damage, while a single high dose had no effect.

Vitamin C activates BK_{Ca} channels

Many researches (Chi et al., 2010; Gribkoff et al., 2001; Hewawasam et al., 2003; Li et al., 2014; Su et al., 2017) have shown that BK_{Ca} channel is one of the molecular targets of the ischemic stroke. Therefore, we tested whether vitamin C could activate the BK_{Ca} channel. First, we investigated the effect of vitamin C on the whole-cell currents of the BK_{Ca} channels expressed in CHO cells. Under control condition, for which the procedure was the same except that no vitamin C was added, the whole cell BK_{Ca} current was decreased after 10 min, probably due to the rundown of the current (Figure 5(a)). Addition of 200 and 500 μ M vitamin C into the bath solution clearly increased the BK_{Ca} channel current (Figure 5(b,c)). Statistical summary showed that the ratio of the whole cell BK_{Ca} channel current at a voltage of + 60 mV before and after 10 min of vitamin C treatment was 0.76 ± 0.06 in control condition. In contrast, the ratio values increased to 1.26 ± 0.08 and 1.39 ± 0.10 after 10 min addition of 200 and 500 μ M vitamin C into the extracellular side of the cells (Figure 5(d)).

The above result indicated that vitamin C could increase the whole cell current of the BK_{Ca} channel.



Figure 3. Effect of long-term and low dose pretreatment with vitamin C on the concentration of vitamin C in the serum and brain. Concentration of vitamin C in the serum (a) or right and left brain (b) for the rats treated with vehicle or treated with vitamin C at a dose of 5 mg.kg⁻¹ once a day or 2.5 mg.kg⁻¹ twice a day for three weeks. The concentration of vitamin C was measured 24 hr after the last administration of vitamin C. **p < 0.01, ***p < 0.005. (c–e) Concentration of vitamin C in the serum (c) and contralateral side (d) or ipsilateral side (e) of the brain for sham-operated, vehicle pretreated MCAO rats and vitamin C pretreated MCAO rats. Vitamin C was intraperitoneally administrated at a dose of 5 mg.kg⁻¹ once a day or 2.5 mg.kg⁻¹ twice a day for three weeks prior to MCAO. The concentration of vitamin C was measured 48 hr after the last administration of vitamin C.

However, it is not known whether cytosolic factors are involved in the channel activation, whether the increased current is due to an increase in the single channel open probability (Po) or the amplitude of the single channel. To address these questions, we turned to the inside-out patch-clamp configuration. Figure 6(a) showed the single channel current traces at membrane potential ranging from -40 to +60 mV without vitamin C, indicating that the Po of channels increased with the depolarization of the membrane potential. In contrast, application of 200 µM vitamin C further increased the Po of the channel at every corresponding membrane potential without changing the single channel amplitude (Figure 6(b)), suggesting that the increase in the whole cell current by vitamin C was due to increase in Po of the single channel but not its single channel amplitude. Next, we studied the effect of vitamin C on the voltage dependent activation of the BK_{Ca} channel. The Po of the BK_{Ca} channels as a function of the membrane potential with or

without vitamin C was fitted by the Boltzmann equation: Po = $1/{1 + e^{[(V_{1/2} - V)/k]}}$, where V_{1/2} is the voltage required for half-maximal activity of the channel and k is the slope factor of the curve. The best fit to the data was obtained with values of 96.7 mV for $V_{1/2}$ and 15.4 mV for k in the absence of vitamin C and with values of 90.7 mV for $V_{1/2}$ and 16.4 mV for k in the presence of $200 \,\mu\text{M}$ vitamin C (Figure 6(c)). Therefore, vitamin C shifted the voltage activation curve toward less positive membrane potentials without affecting the slope factor of the curve, which suggests that vitamin C does not alter the voltagesensitivity of the channel. Figure 6(d) showed the statistical data on the Po of the channel in the control condition and in the presence of vitamin C. In the control condition, Po was 0.031 ± 0.002 , it was slightly decreased to 0.026 ± 0.004 in the presence of $50 \,\mu\text{M}$ vitamin C. In contrast, the Po was significantly increased to 0.062 ± 0.006 , 0.061 ± 0.008 and 0.068 ± 007 when vitamin C concentration was



Figure 4. A single high dose vitamin C has no effect on MCAO. (a) Representative images of brain slices obtained at 24 hr post-MCAO from rats treated with vehicle or treated with a single high dose vitamin C (500 mg.kg^{-1}) 24 hr before MCAO. (b,c) Bar graphs showing the relative infract size (b) and neurological deficit score (c) for a single high dose vitamin C-treated (500 mg.kg^{-1}) vs vehicle-treated MCAO rats. (d,e) Vitamin C concentration in the serum (d) or brain tissue (e) of rats after 24 hr treament with vehicle or a single high dose vitamin C (500 mg.kg^{-1}).



Figure 5. Vitamin C activates BK_{Ca} channels at the whole-cell level. (a–c) Representative whole-cell BK_{Ca} channel currents before (upper panel) and after 10 min (lower panel) application of 0 (a), 200 (b) and 500 (c) μ M vitamin C in the bath solution at Ca²⁺ concentration of 1 μ M. The currents were evoked by step depolarization from a hold potential of -80 mV to test potentials between -80 and +60 mV for 200 ms in 20 mV increments. (d) Ratio values (I/Io) of the whole-cell BK_{Ca} channel current at + 60 mV before (lo) and after 10 min (I) application of different concentration of vitamin C in the bath solution. **p < 0.01.



Figure 6. Vitamin C activates BK_{Ca} channels at the single-channel level. (a,b) Representative single channel current traces of the BK_{Ca} channels in the inside-out configuration at membrane potentials ranging from -80 mV to +60 mV with 20 mV increment before (a) and after application of 200 μ M vitamin C (b) in the bath solution. Arrows indicate the level that all the channels are in their closed state. (c) The relationships between the single channel open probability (Po) of the BK_{Ca} channels and the membrane potentials in the absence and presence of 200 μ M vitamin C are fitted with the Boltzmann equation (see text for detail). (d) Statistical data on Po of the channel at a voltage of +20 mV in the absence and presence of 50, 200, 500 and 1000 μ M vitamin C. ***p < 0.001.

increased to 200, 500 and 1000 μ M at the intracellular side of the patch, respectively. Because inside-out patches are detached from the cell, the result indicated that cytosolic factors were not involved in the activation of the BK_{Ca} channel by vitamin C.

Blockade of BK_{Ca} channels abolishes the neuroprotective effect of vitamin C on MCAO

If the BK_{Ca} channel is the molecular target of vitamin C, then its specific blockers should suppress the neuroprotective effect of vitamin C on MCAO. Representative images in Figure 7(a) showed that penitrem A (PA), a lipid soluble specific BK_{Ca} channel blocker, could suppress the effect of vitamin C on the infarct size of the MCAO rats. Figure 7(b) showed that the percentage of the infarct size was $23.60 \pm 0.92\%$ in the vehicle-treated MCAO rats. Application of vitamin C (5 mg.kg⁻¹.day⁻¹ for three weeks) before MCAO decreased the percentage of the infarct size to $18.26 \pm 1.01\%$. In contrast, application of vitamin C could not decrease the percentage of the infarct size to $16.26 \pm 1.01\%$.



Figure 7. BK_{Ca} channel blocker Penitrem A (PA) suppresses the effect of vitamin C on MCAO rats. (a) Representative images of brain slices for vehicle-treated MCAO rats and vitamin C pretreated (5 mg.kg^{-1} .day⁻¹ for three weeks prior to MCAO) MCAO rats at 24 hr post-MCAO in the absence and presence of PA (0.4 and $4 \mu \text{g.kg}^{-1}$). (b,c) Bar graphs showing relative infarct size (b) and neurological deficit score (c) in vehicle-treated and vitamin C pretreated MCAO rats at 24 hr post-MCAO in the absence and presence of PA.

when PA was given immediately at post-MCAO: the percentage of the infarct size was $21.5 \pm 1.24\%$ and $22.84 \pm 2.18\%$ when PA at a dose of 0.4 and $4 \mu g.kg^{-1}$ was applied immediately after MCAO (Figure 7(b)). Statistical data showed that application of PA also abolished the effect of vitamin C on the neurological deficit score (Figure 7(c)). The neurological deficit score was 2.09 ± 0.16 for vehicle-treated MCAO rats. Application of vitamin C decreased this value to 1.58 ± 0.15 , while 0.4 and $4 \mu g. kg^{-1}$ PA-treatment increased this value to 1.93 ± 0.19 and 2.14 ± 0.14 . To further confirm whether BK_{Ca} channel is the molecular target, we investigated the effect of another blocker of BK_{Ca} channel, paxilline (Pax), on the MCAO rats (Figure 8(a)). The percentage of the infarct size was 22.33 ± 0.98% in the vehicletreated MCAO rats. Vitamin C treatment decreased the value to $18.39 \pm 1.23\%$. In contrast, the value was 23.58 ± 1.99% when Pax was applied immediately after MCAO in the vitamin C pretreated group (Figure 8(b)). In addition, the neurological deficit score was 2.00 ± 0.12 in the vehicle-treated group, 1.36 ± 0.15 in the vitamin C-treated group, and 1.86 ± 0.14 in the vitamin C-treated group with i.p injection of Pax (Figure 8(c)). These results showed that vitamin C could protect MCAO-induced brain damage via activating the BK_{Ca} channels, suggesting that the BK_{Ca} channel is the molecular target of vitamin C.

Discussion

A limitation of this study is that the results were limited to an animal model and cannot be generalized to humans. Another limitation of the current study is that the neurological deficit score and infarct size were assessed only at one timepoint, the day after stroke, and this is only based on a neurological deficit score, and not on any functional measures. Despite of these limitations, in the present study, we showed that pretreatment with vitamin C at a dose of 5 mg.kg⁻¹ per day or 2.5 mg.kg⁻¹ twice a day for three weeks significantly reduced the infarct size induced by MCAO, while single, high dose



Figure 8. BK_{Ca} channel blocker paxiline (Pax) suppresses the effect of vitamin C on MCAO rats. (a) Representative images of brain slices for vehicle-treated MCAO rats and vitamin C pretreated (5 mg.kg⁻¹.day⁻¹ for three weeks prior to MCAO) MCAO rats at 24 hr post-MCAO in the absence and presence of Pax (5 μ g.kg⁻¹). (b,c) Bar graphs showing relative infarct size (b) and neurological deficit score (c) in vehicle-treated and vitamin C pretreated MCAO rats at 24 hr post-MCAO in the absence and presence of Pax.

administration of vitamin C (500 mg.kg^{-1}) had no effect, suggesting that long-term, low-dose intake of vitamin C has beneficial effect on the ischemic stroke. In consistence with this result, it has been reported that pretreatment with vitamin C (100 mg.kg^{-1} , p.o.) on rat with diabetic state for two weeks significantly suppressed the exacerbation of cerebral ischemic injury (Iwata et al., 2014). However, administration of vitamin C (500 mg.day $^{-1}$, iv) to ischemic stroke patients for 10 days beginning on day 1 after ischemic stroke did not substantially improve the clinical and functional status of patients (Lagowska-Lenard et al., 2010). Animal experiment also showed that there was no significant improvements in either infarct size or neurological function when vitamin C at a single dose of 500 mg.kg⁻¹ was administered 15 min after the ischemic insult (Huang et al., 2001). These results suggest that vitamin C plays preventive but not therapeutic role in stroke. This is in good agreement with the conclusion that a high consumption of fruits and vegetables is associated with a reduction in the risk of stroke (Dauchet et al., 2005; Joshipura et al., 1999; Johnsen et al., 2003; Lakkur & Judd, 2015; Padayatty & Levine, 2008), because dietary intake represents long-term habitual exposure of vitamin C before the onset of stroke. Besides, it has been demonstrated that high level of plasma vitamin C could significantly reduce the risk of stroke (Chen et al., 2013a; Gale et al., 1995; Myint et al., 2008).

There is evidence that a rapid increase in the production of reactive oxygen species immediately after acute ischemic stroke rapidly overwhelm antioxidant Defense, causing further brain damage (Rodrigo et al., 2013). Vitamin C acts as part of the intracellular antioxidant network (Padayatty et al., 2003; Rice, 2000). Therefore, the effect of vitamin C on stroke is usually ascribed to its function as antioxidant and free radical scavenger. On the other hand, there is also growing evidence that inflammation plays a key role in the ischemic brain stroke (Anrather & ladecola, 2016; McColl et al., 2007; Vidale et al., 2017). Thus, the effect of vitamin C on ischemic stroke may be due to the antiinflammatory function of vitamin C (Wannamethee et al., 2006). In addition, vitamin C intake has also been demonstrated to be inversely associated with blood pressure (Block et al., 2008; Juraschek et al., 2012). Hence, vitamin C may reduce stroke risk through its pressure-lowering effects. These putative mechanisms could somehow explain the protective effect of vitamin C on the ischemic brain stroke. However, it is not known whether there is a defined molecular target for the effect of vitamin C on the ischemic stroke. In this report, we show that the BK_{Ca} channel is the molecular target of vitamin C on the ischemic brain stroke, which is supported by several pieces of evidence. First, whole-cell recordings showed that vitamin C could enhance whole-cell BK_{Ca} current from extracellular side. Second, single channel recordings indicated that vitamin C could increase single channel open probability without change its amplitude of the BK_{Ca} channel. Most importantly, specific blockers of the BK_{Ca} channel abolished the effect of vitamin C on the MCAO rats.

Excess Ca^{2+} influx into the cells is one of the major pathophysiological mechanisms important for ischemic brain stroke. The BK_{Ca} channels are activated by either the membrane depolarization or the increase in intracellular Ca²⁺. They are expressed abundantly in the brain (Knaus et al., 1996; Sailer et al., 2006; Tseng-Crank et al., 1994). Their activation could prevent excessive Ca²⁺ buildup and abnormal glutamate release (Chi et al., 2010; Gribkoff et al., 2001) and are thus one of the candidates to protect against ischemic brain stroke (Tano & Gollasch, 2014). Indeed, it has been shown that activation of the $\ensuremath{\mathsf{BK}_{\mathsf{Ca}}}$ channel could exerts neuroprotective effect on the ischemic brain damage, which could be antagonized by the BK_{Ca} channel inhibitors (Chi et al., 2010; Gribkoff et al., 2001; Hewawasam et al., 2003; Li et al., 2014; Su et al., 2017). In organotypic hippocampal slice cultures, BK_{Ca} channel opener could reduce cell death induced by oxygen and glucose deprivation or glutamate exposion, which could be reversed by the BK_{Ca} channel blockers (Piwońska et al., 2016; Runden-Pran et al., 2002). In addition, it has been found that MCAO produced larger infarct volume, more severe neurological deficits, and higher post-ischemic mortality in BK(-/-) mice than that of the WT littermates (Liao et al., 2010). These results suggest that neuronal BK_{Ca} channels are important for protection against ischemic brain damage. In the present study, we showed that vitamin C could activate the BK_{Ca} channel. It is worth to mention that the activation of the BK_{Ca} channel by vitamin C is not as potent as many known BK_{Ca} channel openers. This might be beneficial to the ischemic brain damage, because it has been shown that the over-activity of the BK_{Ca} channels could result in more severe hippocampal neuronal damage (Chen et al., 2013b).

Taken together, the present study showed that long-term, low-dose administration of vitamin C could reduce the MCAO induced brain damage. Therefore, long-term, low level activation of the BK_{Ca} channels might be responsible for the neuroprotective effect of vitamin C on the ischemic brain stroke, suggesting that long-term, low-dose intake of vitamin C may serve as a potential strategy to protect against ischemic brain stroke in humans.

Contributors

Z.Q. designed research; L.L., S.L, C.H., L.Z. and Y.Z. performed research; Z.Q., M.Y. and L.L. analyzed data; Z.Q. and L.L. wrote the manuscript. All authors approved the final manuscript.

Disclosure statement

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