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To cite this article: Shu-Juan Zhou, Si-Qian Wang, Yong-Yong Ma, Li-Yuan Tang, Yi-Fen Shi, Bin Liang, Yi Chen & Kang Yu (2016) Association of proton pump inhibitors with the occurrence of gut-derived bacteraemia in patients with haematological malignancy after chemotherapy, Hematology, 21:6, 332-337, DOI: [10.1080/10245332.2016.1142711](https://doi.org/10.1080/10245332.2016.1142711)

To link to this article: <https://doi.org/10.1080/10245332.2016.1142711>



Published online: 10 Mar 2016.



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Association of proton pump inhibitors with the occurrence of gut-derived bacteraemia in patients with haematological malignancy after chemotherapy

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Background: Gut-derived bacteraemia is a major complication in patients with haematological malignancy after chemotherapy.

Objective: Our study aimed to investigate the role of proton pump inhibitors (PPIs) in the occurrence of gut-derived bacteraemia.

Methods: We compared data from 92 hospitalized haematological malignancy patients after chemotherapy with gut-derived bacteraemia, collected from January 2009 to July 2015, with those of 92 contemporaneous, hospitalized haematological malignancy patients without bacteraemia. We evaluated PPIs use and analysed the effects of covariates.

Results: Patients with gut-derived bacteraemia had a significantly higher incidence of PPIs use (69.6%) than that of controls (47.8%). Of the patients with gut-derived bacteraemia, only 44.6% had a documented indication for PPIs therapy. The antibacterial prophylaxis rate was 38.0% in the bacteraemia group and 58.7% in the non-antibacterial group. Based on multivariable logistic regression analysis, only PPIs use ($P = 0.00$, odds ratio (OR) = 0.546) was found to be associated with the risk of bacteraemia whereas antibacterial prophylaxis ($P = 0.00$, OR = 0.652) was protective. There were no significant differences in demographics, malignancy status, length of neutropenia, complications, or steroid use between the gut-derived bacteraemia and control group.

Conclusions: This study suggests a potential association between PPIs use and development of gut-derived bacteraemia in haematological malignancy patients after chemotherapy.

Keywords: Hematologic malignancy, Gut-derived bacteraemia, Proton pump inhibitor (PPI), Prophylactic antibiotics

Introduction

Bacteraemia is a major complication after chemotherapy in patients with haematological malignancy.¹ The prevalence ranges from 12 to 16.4% after chemotherapy in haematological malignancy patients,^{2,3} and 56–65% of such cases in developing countries suffer from gut-derived bacteraemia, which carries a very poor prognosis; indeed, the short-term mortality rate is ~25% even when the diagnosis is made early and treatment started rapidly.^{4–7} Gut-derived bacteraemia usually originates from the intestine.

Gut-derived bacteraemia is resulted from bacterial translocation across the intestinal wall to mesenteric

lymph nodes and then into the bloodstream. Bacteraemia may occur due to alterations of the host immune system, disruption of the normal indigenous bacterial flora, loss of the mucosal barrier, and among other causes.

Proton pump inhibitors (PPIs) are frequently administered to haematological patients during and after chemotherapy for diverse reasons. PPIs are commonly prescribed for dyspepsia and acid peptic disease, for routine stress ulcer prophylaxis, for prevention of non-steroidal anti-inflammatory drug-induced ulcers, etc. PPIs were also found to have the function to lower tolerance to chemotherapy,⁸ and to reduce tumour burden and metastasis.⁹ So that, PPIs are commonly used in haematological department. PPIs use may have potential harmful effects and, in particular,

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carries a risk of bacterial infection.^{10–12} PPIs are designed to shut down the gastric proton pump ($H^+/K^+-ATPase$) of parietal cells, thereby increasing the pH of the stomach and facilitating proliferation of intestinal bacteria.^{11,13,14} PPIs may also impair gastrointestinal motility,¹⁵ predisposing to bacterial translocation.¹⁶ In addition, impairment of neutrophil function by PPIs, which increases the risk of bacterial infection, has been reported.^{17–19} Currently, whether PPIs increase the risk of gut-derived bacteraemia remains unclear.

In this study, we compared the prevalence of PPIs use in patients with haematological malignancy with versus without gut-derived bacteraemia. We sought to investigate the role of PPIs in the occurrence of gut-derived bacteraemia and to determine whether PPIs treatment is appropriate in haematological malignancy patients during and after chemotherapy.

Patients and methods

Patients

In our retrospective case-control study, we identified the medical records of all patients with a confirmed diagnosis of bacteraemia admitted to the Haematology Department of a large urban tertiary care facility between January 2009 and July 2015. This research has been approved by our Institutional Review Board, and patient records was anonymized and de-identified prior to analysis. We included patients who suffered from bacteraemia and excluded those who suffered from lung and upper respiratory airway infection, urinary tract infection, perianal infection, catheter-related infection, skin infection, oral mucositis, and surgery patients, and the patients who recovered from bacteraemia recently. We also excluded patients who underwent bronchoscopy, gastroscopy, parenteral nutrition, and bladder catheter before bacteraemia within the preceding 7 days and those patients with gastrointestinal bleeding within the preceding 14 days.

Microbiological evaluations, which included two separate blood samples obtained from two different anatomical sites for culture, were performed at the onset of fever, according to standard practice. In the absence of an indwelling central venous catheter, two blood samples were obtained from two distinct peripheral veins. When an indwelling central venous catheter was present, one blood sample was obtained through this catheter, and a second was obtained from a peripheral vein.

The diagnosis and source of bloodstream infection were determined by the physician in charge and a specifically trained nurse and were based on objective clinical evidence, microbiological data, and clinical judgment. Blood cultures were performed in aerobic and anaerobic bottles and incubated in an automatic

system. Bacteraemia was identified using standard microbiological methods. Gut-derived bacteraemia cases (group 1) were defined as patients who met the inclusion and exclusion criteria. The non-bacteraemia patients (group 2) were haematology disease patients who underwent chemotherapy but were negative for bacteraemia (negative blood culture). All patients were admitted during the study period. Age, gender, underlying malignancy, malignancy status were compared between the two groups as well.

PPIs use and its therapeutic indications were assessed in the two groups by reviewing medical charts. Patients who received PPIs for at least 1 week prior to bacteraemia in group 1 and for at least 1 week during and after chemotherapy in group 2 were considered as being treated with PPIs, regardless of the dose. Other patients were considered non-users. A single dose of PPIs did not constitute PPIs use. Therapeutic use was defined as appropriate if it corresponded to an indication strongly supported by the medical literature.^{20–22} Acceptable indications for PPIs use were limited to gastro-oesophageal reflux disease, peptic ulcer disease and *Helicobacter pylori* eradication, Barrett's oesophagus, Zollinger–Ellison syndrome, and prevention of non-steroidal anti-inflammatory drug-induced ulcers and pathologic hypersecretory conditions. Data were collected using an anonymous standardized case-record form. Patient data consisted of demographic characteristics (age, gender), underlying malignancy, malignancy status, white blood cell count at the time of bacteraemia diagnosis, duration of neutropenia, antibacterial prophylaxis (we prescribed fluoroquinolone for the patients as antibacterial prophylaxis following the 2008 NCCN guideline for prevention and treatment of cancer-related infections) and corticosteroid therapy, and complications.

Statistical analysis

Data are expressed as percentages or medians (95% confidence intervals (CIs)). Statistical analysis was conducted using the chi-squared test, independent-samples *t*-test, and logistic regression. All statistical testing was two-tailed at the 5% level. The role of PPIs use and other factors potentially associated with bacteraemia was assessed by comparing patients in groups 1 and 2. A multivariable logistic regression analysis was performed to assess the association of PPIs use with the occurrence of bacteraemia while adjusting for other factors. All statistical analyses were performed using the SPSS 13.0 software.

Results

The medical charts of 493 haematological malignancy patients after chemotherapy with bacteraemia were reviewed. Confirmed gut-derived bacteraemia was

found in 92 cases. The non-bacteraemia patients were matched for age, gender, underlying malignancy, and the status of the underlying malignancy and were admitted to our hospital within 6 months of the study admission date.

The clinical characteristics of patients in the two groups are shown in Table 1. There was no significant difference in baseline parameters between the two groups.

There were no differences in the median duration of neutropenia between the groups. During their hospitalization, 84.3% of patients ($n = 184$) had neutropenia for a median duration of 7.7 days. The in-hospital mortality rate in group 1 was 6.5% ($n = 92$).

The pathogens isolated from blood cultures are shown in Table 2. Gram-negative bacilli were the predominant isolates, accounting for 82.6% of the total. The most common enteric organisms were *Escherichia coli* (38.0%), *Klebsiella pneumoniae* (26.1%), and *Enterobacter cloacae* (7.6%). Nine patients (9.8%) suffered from bacteraemia caused by Gram-positive organisms, of which *Streptococcus viridans* was the predominant isolate (5.4%). Seven (7.6%) of the 92 patients had polymicrobial infections (two to three pathogens). All six hospital deaths in the group were attributed to bacteraemia, of which four were Gram-negative, two were polymicrobial bacteraemia.

The proportion of haematological malignancy patients with gut-derived bacteraemia taking PPIs was 69.6%, compared with only 47.8% of those without bacteraemia; this difference was significant

Table 1 Characteristics of patients with and without gut-derived bacteraemia

Characteristic	Group 1 (bacteraemia) ($n = 92$)	Group 2 (non- bacteraemia) ($n = 92$)	<i>P</i>
Demographics			
Age, years*	43.2 ± 15.6	43.5 ± 15.0	0.878
Male, n (%)	45 (48.9)	48 (52.1)	0.768
Underlying malignancy			
ALL, n (%)	20 (21.7)	18 (19.6)	0.856
AML, n (%)	55 (59.8)	57 (62.0)	0.880
NHL, n (%)	11 (12.0)	11 (12.0)	1.00
MDS, n (%)	4 (4.3)	4 (4.3)	1.00
MM, n (%)	2 (2.2)	2 (2.2)	1.00
Status of malignancy	42 (45.7)	42 (45.7)	1.00
CR, n (%)			
Length of neutropenia, days*	7.8 ± 3.9	7.6 ± 6.6	0.807
Diabetes mellitus, n (%)	4 (4.3)	4 (4.3)	1.00
Hepatitis B, n (%)	10 (10.9)	7 (7.61)	0.612

*Data are expressed as medians (95% CIs). ALL, acute lymphoblastic leukaemia; AML, acute myeloid leukaemia; NHL, non-Hodgkin's lymphoma; MDS, myelodysplastic syndrome; MM, multiple myeloma; CR, complete remission.

Table 2 Pathogens and mortality rates

Causative pathogen	Rate ($n = 92$) (%)	Mortality rate ($n = 92$) (%)
Gram-negative	76 (82.6)	
<i>E. coli</i>	35 (38.0)	3 (3.3)
<i>K. pneumoniae</i>	24 (26.1)	1 (1.1)
<i>E. cloacae</i>	7 (7.6)	
<i>Pseudomonas aeruginosa</i>	5 (5.4)	
<i>Proteus mirabilis</i>	2 (2.2)	
<i>Enterobacter agglomerans</i>	1 (1.1)	
<i>Enterobacter aerogenes</i>	1 (1.1)	
<i>Chryseobacterium indologenes</i>	1 (1.1)	
Gram-positive	9 (9.8)	
<i>S. viridans</i>	5 (5.4)	
<i>Leuconostoc pseudomesenteroides</i>	4 (4.3)	
Polymicrobial bacteraemia	7 (7.6)	2 (2.2)

($P = 0.004$). Of the patients with gut-derived bacteraemia, only 44.6% had a documented indication for PPIs therapy. No significant differences were observed between bacteraemia and non-bacteraemia patients with regard to indications for PPIs prescription, the routes of administration of PPIs (i.v. vs. p.o.), or dosage of PPI (q.d. vs. b.i.d.) (Table 3).

Antibacterial prophylaxis was administered to 89 (48.9%) patients. The antibacterial prophylaxis rate was 38.0% in group 1 and 58.7% in group 2 (Table 4) ($P = 0.008$). The use of prophylactic antibiotics had a trend towards influencing the type of pathogen causing bacteraemia (Table 5), albeit not significantly so. Bacteraemia caused by Gram-negative organisms was associated with a 4.3% mortality rate, while no deaths due to bacteraemia caused by Gram-positive organisms were observed (Table 2).

A logistic regression analysis was performed with bacteraemia as the outcome (Table 6), in which only parameters significantly different in the multivariable analysis, i.e. PPIs use and antibacterial prophylaxis, were included. There were no statistically significant differences in demographics, status of malignancy, length of neutropenia, complications, and steroid use between the groups. PPIs use ($P = 0.00$, odds ratio (OR) = 0.546) were associated with bacteraemia, and

Table 3 Details of PPI use

	Group 1 (bacteraemia) ($n = 64$)	Group 2 (non- bacteraemia) ($n = 44$)	<i>P</i>
i.v. administration (%)	23 (35.9)	19 (43.2)	0.430
q.d. PPI dosage (%)	51 (79.7)	33 (75)	0.817
Appropriate PPI therapy (%)	29 (45.3)	22 (50)	0.696

q.d., every day.

Table 4 Acid-suppressive therapy and other medications

Factor	Group 1 (bacteraemia) (n = 92) (%)	Group 2 (non- bacteraemia) (n = 92) (%)	P
PPIs use, n (%)	64 (69.6)	44 (47.8)	0.004
Antibacterial prophylaxis, n (%)	35 (38.0)	54 (58.7)	0.008
Steroid use, n (%)	24 (26.1)	22 (23.9)	0.865

Table 5 Prophylaxis and the causative pathogen of bacteraemia

Causative pathogen	Prophylaxis (n = 35)	Non- prophylaxis (n = 57)	P
Gram-negative, n (%)	25 (71.4)	51 (89.5)	0.177
Gram-positive, n (%)	5 (14.3)	4 (7.0)	
Complex bacteraemia, n (%)	5 (14.3)	2 (3.5)	

antibacterial prophylaxis ($P = 0.00$, OR = 0.652) were negative correlation with bacteraemia.

Discussion

In recent years, PPIs have been associated with several adverse events, including pneumonia,²³ *Clostridium difficile* infection,²⁴ hip fracture,²⁵ interstitial nephritis,²⁶ low serum magnesium and vitamin B12 levels,²⁷ decreased iron absorption,²⁷ and increased cardiac birth defects during pregnancy.²⁸ Newer evidence now associates PPIs with spontaneous bacterial peritonitis^{29,30} and *C. difficile* infection in patients with cirrhosis.¹⁰ Our data support an association between PPIs and the development of gut-derived bacteraemia.

Use of PPIs is increasing in China despite the fact that the approved indications for PPIs use are selective and usually for a limited duration. Studies have shown the occurrence of inappropriate use of acid-suppressive

therapy in hospitalized patients.^{31,32} Because of their perceived safety, PPIs have come to be prescribed for a variety of non-approved indications, often for indefinite periods of time.³³ Up to 71% of patients in general medicine wards receive some sort of acid-suppressive therapy without an appropriate indication.³⁴ There is limited evidence to support the use of chronic acid suppression in patients during and after chemotherapy. There is no evidence suggesting that PPIs have a role in preventing or treating chemotherapy-induced nausea and vomiting. In our study, reasons for PPI therapy were either inappropriate or else undocumented in half of the cases. This suggests the need for strict adherence to the guidelines concerning PPI use in haematological malignancy patients during and after chemotherapy.

The mechanism by which gastric acid suppression might lead to an increased incidence of bacteraemia is poorly understood. The gastrointestinal tract is a primary reservoir for opportunistic bacteria. Study had found bacterial overgrowth in hypochlorhydric subjects,³⁵ the suppression of gastric acid secretion results in gastric bacterial overgrowth³⁶ and duodenal bacterial overgrowth.^{37,38} Gastric acid suppression can also reduce the primary barrier to bacteria within the gastrointestinal tract, and when combined with the intestinal permeability impairment seen in haematological malignancy after chemotherapy, it sets the stage for higher bacterial counts within the small intestine.^{15,18,19} The risk of enteric infections has been reported to be 2.5-fold greater in patients on acid-suppressive therapy compared with patients not receiving acid-suppressive therapy.³⁹ Bacteria within the gut can cross the gastrointestinal mucosal barrier and spread systemically by a process termed bacterial translocation. Factors that can trigger bacterial translocation from the gut include host immune deficiencies and immunosuppression, disturbances in the normal gut ecological balance, and mucosal barrier permeability. So the patients who received PPIs in our study were easily suffered from bacteraemia.

Therapies directed at preventing or limiting bacterial translocation are based on maintaining the host's physiologic defences, which include maintaining a stable gut flora, normal intestinal barrier function, and limiting chemotherapy-induced gut injury. Therefore, the inappropriate use of acid-suppressive therapy should be reduced or stopped. The major clinical therapeutic approach directed at limiting intestinal bacterial overgrowth with potential pathogenic bacteria is selective digestive tract decontamination.⁴⁰ In haematological malignancy patients after chemotherapy who undergo bone marrow suppression, the use of oral non-absorbable antibiotics plus a brief course of systemic antibiotics prevents intestinal bacterial overgrowth and thereby limits gut-derived bacteraemia.

Table 6 PPIs use and gut-derived bacteraemia: multivariable logistic regression analysis

Factor	Multivariable P-value
Age	0.394
Male	0.269
Status of malignancy (CR)	0.149
Length of neutropenia	0.806
Diabetes mellitus	0.397
Hepatitis B	0.234
PPIs use	0.000
Antibacterial prophylaxis	0.000
Steroid use	0.334

As found in this study, prophylactic antibiotics reduce the rate of gut-derived bacteraemia. The antibacterial prophylaxis rate was 38.0% in the gut-derived bacteraemia group, compared with 58.7% in the non-gut-derived bacteraemia group; this difference was significant. The use of prophylactic antibiotics tended to affect the type of pathogen causing bacteraemia, with only 71.4% caused by Gram-negative organisms compared with 14.3% by Gram-positive organisms. If no prophylactic antibiotics were given, the rates were 89.5 and 7.0%, respectively, though it was of no significant difference, but if we include more patients in the bacteraemia group, it might have resulted in detection of a significant difference. Feld⁴¹ found that among patients who received prophylactic antibiotics, Gram-positive bacteraemia was far more common than Gram-negative bacteraemia (75 vs. 25%), compared with ~50% each in patients who did not undergo prophylactic antibiotic treatment.

Our study had several limitations. Firstly, our results suggested an association between bacteraemia and PPIs use. However, an association does not prove causation. We could not confirm that the organisms isolated actually originated from the gut. Only a single objective inclusion criterion was used; i.e. a positive blood culture, although several exclusion criteria were applied. However, we did not culture gastric juice simultaneously with blood. Therefore, it was possible that the organisms originated from a site other than the gut. Moreover, determining the origin of organisms in the neutropenia patients was more difficult. Other limitations of the study included its retrospective design, the limited number of patients, and the fact that patients were not matched for certain factors, such as the intensity of chemotherapy. Although the strong association of PPIs use with bacteraemia suggests a causal relationship, other explanations are possible. PPIs may have been prescribed for dyspeptic symptoms associated with impaired small bowel motility, and such patients may have pre-existing bacterial overgrowth, which can predispose to bacteraemia. Symptoms of abdominal pain resulting from abdominal wall tension may have been treated with PPIs in some cases, and patients with these symptoms may have a greater predisposition to bacteraemia.

Future investigations into the association between acid suppression and bacteraemia appear warranted. A prospective, randomized, placebo-controlled trial would provide definitive evidence that reduction of PPIs use in haematological malignancy patients during and after chemotherapy is associated with a decreased risk of developing bacteraemia.

Conclusion

In conclusion, this study suggests a potential association between PPIs use and development of

bacteraemia in haematological malignancy patients after chemotherapy. Reasons for PPIs use were inappropriate or undocumented in half of the patients. Caution is advised when considering the use of PPIs in haematological malignancy patients who have undergone chemotherapy. The use of prophylactic antibiotics would reduce the rate of gut-derived bacteraemia.

Disclaimer statements

Contributors None.

Funding None.

Conflicts of interest There is no conflict of interest to report regarding this article.

Ethics approval This research has been approved by our Institutional Review Board.

References

- Cherif H, Kronvall G, Björkholm M, Kalin M. Bacteraemia in hospitalised patients with malignant blood disorders: a retrospective study of causative agents and their resistance profiles during a 14-year period without antibacterial prophylaxis. *Hematol J*. 2003;4(6):420–6.
- Engelhart S, Glasmacher A, Exner M, Kramer MH. Surveillance for nosocomial infections and fever of unknown origin among adult hematology–oncology patients. *Infect Control Hosp Epidemiol*. 2002;23(5):244–8.
- Huoi C, Vanhems P, Nicolle MC, Michallet M, Bénet T. Incidence of hospital-acquired pneumonia, bacteraemia and urinary tract infections in patients with haematological malignancies, 2004–2010: a surveillance-based study. *PLoS One*. 2013;8(3):e58121. doi:10.1371/journal.pone.0058121.
- Samonis G, Vardakas KZ, Maraki S, Tansarli GS, Dimopoulou D, Kofteridis DP, et al. A prospective study of characteristics and outcomes of bacteremia in patients with solid organ or hematologic malignancies. *Support Care Cancer*. 2013;21(9):2521–6.
- Velasco E, Soares M, Byington R, Martins CA, Schirmer M, Dias LM, et al. Prospective evaluation of the epidemiology, microbiology, and outcome of bloodstream infections in adult surgical cancer patients. *Eur J Clin Microbiol Infect Dis*. 2004;23(8):596–602.
- Metan G, Demiraslan H, Kaynar LG, Zararsiz G, Alp E, Eser B. Factors influencing the early mortality in haematological malignancy patients with nosocomial Gram negative bacilli bacteraemia: a retrospective analysis of 154 cases. *Braz J Infect Dis*. 2013;17(2):143–9.
- Chen CY, Tsay W, Tang JL, Tien HF, Chen YC, Chang SC, et al. Epidemiology of bloodstream infections in patients with haematological malignancies with and without neutropenia. *Epidemiol Infect*. 2010;138(7):1044–51.
- Ferrari S, Perut F, Fagioli F, Brach Del Prever A, Meazza C, Parafioriti A, et al. Proton pump inhibitor chemosensitization in human osteosarcoma: from the bench to the patients' bed. *J Transl Med*. 2013;11:268. doi:10.1186/1479-5876-11-268.
- Spugnini EP, Citro G, Fais S. Proton pump inhibitors as anti vacuolar-ATPases drugs: a novel anticancer strategy. *J Exp Clin Cancer Res*. 2010;29:44. doi:10.1186/1756-9966-29-44.
- Barletta JF, El-Ibiary SY, Davis LE, Nguyen B, Raney CR. Proton pump inhibitors and the risk for hospital-acquired *Clostridium difficile* infection. *Mayo Clin Proc*. 2013;88(10):1085–90.
- Bauer TM, Steinbrückner B, Brinkmann FE, Ditzel AK, Schwacha H, Aponte JJ, et al. Small intestinal bacterial overgrowth in patients with cirrhosis: prevalence and relation with spontaneous bacterial peritonitis. *Am J Gastroenterol*. 2001;96(10):2962–7.
- Siple JF, Morey JM, Gutman TE, Weinberg KL, Collins PD. Proton pump inhibitor use and association with spontaneous bacterial peritonitis in patients with cirrhosis and ascites. *Ann Pharmacother*. 2012;46(10):1413–8.
- Thorens J, Froehlich F, Schwizer W, Saraga E, Bille J, Gyr K, et al. Bacterial overgrowth during treatment with omeprazole

- compared with cimetidine: a prospective randomised double blind study. *Gut*. 1996;39(1):54–9.
- 14 van Vlerken LG, Huisman EJ, van Hoek B, Renooij W, de Rooij FW, Siersema PD, *et al*. Bacterial infections in cirrhosis: role of proton pump inhibitors and intestinal permeability. *Eur J Clin Invest*. 2012;42(7):760–7.
 - 15 Parkman HP, Urbain JL, Knight LC, Brown KL, Trate DM, Miller MA, *et al*. Effect of gastric acid suppressants on human gastric motility. *Gut*. 1998;42(2):243–50.
 - 16 Lewis SJ, Franco S, Young G, O’Keefe SJ. Altered bowel function and duodenal bacterial overgrowth in patients treated with omeprazole. *Aliment Pharmacol Ther*. 1996;10(4):557–61.
 - 17 Agastya G, West BC, Callahan JM. Omeprazole inhibits phagocytosis and acidification of phagolysosomes of normal human neutrophils *in vitro*. *Immunopharmacol Immunotoxicol*. 2000;22(2):357–72.
 - 18 Yoshida N, Yoshikawa T, Tanaka Y, Fujita N, Kassai K, Naito Y, *et al*. A new mechanism for anti-inflammatory actions of proton pump inhibitors – inhibitory effects on neutrophil-endothelial cell interactions. *Aliment Pharmacol Ther*. 2000;14(Suppl 1):74–81.
 - 19 Zedtwitz-Liebenstein K, Wenisch C, Patruta S, Parschalk B, Daxböck F, Graninger W. Omeprazole treatment diminishes intra- and extracellular neutrophil reactive oxygen production and bactericidal activity. *Crit Care Med*. 2002;30(5):1118–22.
 - 20 American Gastroenterological Association, Spechler SJ, Sharma P, Souza RF, Inadomi JM, Shaheen NJ. American Gastroenterological Association medical position statement on the management of Barrett’s esophagus. *Gastroenterology*. 2011;140(3):1084–91.
 - 21 Laine L, Jensen DM. Management of patients with ulcer bleeding. *Am J Gastroenterol*. 2012;107(3):345–60; quiz 61.
 - 22 Malfertheiner P, Chan FK, McColl KE. Peptic ulcer disease. *Lancet*. 2009;374(9699):1449–61.
 - 23 Herzig SJ, Doughty C, Lahoti S, Marchina S, Sanan N, Feng W, *et al*. Acid-suppressive medication use in acute stroke and hospital-acquired pneumonia. *Ann Neurol*. 2014;76(5):712–8.
 - 24 Proton pump inhibitors: *Clostridium difficile* infections. *Prescrire Int*. 2013;22(142):239–40.
 - 25 Adams AL, Black MH, Zhang JL, Shi JM, Jacobsen SJ. Proton-pump inhibitor use and hip fractures in men: a population-based case-control study. *Ann Epidemiol*. 2014;24(4):286–90.
 - 26 Blank ML, Parkin L, Paul C, Herbison P. A nationwide nested case-control study indicates an increased risk of acute interstitial nephritis with proton pump inhibitor use. *Kidney Int*. 2014;86(4):837–44.
 - 27 Mathieu N. Risk of long-term treatment with proton pump inhibitors. *Rev Prat*. 2008;58(13):1451–4.
 - 28 Talley NJ. Risks of proton-pump inhibitors: what every doctor should know. *Med J Aust*. 2009;190(3):109–10.
 - 29 Bajaj JS, Ratliff SM, Heuman DM, Lapane KL. Proton pump inhibitors are associated with a high rate of serious infections in veterans with decompensated cirrhosis. *Aliment Pharmacol Ther*. 2012;36(9):866–74.
 - 30 Goel GA, Deshpande A, Lopez R, Hall GS, van Duin D, Carey WD. Increased rate of spontaneous bacterial peritonitis among cirrhotic patients receiving pharmacologic acid suppression. *Clin Gastroenterol Hepatol*. 2012;10(4):422–7.
 - 31 Gupta R, Garg P, Kottoor R, Munoz JC, Jamal MM, Lambiase LR, *et al*. Overuse of acid suppression therapy in hospitalized patients. *South Med J*. 2010;103(3):207–11.
 - 32 Hoffmann F, Glaeske G, Schmiemann G. Increased prescribing of proton pump inhibitors in ambulatory care over the years 2005–2013. *Z Gastroenterol*. 2015;53(2):95–100.
 - 33 Naunton M, Peterson GM, Bleasel MD. Overuse of proton pump inhibitors. *J Clin Pharm Ther*. 2000;25(5):333–40.
 - 34 Grube RR, May DB. Stress ulcer prophylaxis in hospitalized patients not in intensive care units. *Am J Health Syst Pharm*. 2007;64(13):1396–400.
 - 35 Saltzman JR, Kowdley KV, Pedrosa MC, Sepe T, Golner B, Perrone G, *et al*. Bacterial overgrowth without clinical malabsorption in elderly hypochlorhydric subjects. *Gastroenterology*. 1994;106(3):615–23.
 - 36 Theisen J, Nehra D, Citron D, Johansson J, Hagen JA, Crookes PF, *et al*. Suppression of gastric acid secretion in patients with gastroesophageal reflux disease results in gastric bacterial overgrowth and deconjugation of bile acids. *J Gastrointest Surg*. 2000;4(1):50–4.
 - 37 Pereira SP, Gainsborough N, Dowling RH. Drug-induced hypochlorhydria causes high duodenal bacterial counts in the elderly. *Aliment Pharmacol Ther*. 1998;12(1):99–104.
 - 38 Fried M, Siegrist H, Frei R, Froehlich F, Duroux P, Thorens J, *et al*. Duodenal bacterial overgrowth during treatment in outpatients with omeprazole. *Gut*. 1994;35(1):23–6.
 - 39 Leonard J, Marshall JK, Moayyedi P. Systematic review of the risk of enteric infection in patients taking acid suppression. *Am J Gastroenterol*. 2007;102(9):2047–56; quiz 57.
 - 40 Gatt M, Reddy BS, MacFie J. Review article: bacterial translocation in the critically ill – evidence and methods of prevention. *Aliment Pharmacol Ther*. 2007;25(7):741–57.
 - 41 Feld R. Bloodstream infections in cancer patients with febrile neutropenia. *Int J Antimicrob Agents*. 2008;32(Suppl 1):S30–3.