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RESEARCH ARTICLE

Moringa oleifera leaves as an inhibitor of human platelet aggregation

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Abstract

Moringa oleifera L. (Moringaceae) leaves were examined for their effect on human platelet aggregation *in vitro*. The aqueous extract of *Moringa oleifera* leaves significantly ($p \leq 0.05$) inhibited platelet aggregation induced by agonists such as adenosine diphosphate, collagen, and epinephrine. The degree of inhibitory activity varied depending on the agonist used, concentration of extract and duration of incubating the extract with platelets. Heat treatment reduced the inhibitory activity of extract against platelet aggregation. In addition, the extracts significantly ($p \leq 0.05$) decreased the amount of malonaldehyde formed in agonist challenged platelets. This study is the first report on the effect of aqueous extract of *Moringa oleifera* leaves against human platelet aggregation. Overall, *Moringa oleifera* leaves have potential to protect platelets against aggregation.

Keywords: *Moringa oleifera*; inhibitory activity; platelet aggregation

Introduction

Platelet aggregation is a physiological process proved to have defined roles in cardiovascular diseases. Normally, in response to a vascular injury, platelets rapidly undergo the processes of adhesion, shape change, secretion, and aggregation through a series of exquisitely coordinated responses (Kroll & Schafer, 1989) that result in the formation of homeostatic plugs or arterial thrombi at the sites of vessel injury to stop blood loss. These thrombi are the source of thromboembolic complications of atherosclerosis, heart attacks, stroke, and peripheral vascular disease (Son et al., 2004). Therefore, compounds that counter aggregation have a protective role regarding thromboembolic disorders (Subramoniam & Ana, 1988).

It is realized that prevention of thrombotic events by dietary means rather than commonly used antithrombotic drugs (e.g., tolmetin, aspirin, heparin, warfarin)

would be more effective and economic in large populations. This has led to interest in exploring the anti-platelet effects of various dietary components including fruits, vegetables, spices, and herbs, and their active constituents (Srivastava, 1989). The anti-platelet effects of several herbs and spices including garlic, onion, ginger, turmeric, cloves, and their active principles have been established (Rattan, 1988).

Moringa oleifera L. (Moringaceae), or drumstick tree, is a rapidly growing perennial soft wood tree that is native to the Indo-Bangla subcontinent and cultivated throughout the tropical belt (Sastri, 1962; Fahey, 2005). All parts of the tree are edible and have long been consumed by humans (Fahey, 2005). The leaves are tri-pinnate, feathery, with many small leaflets, and are rich in β -carotene, vitamin C, vitamin B, calcium, iron, and essential amino acids. Thus, they have been advocated as an outstanding indigenous source of nutrients suitable for utilization in many of the so-called “developing”

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regions of the world to combat malnutrition (Freiberger et al., 1998; Nambiar & Seshadri, 2001; Fahey, 2005). In the Indian indigenous system of medicine, *M. oleifera* leaves are used for the treatment of a variety of common ailments e.g., anemia, anxiety, dry cough, bronchitis, joint pain, scurvy, and psoriasis (Sastri, 1962; Mathur, 2005). A wide range of pharmacological properties such as anti-inflammatory, anti-tumor, antimicrobial, hypotensive, hypocholesterolemic, and hypoglycemic have been reported for *M. oleifera* leaves in the scientific literature (Fahey, 2005). This study is the first report on the effect of aqueous extract of *M. oleifera* leaves against human platelet aggregation.

Materials and methods

Chemicals

Adenosine 5'-diphosphate (ADP), collagen, and epinephrine were purchased from Sigma Chemicals (St. Louis, MO). Malonaldehyde-bis-dimethyl acetal was obtained from Merck-Schuchardt (Germany) and thiobarbituric acid (TBA) from HiMedia Lab (Bombay, India).

Materials

Moringa oleifera leaves were obtained in bulk from a local market in Mysore, India. The taxonomic identity of the plant was confirmed by comparison of the plant material with those of the known identity in the Herbarium specimen identified by Dr. G. R. Janardhana, Department of Studies in Botany, University of Mysore, Mysore, India.

Preparation of aqueous extracts of raw and heat-treated *M. oleifera* leaves

Fresh *M. oleifera* leaves (20 g) were cleaned, washed, and ground to a fine paste with addition of distilled water. The slurry obtained was left overnight at 4°C. Afterwards, the contents were centrifuged at 10,000 rpm for 15 min to obtain a clear extract and volume made up to 50 ml with distilled water. To prepare the extract of heat-treated *M. oleifera* leaves, fresh leaves were heated at 100°C for 15 min and the extract of heat-treated leaves was prepared as mentioned above. The freshly prepared extracts were used for platelet aggregation studies.

Isolation of platelets

Venous blood was collected in 3.8% tri-sodium citrate (9:1 v/v) from healthy volunteers who had not taken any medication for the past 10 days. This citrated blood was centrifuged at 1,100 rpm for 20 min to obtain platelet rich plasma (PRP), which was used within 3 h after

collection. The residual blood was again centrifuged at 2,500 rpm for 20 min to obtain the homologous platelet poor plasma (PPP). Platelet count was adjusted to 1.6×10^7 platelets per μL of PRP (Gerrad, 1982).

Platelet aggregation assay

The aggregation experiments were carried out turbidimetrically in a Dual Path Aggro-meter (Chronolog, Havertown, PA) (Suneetha & Krishnakantha, 2005a). PRP of 450 μL was kept stirred at 1,200 rpm in a glass cuvette at 37°C, while aggregation was induced by agonists such as adenosine diphosphate (ADP) (61 μM), collagen (0.005% in 0.1 N acetic acid), and epinephrine (76 μM). Similarly, PRP was pre-incubated with the extracts at different concentrations for 1 min followed by induction of aggregation by agonists mentioned above. The change in turbidity was recorded at least for 4 min in each case with reference to PPP using an Amersham Pharmacia Biotech Recorder 111, in the presence and absence of extracts. The slopes were calculated for control and experimental samples and percentage inhibition of aggregation due to control was calculated. The concentration of plant extract (mg) at which platelet aggregation was inhibited by 50% (IC_{50}) was obtained by interpolation from linear regression analysis.

Effect of incubation time on platelet aggregation

The effect of duration of incubation of extracts with platelets on platelet aggregation was determined at IC_{50} of each extract. PRP (450 μL) was pre-incubated with sample extract for different times (1, 3, 5, & 10 min) before induction of aggregation, and percentage inhibition of platelet aggregation was calculated.

Estimation of malonaldehyde (MA) in agonist challenged platelets

The amount of MA formed in agonist challenged platelets was determined according to the method of Maguire and Csonka-Khalifah (1987). After aggregation, platelets (450 μL) were taken and mixed with 20 μL of 1% butylated hydroxytoluene (BHT) in ethanol and 100 μL of 100% trichloroacetic acid in 3 N HCl. The pellet was centrifuged at 10,000 rpm for 10 min to obtain 450 μL of supernatant to which 100 μL of TBA reagent (0.12 M TBA in 0.26 M Tris-HCl solution) was added. The chromophor generated was measured at 532 nm, and MA equivalents were calculated using the following equation:

$$\text{MA equivalents (nmol/} 1.6 \times 10^7 \text{ platelets)} = (\text{Absorbance/156}) \times \text{total volume}$$

Statistical analysis

Data were recorded as means \pm standard deviation of triplicate measurements. Analyses of variance were performed by ANOVA test and significance differences between the means were determined by Duncan's multiple-range test ($p \leq 0.05$) (Steel & Torrie, 1980).

Results and discussion

Effect of *M. oleifera* extracts on human platelet aggregation

Aqueous extract of raw *M. oleifera* leaves inhibited platelet aggregation induced by the agonists, collagen, ADP, and epinephrine in a dose-dependent manner (Figure 1). In the concentration range of 0.2–1 mg, the extract inhibited aggregation by 25.93%–89.26% in the case of collagen, and 20.65%–65.39% in the case of ADP. This indicated that *M. oleifera* extract was a potent inhibitor of collagen-induced platelet aggregation. When epinephrine was used as the agonist, the extract showed a dose-dependent activity (12.54%–38.18%) in the concentration range of 0.2–0.8 mg. Increasing the concentration of extract to 1 mg appeared to suppress epinephrine-induced aggregation by almost 100%. The IC_{50} values of *M. oleifera* extract on collagen- and ADP-induced aggregation were 0.48 mg and 0.70 mg (Table 1), respectively, indicating that 50% inhibition of aggregation can be achieved by a significantly lower concentration of extract, when aggregation was induced by collagen.

In general, plant foods are subjected to processing such as boiling, pressure-cooking, steaming, roasting, frying, etc., before consumption. Since in India, *M. oleifera* leaves are commonly used in cooked form

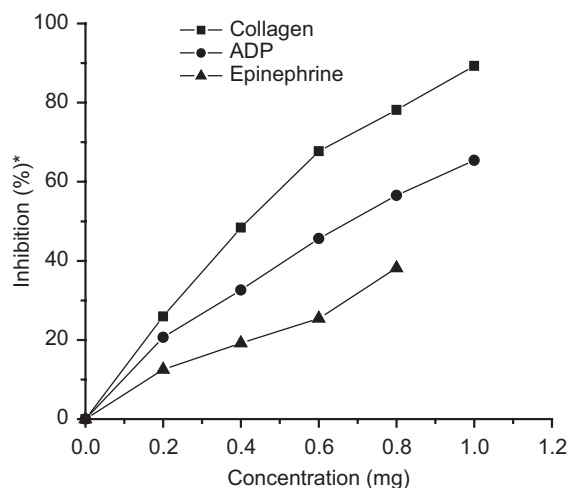


Figure 1. Effect of aqueous extract of *Moringa oleifera* leaves on human platelet aggregation induced by collagen, ADP and epinephrine. *Percentage inhibition of aggregation relative to control.

(preparation of curry), it was reasonable to examine the effect of heat treatment on their inhibitory activity against platelet aggregation. The effect of extract obtained from heat-treated (100°C, 15 min) leaves on platelet aggregation is shown in Figure 2. In the concentration range of 0.2–1 mg, the inhibitory effect of extract against agonist-induced platelet aggregation was 17.64%–53.97% (in the case of collagen), and 13.25%–42.47% (in the case of ADP). The inhibitory effect of extract against epinephrine-induced aggregation reached its maximum (44.9%) with 0.6 mg of extract, beyond which a decline in the activity of the extract was observed. From these results it is evident that 50% inhibition of ADP- and epinephrine-induced aggregation could not be achieved with the concentration range used in this study, i.e., there was poor inhibitory activity after heat processing of the sample. The IC_{50} value of extract against collagen-induced aggregation was 0.87 mg, which was significantly ($p \leq 0.05$) higher than that of raw leaf extract (0.48 mg) (Table 1). This indicated that anti-aggregatory potential of *M. oleifera* leaves was reduced by heat processing, which could be attributed to the loss of active components responsible for activity of extract. However, the residual activity in the extract was about 54%, indicating that it can be still considered as a good inhibitor of platelet aggregation.

Table 1. IC_{50} (mg)* of aqueous extracts of *Moringa oleifera* leaves on agonist-induced platelet aggregation.

| Extract | Collagen | ADP | Epinephrine |
|--------------|-----------------|-----------------|-------------|
| Raw | 0.48 \pm 0.03 | 0.70 \pm 0.04 | ND |
| Heat-treated | 0.87 \pm 0.04 | ND | ND |

Values are mean \pm SD. Duration of incubation was 1 min.

*The concentration required to inhibit platelet aggregation by 50%. ND, not determined.

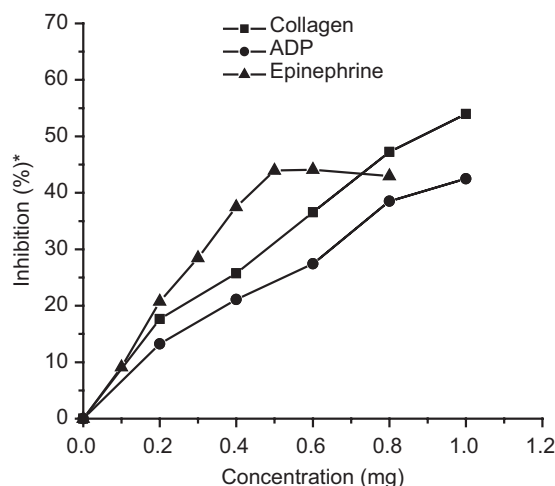


Figure 2. Effect of aqueous extract of heat-treated (100°C, 15 min) *Moringa oleifera* leaves on human platelet aggregation induced by collagen, ADP, and epinephrine. *Percentage inhibition of aggregation relative to control.

It has been shown that about 86-91% of the active principle of spices was lost in turmeric, 13-17% in black pepper powder, and 4-19% in red pepper powder when boiled for 15 min (Srinivasan et al., 1992). Similarly, a two-fold decrease in the consistency range was observed when garlic and onion were processed at high temperature (Ahmed, 2000).

The aqueous extracts of *M. oleifera* leaves (raw and heat-treated) were able to inhibit platelet aggregation induced by collagen, ADP and epinephrine. Reports indicate that activation of platelets with agonists brings about changes in the cyclo-skeletal structure, resulting in the loss of discoidal shape and pseudopodal projections (Mukherjee et al., 1990). It has been shown that ADP and epinephrine induce aggregation by reducing the intra-platelet c-AMP levels and inhibiting adenylate cyclase activity (Hawiger et al., 1980). Collagen-induced platelet aggregation has been reported to be associated with a burst of hydrogen peroxide (Pignatelli et al., 1998), a pro-oxidant which stimulates the arachidonic acid (AA) metabolism by contributing to the platelet production of thromboxane A_2 (Pignatelli et al., 1999). Epinephrine-induced platelet aggregation is mediated through α_2 -adrenergic receptors that also decrease the PGE_1 stimulated c-AMP levels (Figures et al., 1986). The mode of action of these platelet agonists is to help increase the cytosolic levels of calcium either due to release from internal stores or through calcium reflux (Shah et al., 1999). A rise in the cytosolic levels of calcium accompanies platelet activation through stimulation of enzymes, which were otherwise not fully functional at low levels of calcium concentration present in resting platelets (Heemsherk & Sage, 1994). Calcium antagonists usually may not bind to the specific receptor sites but may cause the thickening of membranes due to the insertion of these antagonists into the membrane bilayers, thus affecting the calcium mobilization (Blache et al., 1987). The agonists, collagen, ADP, and epinephrine may also activate the surface glycoprotein receptors IIb-IIIa (GP IIb-IIIa) as well as interaction with the von Willebrand factor found in plasma (Scott et al., 1991; Bloekmans et al., 1995). Hence, the suppression of platelet aggregation indicated by *M. oleifera* extract in this study might be attributed to not only competition of extract components with the GP IIb - IIIa receptors along with the agonists, but also their interaction with membrane bilayers, causing the thickening of membranes and thus affecting calcium mobilization. It may also be likely that these extracts prevent reduction of intra-platelet c-AMP levels and adenylate cyclase stimulation caused by ADP and epinephrine. Inhibition of collagen-induced aggregation with the extract might be due to blocking the release of hydrogen peroxide, which will otherwise stimulate AA metabolism.

Effect of incubation time on inhibition of platelet aggregation

Human platelet aggregation is usually followed with the inhibitors being incubated for one min in the normal course. In this study, it was also of interest to determine the effect of longer exposure of platelets to inhibitors on the inhibition of platelet aggregation. The effect of incubation time on inhibition of platelet aggregation with the extracts (raw and heat-treated) is presented in Table 2. The maximum inhibition of collagen-induced platelet aggregation (69.84%) with the extract of raw sample was achieved due to incubation with PRP for 5 min. Further incubation (10 min) did not affect the activity of extract significantly ($p \leq 0.05$). The inhibition of ADP-induced platelet aggregation significantly ($p \leq 0.05$) increased with increase in duration of incubation. The extract incubated with platelets for 10 min, exhibited 83.78% anti-aggregatory activity on ADP. Epinephrine-induced platelet aggregation was highly affected by increasing the time of incubation. Maximum inhibition was seen at 3 min incubation for epinephrine-induced aggregation. When the extract of heat-treated leaves was incubated with PRP for 3 min, a 7% decline in the inhibition of platelet aggregation was noted.

The results showed that the efficacy of *M. oleifera* extract in inhibiting platelet aggregation was time-dependent. The increase in the inhibitory activity observed might be attributed to a better uptake or interactions of extract component(s) into the platelets or a better incorporation of active components into the platelet membrane bilayers during longer incubation. Prolonged incubation did not improve the anti-aggregatory activity of the extract of heat-treated *M. oleifera* leaves. The inhibitory activity of aqueous extracts of coriander, saffron and cardamom against platelet aggregation induced by ADP, collagen and epinephrine were found to be time-dependent at IC_{50} (Suneetha & Krishnakantha, 2005a, 2005b, 2005c).

Table 2. Effect of incubation time on inhibition of platelet aggregation with aqueous extracts of *Moringa oleifera* leaves at IC_{50} .

| | Inhibition (%)* | | | |
|--------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|
| | 1 min | 3 min | 5 min | 10 min |
| Raw | | | | |
| Collagen | 49.56 \pm 2.24 ^c | 58.73 \pm 1.80 ^b | 69.84 \pm 2.50 ^a | 70.63 \pm 1.50 ^a |
| ADP | 50.10 \pm 1.70 ^d | 67.72 \pm 0.98 ^c | 74.77 \pm 2.80 ^b | 83.78 \pm 2.35 ^a |
| Epinephrine [†] | 38.18 \pm 1.10 ^b | 100 ^a | ND | ND |
| Heat-treated | | | | |
| Collagen | 49.86 \pm 1.32 ^a | 42.35 \pm 1.60 ^b | ND | ND |

Values are mean \pm SD. Values bearing different superscripts a, b, c ... in the same row differ significantly ($p \leq 0.05$) from each other.

* Percentage inhibition of platelet aggregation relative to control.

[†] Due to 0.80 mg of extract.

ND, not determined

Table 3. Effect of aqueous extracts of *Moringa oleifera* leaves on malonaldehyde (MA) formed in agonist challenged platelets at IC₅₀.

| Sample | MA formed (nmol/1.6 × 10 ⁷ platelets) | | |
|-------------------------|--|-----------------------------|-----------------------------|
| | Collagen | ADP | Epinephrine |
| Control* | 556.25 ± 21.30 ^a | 501.41 ± 22.47 ^a | 391.73 ± 17.50 ^a |
| <i>Moringa oleifera</i> | | | |
| Raw | 365.06 ± 13.45 ^c | 352.55 ± 15.90 ^c | 274.21 ± 12.20 ^b |
| Heat-treated | 413.53 ± 10.35 ^b | ND | ND |

Values are mean ± SD. Values bearing different superscripts a, b, c... in the same column differ significantly ($p \leq 0.05$) from each other.

*Control represents platelets stimulated aggregation without addition of any inhibitor.

[†]Due to 0.80 mg of extract.

ND, not determined.

Estimation of malonaldehyde (MA) in agonist challenged platelets

Malonaldehyde is one of the end products of the cyclooxygenase pathway of AA metabolism (Rattan, 1988). It is believed that lipid peroxides (e.g., MA) along with thromboxane A₂ (TXA₂) increase the platelet sensitivity to agonists, thus leading to coronary heart diseases (Neiva et al., 1999). The amount of MA released in agonist-challenged platelets decreased significantly ($p \leq 0.05$) at IC₅₀ of *M. oleifera* extract with different agonists (Table 3). The reduction in MA released during collagen-induced aggregation with the extract of raw leaves was higher than that of heat-treated leaves. The extract of raw leaves exhibited 34.37%, 29.68%, and 30% inhibitory activity toward MA formation during collagen-, ADP-, and epinephrine-mediated aggregation, respectively. The extract of heat-treated leaves exhibited 25.65% inhibitory activity against formation of MA in collagen-challenged platelets. The results indicate that the extract possessed component(s), which were able to inhibit the formation of release products through degradation of AA. It may also be likely that the cyclooxygenase pathway of AA metabolism was being affected, thus resulting in a decrease in platelet aggregation. Inhibition of cyclooxygenase pathway can lead to accumulation of AA, which in turn converts into 12-HPETE, a potent anti-aggregatory agent, through lipooxygenase pathway of AA metabolism (Rattan, 1988). Thus, the inhibition of platelet aggregation might be attributed to combined effects of reduced formation of MA and TXA₂ and increased formation of 12-HPETE. It has been reported that ginger, omum, cloves, cumin, and turmeric increased the formation of platelet lipooxygenase products (Srivastava, 1984, 1988, 1989; Srivastava & Justesen, 1987). Onion has been shown to suppress both the cyclooxygenase and lipooxygenase pathways in platelets (Srivastava, 1986).

Conclusions

The results of this investigation revealed that *M. oleifera* leaves have components that are capable of

inhibiting human platelet aggregation *in vitro*. Their anti-aggregatory activity was dependent on the concentration of extract, duration of incubation with platelets, and type of inducer used. The extract appeared to suppress aggregation process mediated by agonists through the inhibition of cyclooxygenase pathway of AA metabolism. Further investigations are needed to isolate, identify and purify the active components, and to elucidate the exact biochemical interactions between the component(s) of the extracts and platelets, which may result in inhibition of platelet aggregation. Regular and increased consumption of *M. oleifera* leaves is recommended in order to exploit their nutritional and beneficial health effects such as their capacity as anti-platelet agents.

Declaration of interest: The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

References

- Ahmed J (2000): Effect of temperature on rheological characteristics of garlic and onion pastes. *J Food Sci Tech* 37: 409–411.
- Blache D, Ciavatti M, Ojeda C (1987): The effect of calcium channel blockers on blood platelet function especially calcium uptake. *Biochim Biophys Acta* 923: 401–412.
- Bloekmans D, Peckmyn H, Vermeylen J (1995): Platelet activation. *Blood Rev* 9: 143–156.
- Fahey JW (2005): *Moringa oleifera*: A review of the medical evidence for its nutritional, therapeutic, and prophylactic properties. Part 1. *Trees Life J* 1: 5–19.
- Figures WR, Searce LM, Wachtfogel Y, Chen J, Colman RF, Colman RW (1986): Platelet ADP receptor and α_2 -adrenoreceptor interaction. *J Biol Chem* 261: 5981–5986.
- Freiberger CE, Vanderjagt DJ, Pastuszyn A, Glew RS, Mounkaila G, Millson M, Glew RH (1998): Nutrient content of the edible leaves of seven wild plants from Niger. *Plant Foods Hum Nutr* 53: 57–69.
- Gerrad JM (1982): Platelet aggregation and the influence of prostaglandins, in: Lands WEM, Smith WL, eds. *Prostaglandins and Arachidonate Metabolites* (Methods in Enzymology series, Vol. 86). London, Academic Press, pp. 642–643.
- Hawiger J, Parkinson S, Timmons S (1980): Prostacyclin inhibits mobilization of fibrinogen binding sites on human ADP- and thrombin-treated platelets. *Nature* 283: 195–197.
- Heemsherk JWM, Sage O (1994): Calcium signaling in platelets and other cells. *Platelets* 5: 295–316.
- Kroll MH, Schafer AI (1989): Biochemical mechanisms of platelet activation. *Blood* 74: 1181–1195.
- Maguire MH, Csonka-Khalifah L (1987): Vinca alkaloids inhibit conversion of arachidonic acid to thromboxane by human platelet microsomes: Comparison with other microtubule-active drugs. *Biochim Biophys Acta* 921: 426–436.
- Mathur BS (2005): *Moringa oleifera*: A potential life-saver. Available at: http://www.treesforlife.org/project/moringa/book/moringa_book_view.pdf. (Accessed 2005).
- Mukherjee G, Chatterjee GC, Banerjee D, Bhattacharya DK (1990): Differential effect of retinoic acid on ADP and collagen induced platelet aggregation. *Indian J Exp Biol* 28: 49–952.
- Nambiar VS, Seshadri S (2001): Bioavailability trials of β -carotene from fresh and dehydrated drumstick leaves (*Moringa oleifera*) in a rat model. *Plant Foods Hum Nutr* 56: 83–95.
- Neiva TJC, Morais L, Polack M, Simoes MD, D'Amico EA (1999): Effect of catechins on human platelet aggregation and lipid peroxidation. *Phytother Res* 13: 597–600.

- Pignatelli P, Pulcinelli FM, Lenti L, Gazzania PP, Violi F (1998): Hydrogen peroxide is involved in collagen induced platelet activation. *Blood* 91: 484-490.
- Pignatelli P, Pulcinelli FM, Lenti L, Gazzania PP, Violi F (1999): Vitamin E inhibits collagen induced platelet activation by blunting hydrogen peroxide. *Atheroscler Thromb Vasc Biol* 19: 2542-2547.
- Rattan ISS (1988): Science behind spices: Inhibition of platelet aggregation and prostaglandin synthesis. *Bioassays* 5: 161-162.
- Sastri BN (1962): *The Wealth of India, Raw Materials*. New Delhi, India, Council of Scientific and Industrial Research, pp. 425-429.
- Scott JP, Montgomery RR, Retzimer GS (1991): Dimeric ristocetin flocculates proteins, binds to platelets and mediates von Willebrand factor-dependent agglutination of platelets. *J Biol Chem* 266: 8149-8155.
- Shah BH, Nawaz Z, Petrani SA, Roomi A, Mahmood H, Saeed SA, Gilani AH (1999): Inhibitory effect of curcumin, a food spice from turmeric, on platelet-activating factor- and arachidonic acid-mediated platelet aggregation through inhibition of thromboxan formation and Ca^{2+} signaling. *Biochem Pharmacol* 58: 1167-1172.
- Son DJ, Cho, MR, Jin YR, Kim Sy, Park YH, Lee SH, Akiba S, Sato, T, Yun YP (2004): Antiplatelet effect of green tea catechins: A possible mechanism through arachidonic acid pathway. *Prostaglandins Leukot Essent Fatty Acids* 71: 25-31.
- Srinivasan K, Sambiah K, Chandrasekhar N (1992): Loss of active principles of common spices during domestic cooking. *Food Chem* 43: 271-274.
- Srivastava KC (1984): Effects of aqueous extracts of onion, garlic and ginger on platelet aggregation and metabolism of arachidonic acid in the blood vascular system. *Prostaglandins Leukot Med* 13: 227-235.
- Srivastava KC (1986): Onion exerts antiaggregatory effects by altering arachidonic acid metabolism in platelets. *Prostaglandins Leukot Med* 24: 43-50.
- Srivastava KC (1988): Extract of a spice omum (*Trachyspermum ammi*) shows antiaggregatory effects and alters arachidonic acid metabolism in human platelets. *Prostaglandins Leukot Essent Fatty Acids* 33: 1-6.
- Srivastava KC (1989): Extracts from two frequently consumed spices - cumin (*Cuminum cyminum*) and turmeric (*Cucuma longa*) - inhibit platelet aggregation and alter eicosanoid biosynthesis in human blood platelets. *Prostaglandins Leukot Essent Fatty Acids* 37: 57-64.
- Srivastava KC, Justesen U (1987): Inhibition of platelet aggregation and reduced formation of thromboxane and lipoxygenase products in platelets with oil of cloves. *Prostaglandins Leukot Med* 29: 11-18.
- Steel RGD, Torrie JH (1980): *Principles and Procedures of Statistics: A Biometrical Approach*, 2nd edn. New York, NY, McGraw-Hill, p. 187.
- Subramoniam A, Ana MNS (1988): Influence of certain dietary plant constituents on platelet aggregation. *J Food Safety* 9: 201-214.
- Suneetha WJ, Krishnakantha TP (2005a): Inhibition of human platelet aggregation and membrane lipid peroxidation by food spice, saffron. *Mol Cell Biochem* 278: 59-63.
- Suneetha WJ, Krishnakantha TP (2005b): Antiplatelet activity of coriander and curry leaf spices. *Pharm Biol* 43: 230-233.
- Suneetha WJ, Krishnakantha TP (2005c): Cardamom extract as inhibitor of human platelet aggregation. *Phytother Res* 19: 437-440.