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To cite this article: Manzoor Ahmad, Waqar Ahmad, Mansoor Ahmad, Muhammad Zeeshan, Obaidullah & Farzana Shaheen (2008) Norditerpenoid alkaloids from the roots of *Aconitum heterophyllum* Wall with antibacterial activity, Journal of Enzyme Inhibition and Medicinal Chemistry, 23:6, 1018-1022, DOI: [10.1080/14756360701810140](https://doi.org/10.1080/14756360701810140)

To link to this article: <https://doi.org/10.1080/14756360701810140>



Published online: 20 Oct 2008.



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SHORT COMMUNICATION

Norditerpenoid alkaloids from the roots of *Aconitum heterophyllum* Wall with antibacterial activity

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(Received 15 August 2007; accepted 25 September 2007)

Abstract

Two new aconitine-type norditerpenoid alkaloids 6-dehydroacetylsepaconitine (1) and 13-hydroxylappaconitine (2), along with three known norditerpenoid alkaloids lycotoonine, delphatine and lappaconitine were isolated from the roots of the *Aconitum heterophyllum* Wall. These compounds exhibited significant antibacterial activity. The structure of compound 1 and 2 were deduced on the basis of their spectral data.

Keywords: Norditerpenoid alkaloids, *Aconitum heterophyllum* Wall, antibacteria

Introduction

Genus *Aconitum* is a rich source of diterpenoid alkaloids, many of which exhibited a broad spectrum of biological activities. Lappaconitine hydrobromide has been used as an antiarrhythmic drug [1]. The methyllycaconitine perchlorate is used in curare-mimetic preparation [2]. Some aconitine and mesaconitine derivatives possess potent analgesic and anti-inflammatory activities [3]. The methyllycaconitine and lycaconitine exhibited neuronal nicotinic acetylcholine receptor affinity [4]. Lycaconitine, obtained from several *Aconitum* species, was found to be effective against multi-drug resistance cancers. *Aconitum* plants are widely used in Chinese and Indian traditional systems of medicine [5–6]. Turkish *Aconitum* species are used externally in the treatment of rheumatic pain and sciatica and also against body lice [7].

Previously, heterophyllisine, heterophylline, heterophyllidine, heteratisine, atisine, atidine, F-dihydroatisine, hetisine, benzoylheteratisine and atisenol were reported from *A. heterophyllum* [8–11]. In the present paper, we describe the isolation and structure elucidation

of two new antibacterial norditerpenoid alkaloids 6-Dehydroacetylsepaconitine (1) and 13-hydroxylappaconitine (2), along with three known alkaloids lycotoonine (3), delphatine (4) and lappaconitine (5).

Experimental

General experimental

Optical rotations were measured on a JASCO DIP 360 polarimeter. IR spectra were recorded on a JASCO 302-A spectrophotometer. EI-MS and HREI-MS were recorded on JMS HX 110 with data system and on JMS-DA 500 mass spectrometers. The ¹H- and ¹³C-NMR spectrums were recorded on Bruker NMR spectrometers operating at 400 MHz, (100 and 125 MHz for ¹³C). The chemical shifts values are reported in ppm (δ) units and the coupling constants (J) are given in Hz.

Chromatographic conditions

For TLC, precoated aluminium sheets (silica gel 60F-254, E. Merck) were used. Visualization of the TLC

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plates was achieved under UV at 254 and 366 nm and by spraying with Dragendorff's reagent. Solvent system; "*n*-hexane–acetone–diethylamine 8:2:10 drops", was used.

Plant material

The roots (5 kg dry wt) of *Aconitum heterophyllum* Wall, were collected from District Swat of Pakistan at an elevation of 2000 m in August 2005 and identified by Mr Mehboob ur Rahman (Assistant Professor) Department of Botany, Jahanzeb Post Graduate College, Saidu Sharif, Swat, NWFP, Pakistan. The voucher specimen (HA-014) is deposited in the herbarium of the botany department.

Extraction and isolation

Dried and powdered roots (5 Kg) of the plant were extracted exhaustively with *n*-hexane which solvent extract *n*-hexane (3 × 8 L) followed by 80% EtOH (3 × 10 L) extraction at room temperature for 7 days (3-times). The filtrate was evaporated in *vacuo* to yield 60 g of residue. The residue was acidified to pH 1.5 by 0.5 N H₂SO₄ and extracted with CH₂Cl₂ (3 × 2 L) collected alkaloidal mixture (18 g). The acidic aqueous solution was basified (pH 8–10) by using 10% KOH and extracted with CH₂Cl₂ (5 × 2 L) to yield 13.8 g of crude acidic compounds fraction. The crude acidic compounds fraction was fractionated on silica gel column (260 g), five combined fractions were obtained. On repeated flash column chromatography using solvent system *n*-hexane–acetone–diethylamine (9:1:10 drops per 100 ml) 6-dehydroacetylsepaconitine (1), 13-hydroxylappaconitine (2), along with lycoctonine, delphatine and lappaconitine known alkaloids were obtained.

6-dehydroacetylsepaconitine (1), amorphous powdered (13 mg). mp 122–125°C; [α]_D³⁰ + 23.33° (*c* 0.8, CHCl₃); UV (EtOH) λ_{\max} (log ϵ) 310 (3.75), 254 (4.22), and 225 (4.46) nm; IR ν_{\max} CHCl₃, 3492 (OH groups), 1700 (ester carbonyl, amide carbonyl), 1600, 1280, 1250, and 750 cm⁻¹ (1,2-substituted aromatic ring), 1083 (simple ether bonds); ¹H-NMR (400 MHz, CDCl₃): δ 1.17 (3H, t, \mathcal{J} = 7.0 Hz, N-CH₂CH₃), δ 2.82 (1H, br s, C-17), δ 3.85 (1H, t, \mathcal{J} = 7.9 Hz, C-1), δ 3.71 (1H, d, \mathcal{J} = 4.8 Hz, C-14), δ 2.45 (1H, br s, C-5), δ 2.11 (1H, br s, C-7), δ 3.24 (1H, t, \mathcal{J} = 8.4 Hz, C-16), δ 11.0 (NH). ¹³C-NMR (Table I). HREI-MS ($M^{+}m/z$, 614.6828)

13-hydroxylappaconitine (2), amorphous powdered (10 mg). mp 145–147°C; [α]_D³⁰ + 10.33° (*c* 1.0, CHCl₃); IR ν_{\max} CHCl₃, 3492 (OH groups), 1720 (ester carbonyl, amide carbonyl), 1608, 1270, 1255, and 790 cm⁻¹ (1,2-substituted aromatic ring), 1089 (simple ether bonds); ¹H-NMR (400 MHz, CDCl₃): δ 1.03 (3H, t, \mathcal{J} = 7.0 Hz, N-CH₂CH₃), δ 3.41 (1H, br s, H-17), δ 3.20 (2H, dd, \mathcal{J} = 9.0 and 6.0 Hz, H-1 and

Table I. ¹³C-NMR data of compounds 1 and 2 (CDCl₃).

| C. No | 6-dehydroacetylsepaconitine (1) | | 13-hydroxylappaconitine (2) | |
|-------------------|---------------------------------|-----------------|-----------------------------|-----------------|
| | δ_C | Multiplicity | δ_C | Multiplicity |
| 1 | 77.7 | CH | 84.8 | CH |
| 2 | 26.5 | CH ₂ | 26.7 | CH ₂ |
| 3 | 31.6 | CH ₂ | 33.1 | CH ₂ |
| 4 | 84.6 | C | 84.6 | C |
| 5 | 44.4 | CH | 47.9 | C |
| 6 | 206 | C | 27.2 | CH ₂ |
| 7 | 46.8 | CH | 47.8 | CH |
| 8 | 79.5 | C | 68.4 | C |
| 9 | 79.6 | C | 79.2 | C) |
| 10 | 78.9 | C | 43.8 | CH |
| 11 | 55.7 | C | 53.1 | C |
| 12 | 37.4 | CH ₂ | 35 | CH ₂ |
| 13 | 34.5 | CH | 78.1 | CH |
| 14 | 87.7 | CH | 87.4 | CH |
| 15 | 44.7 | CH ₂ | 32.4 | CH ₂ |
| 16 | 82.7 | CH | 82.2 | CH |
| 17 | 61.5 | CH | 58 | CH |
| 19 | 55.5 | CH ₂ | 55.8 | CH ₂ |
| 1' | 115.7 | C | 115.7 | C |
| 2' | 141.6 | C | 140.1 | C |
| 3' | 120.2 | CH | 120.3 | CH |
| 4' | 134.4 | CH | 134.4 | CH |
| 5' | 122.3 | CH | 122.3 | CH |
| 6' | 130.9 | CH | 131 | CH |
| N-CH ₂ | | | | |
| | | | | |
| CH ₃ | 48.9 | CH ₂ | 49 | CH ₂ |
| | | | | |
| C=O | 13.4 | CH ₃ | 13.5 | CH ₃ |
| | | | | |
| C=O | 167 | C | 159.7 | C |
| | | | | |
| N-C=O | | | | |
| | | | | |
| CH ₃ | 169.3 | C | 168 | C |
| | 21.4 | CH ₃ | 25.2 | CH ₃ |

H-16), δ 4.32 (1H, s, H-14), δ 2.45 (1H, br s, H-5), δ 2.35 (1H, br s, H-7), δ 8.68 and 7.92 (each 1H, dd, Ar-H) 7.50 and 7.04 (each 1H, t, Ar-H). ¹³C-NMR (Table I). FAB (+ ve and - ve) ($M^{+}m/z$, 601.0 and 599.0).

Antibacterial activity

All of the isolated compounds were screened against strains of *Escherichia coli*, *Bacillus subtilis*, *Shigella flexneri*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Salmonella typhi*. For antibacterial screening, 3 mg of sample was taken and dissolved in 3 ml of DMSO. Molten nutrient agar (45 mL) was poured on sterile petri plates, where it was allowed to solidify. Bacterial spread were made on these nutrient agar plates by dispensing 7 mL of sterile soft agar containing 100 μ L of test-organism culture. Wells were dug with the help of a 6-mm sterile metallic borer at appropriate distance. Then, 100 μ L of sample was poured into each well, and the plates were incubated at 37°C for 24 h. The results, in terms of inhibition zone, were noted. The drug Imipenem, a broad-spectrum β -lactam antibiotic, was used as a positive control.

Table II. Table-2 Antibacterial activities of compounds 1-5. Relative to the standard drug Imipenem. For details, see Experimental.

| Compound | EC | BS | SF | SA | PA | ST |
|----------|----|----|----|----|----|----|
| Imipenem | 30 | 33 | 27 | 33 | 24 | 25 |
| 1 | — | 15 | 12 | 24 | — | 15 |
| 2 | 10 | 15 | 10 | — | 13 | 17 |
| 3 | 11 | 16 | 15 | 15 | 17 | 17 |
| 4 | — | 12 | — | 15 | 16 | 18 |
| 5 | — | 15 | 15 | — | 17 | 17 |

Inhibition zones are given in mm. Abbreviations: EC, *Escherichia coli*; BS, *Bacillus subtilis*; SF, *Shigella flexneri*; SA, *Staphylococcus aureus*; PA, *Pseudomonas aeruginosa*; ST, *Salmonella typhi*.

As a negative control, DMSO was used. The results are summarized in Table II.

Results and discussion

6-dehydroacetylsepaconitine (1) was obtained as a white amorphous powdered, and was assigned the molecular formula $C_{32}H_{42}N_2O_{10}$, on the basis of HREI-MS (m/z 614.6828, calcd. 614.6834). The mass spectrum of 1 was characteristic for diterpene bases of the C-18 series esterified through hydroxyl group at C-4 (lappaconitine, sepaconitine, *N*-deacetylappacinitine, etc), where the maximum peak corresponds to the ejection of a molecule of acid from the molecular ion. In the mass spectrum of compound 1, the peak of the $(M - 179)^+$ ion was the maximum peak and corresponded, as in the mass spectrum of *N*-acetylsepaconitine, to the ejection of acetylanthranilic acid [12]. The IR spectrum showed of compound 1 showed absorption bands at 3492 (OH

groups), 1700 broadened band, ester carbonyl, amide carbonyl), and bands at 1600, 1280, 1250, and 750 cm^{-1} (1,2-substituted aromatic ring), 1083 (simple ether bonds). The ^1H - and ^{13}C -NMR spectra of 6-dehydroacetylsepaconitine (1) exhibited a close resemblance to that of the known compound *N*-acetylsepaconitine (3) [12] except the presence of carbonyl group at C-6, instead of methylene group in compound 1.

The ^1H -NMR spectrum of 6-dehydroacetylsepaconitine (1) exhibited signals for *N*-ethyl, three methoxy groups and several methine protons with geminal oxygen substituents. In the down field region of the spectrum a doublet of one proton at δ 3.71 ($J = 4.8\text{ Hz}$), characteristic for H-14. Triplet of three protons integration at δ 1.17 ($J = 7.0\text{ Hz}$), was due to the methyl of *N*-ethyl group. Similarly, in the down field region a triplet of one proton at δ 3.85 ($J = 7.9\text{ Hz}$), was assigned to the H-1, geminal to methoxy group. A broad singlet of one proton at δ 2.82 was assigned to the H-17, whereas, a broad singlet of one proton at δ 2.45, was assigned to the H-5, while, another singlet of one proton at δ 2.11 was assigned to H-7. The ^{13}C -NMR spectrum (BB, DEPT) (Table I), showed thirty two signals, including five methyls, six methylene, eleven methine, and ten quaternary carbons. Comparing the ^{13}C -NMR data of compound 1 with those of the reported compound *N*-acetylsepaconitine (3), (Table I), the appearance of a new quaternary carbon signal at δ 206.9, and the disappearance of CH_2 signal at δ 24.5, indicated the presence of carbonyl group at C-6. The ^1H - ^{13}C correlation was determined by the HMQC spectrum,

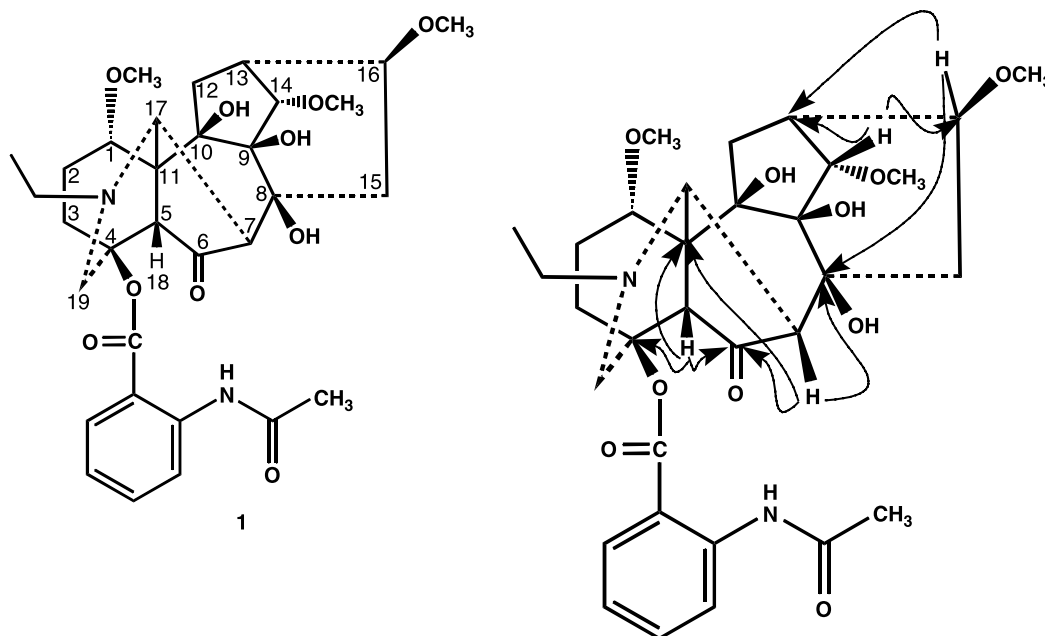


Figure 1. Selective HMBC interactions in compound 1.

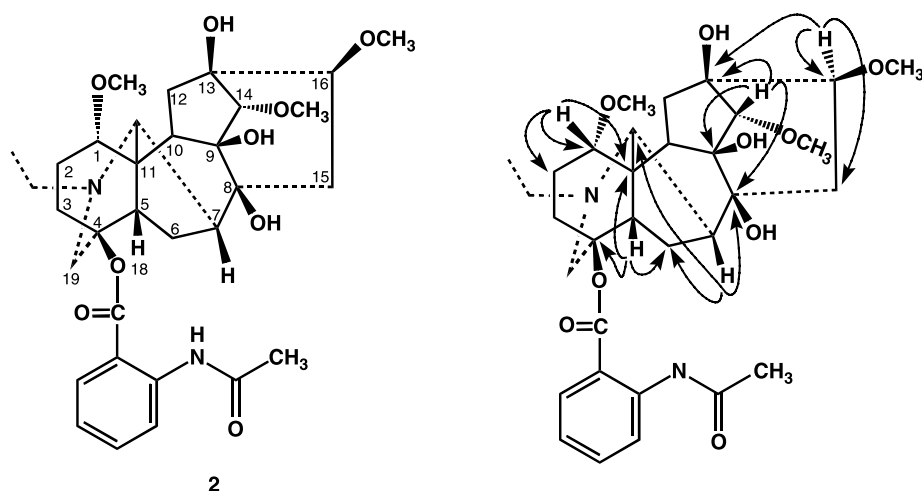
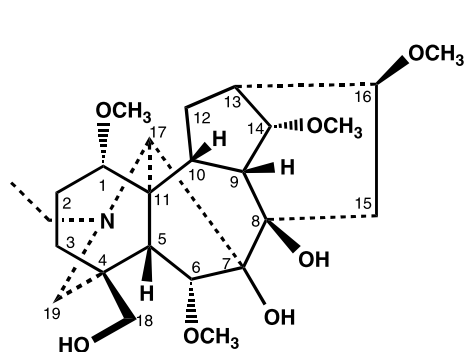


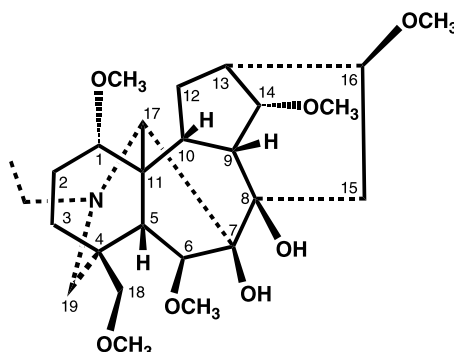
Figure 2. Selective HMBC interactions in compound 2.

while the long range ^1H - ^{13}C connectivities were obtained through HMBC technique (Figure 1). The H-5 (δ 2.45) showed ^2J and ^3J correlation with C-4 (δ 84.6), C-11 (δ 55.7), C-6 (δ 206.9) and C-17 (δ 61.5), whereas H-7 (δ 2.11), exhibited ^1J and ^2J correlation with C-6 (δ 206.9), C-7 (δ 46.8), C-8 (δ 74.5), and C-11 (δ 55.7), while, H-14 (δ 3.71), showed correlation with C-13 (δ 34.5), C-16 (δ 82.7).

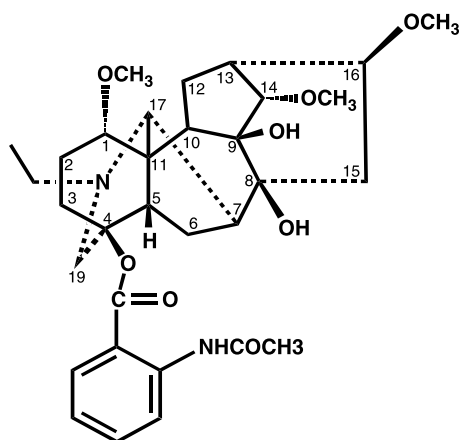
Thus on the basis of above spectral data, the structure of compound 1 was deduced as 6-dehydroacetylsepaconitine.



Lycoctonine (3)



Delphatine(4)



Lappaconitine(5)

13-hydroxylappaconitine (2) was obtained as a white amorphous powdered, and was assigned the molecular formula $\text{C}_{32}\text{H}_{44}\text{N}_2\text{O}_9$, on the basis of HREI-MS (m/z 600.7003, calcd. 600.7012). The IR spectrum showed of compound 2 showed absorption bands at 3492 (OH groups), 1700 broadened band, ester carbonyl, amide carbonyl), and bands at 1600, 1280, 1250, and 750 cm^{-1} (1,2-substituted aromatic ring), 1083 (simple ether bonds). The mass fragmentation of 2 is characteristic of alkaloids with aconitine skeleton. The ^1H - and ^{13}C -NMR spectra of 13-hydroxylappaconitine (2) exhibited a close resemblance to that of the known compound *lappaconitine* (5) [13], except the presence of hydroxyl group at C-13, instead of methine group in compound 2.

The ^1H -NMR spectrum of 13-hydroxylappaconitine (**2**) exhibited signals for *N*-ethyl, three methoxy groups and several methine protons with geminal oxygen substituents. In the down field region of the spectrum a singlet of one proton at δ 4.32 was assigned to H-14. Similarly, a broad singlet of one proton in the down field region at δ 10.78 was assigned to the amide proton. Triplet of three protons at δ 1.03 ($J = 7.0$ Hz), was due to the methyl of *N*-ethyl group. Similarly, a double doublet of two protons at δ 3.21 ($J = 9.0, 6.0$ Hz), was assigned to the H-1 and H-16, geminal to methoxy group. A broad singlet of one proton at δ 2.45 was assigned to the H-5. The ^{13}C -NMR spectrum (BB, DEPT) (Table I), showed thirty two signals, including five methyls, seven methylene, eleven methine, and nine quaternary carbons. Comparing the ^{13}C -NMR data of compound **2** with those of the reported compound lappaconitine (**5**), (Table I), the appearance of a new quaternary carbon signal at δ 78.1, and the disappearance of CH signal at δ 49.0, indicated the presence of hydroxyl group at C-13. The ^1H - ^{13}C correlation was determined by the HMQC spectrum, while the long range ^1H - ^{13}C connectivities were obtained through HMBC technique (Figure 2). The H-5 (δ 2.45) showed correlation with C-4 (δ 84.6), C-11 (δ 53.1), and C-6 (δ 27.2), whereas H-7 (δ 2.11), exhibited correlation with C-6 (δ 27.2), C-7 (δ 47.8), C-8 (δ 68.4), and C-17 (δ 58.0), while, H-14 (δ 3.71), showed correlation with C-13 (δ 78.1), C-14 (δ 87.4), C-9 (δ 79.2) and C-8 (δ 68.4), Similarly, H-16 (δ 3.33) exhibited interaction with C-16 (δ 82.2), C-15 (δ 32.4) and C-13 (δ 78.1).

Thus on the basis of above spectral data, the structure of compound **2** was deduced as 13-hydroxylappaconitine.

Antibacterial activity: Compounds **1** and **2** showed significant antibacterial activity against

Staphylococcus aureus. Compound **3** showed significant activities against *Salmonella typhi* and *Pseudomonas aeruginosa*, while compounds **4** and **5** showed significant activity against *Salmonella typhi* and *Pseudomonas aeruginosa* (Table II).

Acknowledgements

We are thankful to Professor Mehboob-ur-Rahman for helping in the collection and identification of the plant.

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