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Published online: 20 Oct 2008.

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Enzyme kinetic and molecular modelling studies of sulphur-containing substrates of phenylalanine 4-monooxygenase

N. G. PATEL[1], C. ILIADOU[1], B. BOONYAPIWAT[1], D. J. BARLOW[1], B. FORBES[1], S. C. MITCHELL[2], & G. B. STEVENTON[1]

[1]*Pharmaceutical Sciences Division, School of Biomedical and Health Sciences, King's College London, Franklin Wilkins Building, 150 Stamford Street, London SE1 9NH, UK, and* [2]*Faculty of Medicine, Department of Biomolecular Medicine, Imperial College London, Sir Alexander Fleming Building, South Kensington, London SW7 2AZ, UK*

Abstract

Previous investigations into the binding of substrates/cofactors to the PAH active site have only concentrated on Phe, thienylalanine and BH_4. This is the first reported investigation to model aliphatic thioether amino acid substrates to PAH. The clearance of the thioether substrates (4.82-79.09% of Phe) in the rat and human (1.19-37.41% of Phe) showed species differences. The xenobiotic thioether substrates (SMC and SCMC) were predicted to be poor substrates for PAH by the molecular modelling investigation and this has now been confirmed by the *in vitro* enzyme kinetic data. However, reaction phenotyping investigations have found that PAH was the major enzyme involved in the metabolism of SCMC *in vitro* and *in vivo*.

Keywords: *Phenylalanine 4-monooxygenase, thioether substrates, S-oxidation, inhibition, computer modelling*

Introduction

The role played by phenylalanine 4-monooxygenase (PAH) in the conversion of L-phenylalanine (Phe) to L-tyrosine (Tyr) has been under active investigation ever since it was first associated with the disease phenylketonuria (PKU) [1]. The genetic mutations that result in PKU and hyperphenylalaninemia due to PAH deficiencies are now well known [2]. The PAH enzyme is regulated by a variety of pre- and post-translational mechanisms *in vivo* and *in vitro*. At the pre-translational level *PAH* is activated by the glucocorticoid receptor [1]. However, at the post-translational level PAH is activated by a number of processes [3].

PAH is a homo tetrameric protein. The monomer subunits have no catalytic activity but the dimer and tetramers all possess enzyme activity and the full enzyme is in fact a dimer of dimers [3]. Each monomer comprises, an N-terminal regulatory domain (amino acids 1-142), a catalytic domain (amino acids 143-410) and a C-terminal polymerization domain (amino acids 411-452). The major physiological control of PAH activity *in vivo* is by binding of Phe and/or phosphorylation of the allosteric enzyme [3]. This results in changes in both K_m and V_{max} for Phe but does not alter the enzyme's substrate specificity. The binding of lysophosphatidycholine, modification of Cys^{237} or limited proteolysis all can activate PAH. These activation mechanisms not only alter the K_m and V_{max} for Phe, but also alter the substrate specificity of the enzyme as well [3]. L-Methionine (Met), S-methyl-L-cysteine (SMC), S-carboxy-methyl-L-cysteine (SCMC) and nor-leucine are all substrates for activated PAH [3,4].

Correspondence: G. B. Steventon, Pharmaceutical Sciences Division, School of Biomedical and Health Sciences, King's College London, Franklin Wilkins Building, 150 Stamford Street, London SE1 9NH, UK. Tel: 44 207 848 4842. Fax: 44 207 848 4800. E-mail: glyn.steventon@kcl.ac.uk

Both SMC and SCMC are mucolytic drugs used in the treatment of chronic obstructive pulmonary disease (COPD) and otitis media with effusions (OME). The clinical efficacy of these two drugs in patients has been the subject of debate for a number of years. However, it is clear that while some patients respond very well to the mucolytic drugs others show little or no benefit. This observed variation in response to these mucolytic agents could be linked to the human genetic polymorphism in the S-oxidation of SCMC in man [5]. The major urinary metabolites of SCMC in man are the S-oxides (SCMC S-oxide, SMC S-oxide, N-acetyl SMC S-oxide, N-acetyl-SCMC S-oxide and thiodiglycolic acid S-oxide). Thus the polymorphism in the S-oxidation of SCMC will have a major effect on the clinical efficacy of SCMC since the mechanism of action of the drug is now attributed to its activity as a free radical scavenger. The thioether drug is converted to SCMC S-oxide by reactive oxygen intermediates [6]. Since 65% of the population are extensive metabolisers, 32% are intermediate metabolisers and 3% are poor metabolisers with regard to the S-oxidation of SCMC[7], the enzyme responsible for the S-oxidation of this drug will have a major effect on the level of the free radical scavenger in the body.

The role that PAH plays in the metabolism of the thioether mucolytic drugs is poorly understood, and the aims of this investigation were to model the binding of the thioether drugs to the PAH active site and to compare the predicted binding interactions with the K_m and V_{max} values determined experimentally.

Materials and methods

Molecular modelling of human PAH-substrate complexes

Model building and visualisation of the PAH-substrate complexes was performed using HyperChem™ version 7.1 and WebLab ViewerLite® version 5.0.

The template selected for the model building was provided by the 2.0 Å resolution crystal structure for the catalytic domain of human PAH ternary complex with norleucine and tetrahydrobiopterin (BH_4)[8]. Atomic co-ordinates for this complex were obtained from the Protein Data Bank [9] (structure code: 1 mmt).

The docking of substrates into the PAH active site was performed manually but guided by the pose of the crystal structure ligand, norleucine. The resulting models were then refined by means of potential energy minimization using the AMBER force field [10].

In vitro *PAH enzyme assays*

Enzyme assays. Direct PAH activity was determined by the method of Pey et al. [11] with the following modifications. All incubations were carried out at 37°C

for 10 min. Each enzyme assay contained (rat pooled cytosolic fractions, 2.0 mM phenylalanine/40.0 mM SCMC/50.0 mM SMC/20.0 mM Met, human pooled cytosolic fractions 2.0 mM phenylalanine/20.0 mM SCMC/40.0 mM SMC/15.0 mM Met), 50.0 mM potassium phosphate buffer (pH 6.8), catalase (7,800 units/mL), human/rat liver cytosol (200 μg/mL), 6.0 mM DTT, 100 μM BH_4 in a total volume of 1.0 mL. Reactions were terminated by addition of 100 μl of 10% (w/v) TCA. Acid supernatants were prepared by centrifugation at 3,000 x g for 10 min. Blanks were assayed as described but using heat (100°C) in-activated rat or human cytosolic fractions.

Indirect PAH activity for the S-oxidation of Met, SCMC, SMC and S-methyl-ergothionine (SME) was determined using the method of Pey et al.[11]. All incubations were carried out at 37°C. The Δ340 nm was measured for 1.0 min by using $\varepsilon_{340\,nm} = 6220$ $M^{-1}\cdot cm^{-1}$. Each enzyme assay contained (rat pooled cytosolic fractions, 2.0 mM phenylalanine/40.0 mM SCMC/50.0 mM SMC/30.0 mM SME/20.0 mM Met, human pooled cytosolic fractions 2.0 mM phenylalanine/20.0 mM SCMC/40.0 mM SMC/ 2.0 mM SME/15.0 mM Met), 50.0 mM potassium phosphate buffer (pH 6.8), catalase (7,800 units/mL), human/rat liver cytosol (200 μg/mL), 200 μM NADH(H⁺), 1.0 units/mL dihydropteridine reductase, 100 μM BH_4 in a total volume of 1.0 mL. Blanks were assayed as described but containing both 200 μg/mL PH8 (monoclonal antibody against PAH) and 1.0 mM 4-chlorophenylalanine (specific PAH inhibitor).

HPLC assays

The quantification of the S-oxides of Met, SCMC and SMC was carried out by HPLC as reported by Hirschberger et al. [12], and the quantification of Tyr as reported by Doskeland et al. [13].

Synthesis of S-carboxymethyl-L-cysteine and S-methyl-L-cysteine S-oxides

The synthesis of the mixture of SCMC (*S/R*) sulphoxide and SMC (*S/R*) diastereoisomers was based on the methods of Meese [14] and Meese et al. [15].

Synthesis of S-methyl-ergothionine

The synthesis of S-methyl-ergothionine (SME) was based on the method of Theodoropoulos [16].

Data analysis

Enzyme kinetic data were analysed using WinNonLin version 4.1 and Leonora version 1.0. Statistical analysis was undertaken using SPSS version 10.0.

Table I. Molecular modelling of human PAH active site.

Substrate	Distance of oxidation site from (Å)			
	Fe^{2+}	H_2O	BH_4 (oxygen)	
Phe	3.09*	2.17	4.53	5.19
Met	3.69*	3.89	6.36	6.24
SMC	4.92	4.40	6.98	6.65
SCMC	4.93	4.40	6.96	6.65
ERG	3.14*	1.88	3.71	5.46

*Distance <4.2 Å is adequate for the introduction of molecular O_2 [18].

Results and discussion

Molecular modelling studies

Results obtained in the molecular modelling studies are summarised in Table I and Figures 1a–1e. For each complex, attention was focused on the proximity of the substrate to the active site Fe^{2+}, BH_4 cofactor, molecular oxygen and bound water. In addition to the modelling of the PAH-Phe, PAH-SMC, and PAH-SCMC complexes, studies were also made of the complex formed between the PAH enzyme and ergothionine (Erg). This latter compound was identified as a potential substrate for the enzyme following a manual search for commercially available compounds having the appropriate molecular properties and dimensions.

For the PAH-Phe complex (Table I and Figure 1) the Arg[270] amino acid from one side of the active site channel is involved in an ionic interaction with the α-carboxyl group of the substrate. The distance between the carboxyl oxygen atoms and the amino nitrogen atoms of Arg[270] are 1.98 Å and 3.15 Å. On the opposite side of the channel, the side chain OH of Ser[349] hydrogen bonds with the α-amino group of Phe and the phenyl ring of Phe is in close proximity to Pro[281]. The site of oxidation of Phe is 2.17 Å from the bound water molecule and 3.09 Å from the Fe^{2+} ion.

The latter distance is <4.2 Å and thus deemed adequate for intercalation of molecular oxygen in the hydroxylation reaction[17]. The site of oxidation on the phenyl ring is 4.33 Å and 5.19 Å from the oxygen atoms on the cofactor, BH_4.

For the aliphatic amino acids, Met, SMC and SCMC, the distances of the site of oxidation from the Fe^{2+} were 3.69 Å, 4.92 Å and 4.93 Å respectively (Table I). The predicted substrate Erg, has a sulphur atom at 3.14 Å from the Fe^{2+}. Thus from the modelling studies the preferred substrates for human PAH in terms of orientation and conformation within the catalytic site were Phe > Erg > Met > SMC ≈ SCMC.

Enzyme kinetic studies

The *in vitro* enzyme kinetics of both female rat and human PAH in pooled cytosolic fractions were investigated in order to assess the predicted substrate specificity. A summary of the results obtained in these is presented in Table II.

The rate of clearance of the substrates via rat PAH was Phe > Met > SCMC > SMC (Cl_E 14.30 ± 0.72, 8.24 ± 0.41, 3.26 ± 0.16, 0.69 ± 0.03 μl min^{-1} mg protein^{-1}). The predicted substrate, Erg was assayed via the coupled enzyme assay since no Erg sulphenic or sulphinic acid could be synthesed or found commercially. No activity was detected but when an inhibitor investigation of Erg and PAH was undertaken to determine the K_i and type of inhibition versus Phe, the predicted substrate was found to be an activator of rat PAH activity *in vitro* (results not shown). Since the sulphur containing substrates Met, SMC and SCMC are all sulphides and Erg was a sulphydryl compound, the sulphydryl group was converted to a sulphide group via methylation. The new potential substrate, S-methyl-ergothionine (SME) was found to be a substrate for rat PAH in the coupled assay with a
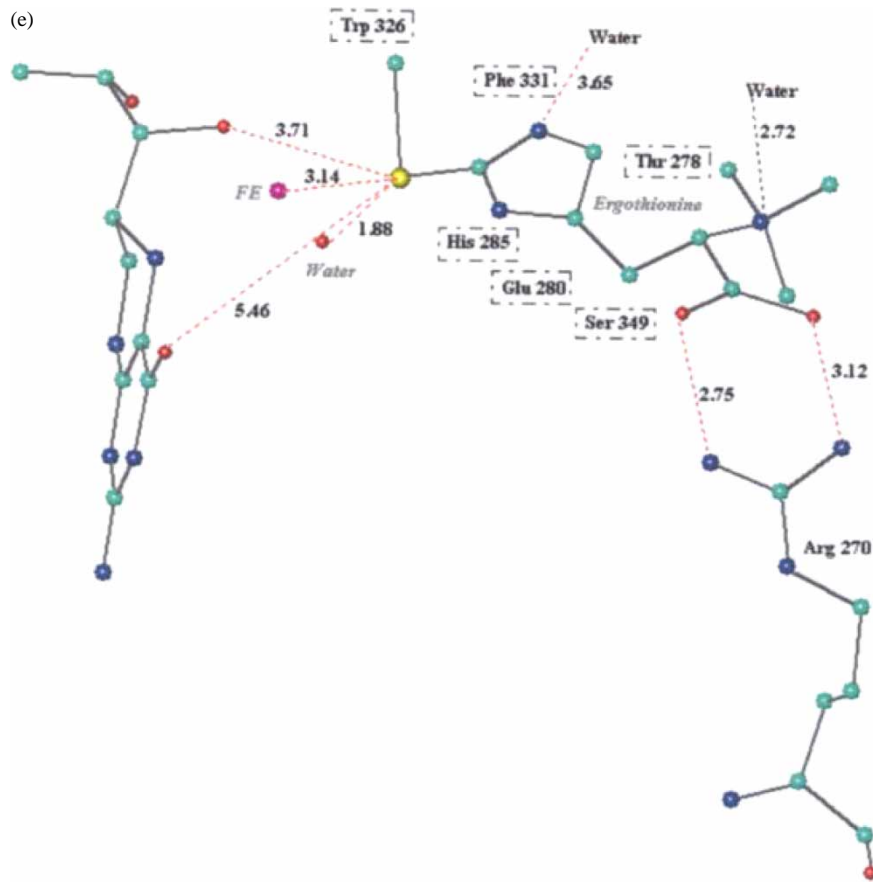
(a)

(b)

(c)

(d)
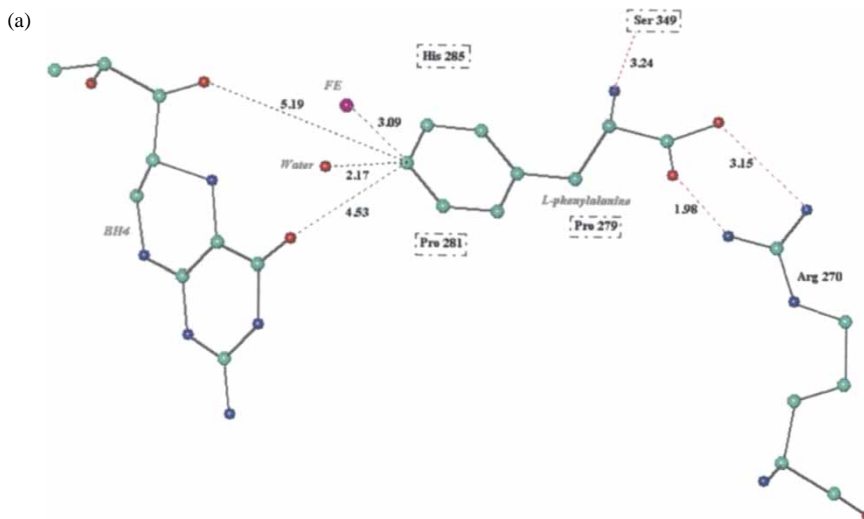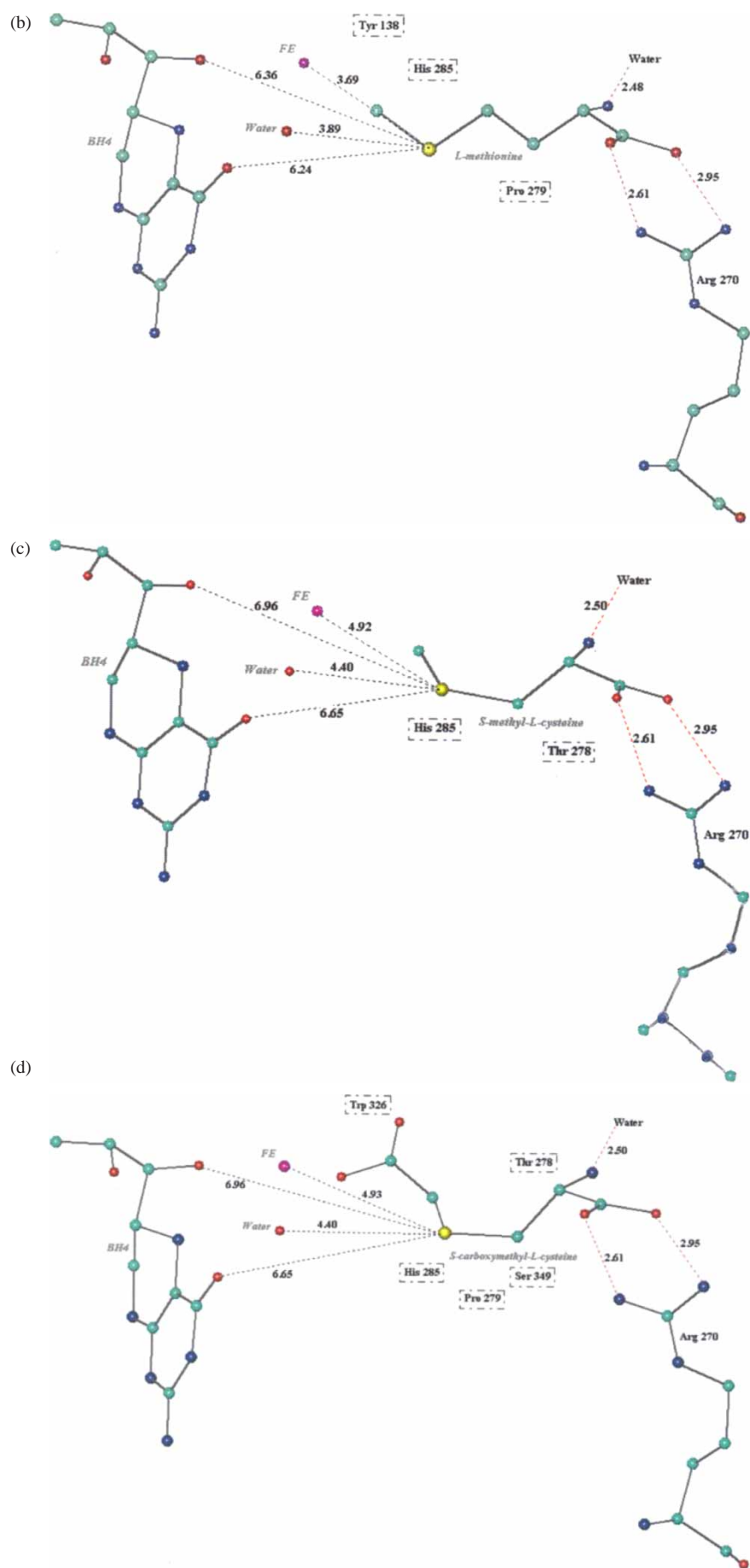
Figure 1. (a) 3-D schematic diagram of Phe-PAH interactions. (b) 3-D schematic diagram of Met-PAH interactions. (c) 3-D schematic diagram of SMC-PAH interactions. (d) 3-D schematic diagram of SCMC-PAH interactions. (e) 3-D schematic diagram of SME-protein interactions. Molecular models of the predicted complexes formed between PAH and the substrates Phe (a), Met (b), SMC (c), SCMC (d) and SME (e). Atoms are colour coded as C: green, N: blue, O: red, Fe: pink, S: yellow. Potential hydrogen bonds are indicated by dashed red lines, and PAH residues in van de Waals contact with the ligands are boxed. All inter atomic distances are given in Å.

clearance value of $11.31 \pm 8.9\,\mu l\,min^{-1}\,mg\,protein^{-1}$. Thus, from the clearance date in Table II, the substrate specificity of rat PAH was Phe > SME > Met > SCMC > SMC. The inhibitor binding constant (K_i) and type of inhibition were also determined experimentally. All substrates (SME, Met, SCMC, SMC) were found (via Dixon plots) to be competitive inhibitors of the C-oxidation of Phe (results not shown). The experimentally determined K_i values for the

inhibitors were not significantly different from their experimentally determined K_m values ($P > 0.05$, Student t-test) indicating that the binding constants for the compounds acting as substrates and inhibitors were the same. The assay of PAH using Phe, Met, SCMC and SMC was undertaken by two assays. The direct assay involved HPLC identification and quantification of the product of the enzyme reactions (Tyr, Met *R/S* S-oxides, SCMC *R/S* S-oxides and SMC

Table II. Enzyme kinetic data for activated pooled rat cytosolic fraction PAH assays using Phe, Met, SME, SMC and SCMC as substrates.

Substrate	n	K_m (mM)	V_{max} (nmole min^{-1} mg protein^{-1})	Cl_E (μl min^{-1} mg protein^{-1})	K_i (mM)	Coupling efficiency
Phe	6	0.26 ± 0.08	3.72 ± 4.74	14.30 ± 0.72	ND	0.95 ± 0.05
Met	6	7.75 ± 3.01	63.88 ± 6.81	8.24 ± 0.41	8.11 ± 0.44	0.94 ± 0.06
SMC	6	43.25 ± 7.76	29.73 ± 3.79	0.69 ± 0.03	41.53 ± 2.49	0.94 ± 0.05
SCMC	6	14.73 ± 8.91	48.11 ± 10.69	3.26 ± 0.16	13.14 ± 0.66	0.95 ± 0.05
SME	6	0.45 ± 0.05	5.09 ± 1.34	11.31 ± 8.9	0.50 ± 0.04	ND

n = number of experiments. Each experiment was determined in duplicate with associated blanks. PAH was activated by pre-incubation with 1.0 mM lysophosphatidylcholine for 3.0 min at 37°C. $Cl_E = V_{max}/K_m$. Coupling efficiency = ratio of metabolite produced (Tyr, Met S-oxides, SMC S-oxides, SCMC S-oxides)/NADH oxidised using 2.0 mM phenylalanine/40.0 mM SCMC/50.0 mM SMC/20.0 mM Met and 100 μM BH$_4$. ND = not determined.

Table III. Enzyme kinetic data for activated pooled human cytosolic fraction PAH assays using Phe, Met, SME, SMC and SCMC as substrates.

Substrate	n	K_m (mM)	V_{max} (nmole min^{-1} mg $protein^{-1}$)	Cl_E (μl min^{-1} mg $protein^{-1}$)	K_i (mM)	Coupling efficiency
Phe	6	0.64 ± 0.03	91.21 ± 5.47	142.51 ± 0.71	ND	0.96 ± 0.04
Met	6	3.10 ± 0.16	59.13 ± 2.95	18.59 ± 1.01	3.50 ± 0.18	0.95 ± 0.05
SME	6	0.30 ± 0.03	15.97 ± 1.25	53.31 ± 2.67	0.41 ± 0.03	ND
SMC	6	18.32 ± 1.00	31.28 ± 1.41	1.70 ± 0.09	17.32 ± 0.77	0.94 ± 0.05
SCMC	6	4.60 ± 0.23	50.26 ± 2.57	10.92 ± 0.16	5.01 ± 0.25	0.95 ± 0.05

n = number of experiments. Each experiment was determined in duplicate with associated blanks. PAH was activated by pre-incubation with 1.0 mM lysophosphatidylcholine for 3.0 min at 37°C. $Cl_E = V_{max}/K_m$. Coupling efficiency = ratio of metabolite produced (Tyr, Met S-oxides, SMC S-oxides, SCMC S-oxides)/NADH oxidised using 1.0 mM phenylalanine/20.0 mM SCMC/40.0 mM SMC/15.0 mM Met and 100 μM BH_4. ND = not determined.

R/S S-oxides). The same assays were repeated using the coupled assay (assaying the disappearance of the cofactor $NADH(H^+)$ at 340 nm). The correlation between the disappearance of cofactor and the appearance of product (coupling factor) was 0.94-0.95 for all substrates investigated. It was thus concluded that the coupled assay could be used to investigate substrate specificity when authentic standards of the products were not available.

When the experiments using pooled female Wistar rat cytosolic fractions were repeated in pooled human female cytosolic fractions, a similar pattern of results was obtained (Table III). The rate of clearance of the substrates via human PAH was Phe > Met > SMC > SMC (Cl_E 142.51 ± 0.71, 18.59 ± 1.01, 10.92 ± 0.16, 1.70 ± 0.09 μl min^{-1} mg $protein^{-1}$). Again no activity was detected when Erg was investigated for substrate activity in the coupled assay but when an inhibitor investigation of Erg and PAH was undertaken to determine the K_i and type of inhibition versus Phe, the predicted substrate was also found to be an activator of human PAH activity *in vitro* (results not shown). The new potential substrate, S-methyl-ergothionine (SME) was found to be a substrate for human PAH in the coupled assay with a clearance value of 53.31 ± 2.67 μl min^{-1} mg $protein^{-1}$. Thus, from the clearance date in Table III, the substrate specificity of human PAH was Phe > SME > Met > SCMC > SMC. The inhibitor binding constant (K_i) and type of inhibition were also determined experimentally. All substrates (SME, Met, SCMC, SMC) were found (via Dixon plots) to be competitive inhibitors of the C-oxidation of Phe (results not shown). The experimentally determined K_i values for the inhibitors were again not significantly different from their experimentally determined K_m values (P > 0.05, Student t-test), indicating that the binding constants for the compounds acting as substrates and inhibitors were the same. The assay of human PAH using Phe, Met, SCMC and SMC was again undertaken by both direct and coupled assays as with the rat PAH study. The correlation between the disappearance of cofactor and the appearance of product (coupling factor) was 0.94-0.96 for all

substrates investigated. It was again concluded, therefore, that the coupled assay could be used to investigate substrate specificity when authentic standards of products were not available in human and rat pooled cytosolic fractions for PAH activity.

Acknowledgements

References

[1] Kaufman S. Adv Enzymol Rel Areas Mol Biol 1993;67: 77–264.
[2] PAHdb: www.mcgill.ca/pahdb
[3] Hufton SE, Jennings IG, Cotton RG. Biochem J 1995;311: 353–366.
[4] Kaufman S, Mason K. J Biol Chem 1982;257:14667–14678.
[5] Mitchell SC, Waring RH, Haley CS, Idle JR, Smith RL. Br J Clin Pharmacol 1984;18:507–521.
[6] Brandolini L, Allegretti M, Berdini V, Cervellera MN, Mascagni P, Rinaldi M, Melillo G, Ghezzi P, Mengozzi M, Bertini R. Eur Cytokine Net 2003;14:20–26.
[7] Steventon GB, Sturman S, Waring RH, Williams AC. Drug Metab Drug Inter 2001;18:79–98.
[8] Andersen OA, Stokka AJ, Flatmark T, Hough E. J Mol Biol 2003;333:747–757.
[9] Merman H, Westbrook J, Feng Z, Gilliland G, Bhat TN, Weissig H, Shindyalov IN, Bourne PE. Nuc Acids Res 2000;28:235–242.
[10] Case DA, Cheatham TE, Darden T, Gohlke H, Luo R, Merz KM, Onufriev A, Simmerling C, Wang B, Woods RJ. J Comput Chem 2005;26:1668–1688.
[11] Pey AL, Martinez A, Charubala R, Maitland DJ, Teigen K, Calvo A, Pfeiderer W, Wood JM, Schallreuter KU. FASEB J 2006;20:E1451–E1464.
[12] Hirschberger LL, de la Rosa J, Stipanuk MH. J Chromatog 1985;343:303–313.
[13] Doskeland AP, Doskeland SO, Ogreid D, Flatmark T. J Biol Chem 1984;259:11242–11248.
[14] Meese CD. Arch Pharm (Weinheim) 1987;320:473–474.
[15] Meese CD, Spechat D, Hofmann U. Arch Pharm (Weinheim) 1990;323:957–965.
[16] Theodoropoulos D. Acta Chem Scand 1959;13:383–384.
[17] Teigen K, Froystein NA, Martinez A. J Mol Biol 1999;294: 807–823.
[18] Andersen OA, Flatmark T, Hough E. J Mol Biol 2002;320: 1095–1108.