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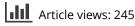
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Immunotherapy for brain tumors: *quo vadis*?

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At a recent international brain tumor advisory meeting held in Scottsdale, AZ, USA the topic of immunotherapy for brain tumors was hotly debated. After an intriguing presentation highlighting the most recent attempts at creating a successful vaccine, one of my esteemed colleagues asked a poignant question: "it would appear to me" he said, "that we have been listening to similar presentations for the last 10 years, yet there really has not been any significant clinical efficacy shown with this approach - where are we heading?" Clearly, this statement struck at the heart of those of us involved in developing novel immunological approaches for malignant brain tumors. Whether our work was called into question, or some of our egos were bruised, the fact remains that our success in this arena has been limited. This is not to say that we have not evolved and advanced our understanding of the role that the immune system plays in the CNS; in fact, the last 10 years has seen an increasing number of clinical trials involving immunomodulatory cytokines, dendritic cells (DCs), and gene therapy vectors, all of which have advanced our knowledge and understanding of brain tumor immunology. However, clinical efficacy has lagged behind these endeavors and therefore, the author thought it would be appropriate to bring us all up to date, and explore the potential benefits and limitations of brain tumor immunotherapy.

The field of tumor immunology, especially as it relates to the CNS, has undergone a significant evolution. The brain has historically been viewed as immune privileged, meaning that it is somehow protected from the immune system. The presence of a blood-brain barrier, lack of organized lymphoid tissue or significant lymphatic drainage, poor major histocompatibility complex (MHC) expression in normal brain parenchyma, and presence of immunoregulatory factors all contribute to the perception of immune privilege. While the above properties contribute to the distinctive immunoreactivity of the CNS, they do not render it completely privileged. They do, however, necessitate an understanding of intricacies of the immune system as it relates to CNS antitumor response.

Cell-mediated responses to neoplasms in the CNS require efficient antigen presentation. The precise cell type and mechanism responsible for antigen presentation in the brain has been debated; however, the ability to ingest or phagocytose foreign antigens, expression of MHC complexes, and the ability to induce T-lymphocyte responses, are all critical to an effective antitumor response. Possible antigen-presenting cells (APCs) in the CNS include astrocytes, endothelial cells, capillary pericytes, DCs and microglia. However, since the 1990s, microglia have emerged as the most likely candidates for antigen presentation in the brain. In addition to their hematopoietic origin, microglial cells possess many characteristics that APCs require, including the expression of MHC class I and II molecules and the expression of costimulatory molecules.

In addition to the issues related to antigen presentation, a second potential obstacle to CNS tumor reactivity is the relative lack of MHC expression and antigenicity in normal and malignant brain tissue. However, there is now experimental evidence that MHC antigens are upregulated at sites of brain injury, degenerative disease, tumor, or after exposure to cytokines, such as interferon (IFN)-y. In addition to antigenpresentation complexes, most human glioma cells have been shown to express Fas/Apo-1 and Fas ligand, which allow these cells to undergo Fas/Fas-ligand interactions with activated T lymphocytes, resulting in apoptosis. While no universal glioma-specific antigen presentation on these MHC molecules has been found, several tumor-associated antigens shared by histogenetically related tumors have been identified (e.g., tenascin, glycoprotein [gp]240, gp1000, epidermal growth factor receptor, tyrosinase, tyrosinase-related proteins 1 and 2, interleukin [IL]-13 α 2R, melanoma-associated gene [MAGE]-1 and MAGE-3).

Besides evading host immune responses, malignant brain tumors actively suppress the local immune environment. T-lymphocyte function in the setting of primary intracranial

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tumors has been shown to be defective with abnormal T-cell receptor-mediated signaling and a depressed proliferative response to mitogen. Moreover, tumorinfiltrating lymphocytes exhibit a propensity for apoptosis. The partial reversibility of T-cell suppression following brain tumor removal along with the demonstration of downregulated function of T cells harvested from brain tumor patients or normal patients exposed to glioma supernatants, pro-

vides strong evidence of glioma-produced immunosuppressive factors. Transforming growth factor (TGF)- β_2 has been one of the most extensively studied of all immunosuppressive factors identified thus far. It has been shown to be significantly overexpressed in patients with glioblastoma multiforme, while virtually absent in normal brain tissue. Additionally, TGF- β_2 has been shown to effectively limit T-cell and B-cell proliferation, IL-2 receptor induction, cytokine production, natural killer (NK) cell activity, cytotoxic T-lymphocyte development, lymphokine-activated killer (LAK) cell generation and the cytotoxic response of tumor-infiltrating lymphocytes. Other factors produced by glioma cells that may play a role in immune regulation and tumor cell escape include prostaglandin (PG)E₂ and IL-10.

In spite of the unique anatomical and physiological barriers posed by the CNS, there is clear evidence that priming of T cells to intracranial tumor antigens does occur although it can be significantly augmented with local immunotherapy. These data led to the investigation of various types of peripheral vaccinations as treatment strategies for brain tumors. First and foremost among them was the development of cytokineexpressing vaccines. Cytokines play critical roles in CNS tumor immunology by enhancing T-cell activation and MHC antigen expression. Cytokine gene transfer in cancer models, resulting in tumor rejection, has been demonstrated with IL-2, -4, -12, -18 and -23, IFN- α and granulocyte–macrophage colony-stimulating factor (GM-CSF). The use of peripheral, cytokine-secreting vaccines for intracranial cancers has, therefore, been explored in the hope of overriding the tumor-induced immuno-suppressive microenvironment and enhancing the activation and infiltration of host-derived APCs and leukocytes.

Several studies have demonstrated the efficacy of peripheral vaccinations with GM-CSF-transfected tumor cells against brain tumors in animal models [1–9]. Indeed, some studies have suggested intensified antitumoral responses when GM-CSF vaccines were coupled with those expressing IL-4 or the costimulatory molecule B7-2. In this setting, antitumor cyto-toxicity only exceeded that of peripheral blood mononuclear cells (PBMCs) stimulated with wild-type tumors alone, when PBMCs were stimulated with both wild-type tumor and B7-2/GM-CSF-transduced cells. Finally, peripheral tumor cell inoculation coupled with continuous, peripheral GM-CSF alone or with IL-2 or IL-12 resulted in improved survival rates

in the treatment of intracranial glial tumors in rat models. It is postulated that the antitumor activity of GM-CSF is related to its potency in generating DCs. Whether GM-CSF alone or in combination with other factors will result in CNS antitumor efficacy in humans remains to be seen.

The utility of IL-2 in the treatment of brain tumors has been investigated using multiple methods [10–17]. IL-2secreting autologous fibroblasts injected

into glioma cells have been demonstrated to induce cytotoxic CD8⁺ T cells in the blood of a patient with malignant glioma. Intracerebral injection of IL-2-secreting fibroblasts has also been found to be effective as a protective treatment in preventing the development of murine brain tumors when the tumor cells are introduced into the same site where the fibroblasts were injected earlier. Of note, this treatment has proven effective for metastatic brain tumors, including established intracerebral breast carcinoma. Combining peripheral vaccination and intratumoral injection of IL-2-expressing cells or IL-2-activated NK cells has also shown promise in the elimination of brain tumors in animal models. Also, injection of IL-2-generated LAK cells into tumor resection cavities of humans with histologically confirmed recurrent glioblastoma multiforme (GBM) was found to be safe and resulted in higher median survival rates than those reported in other published series of patients that underwent reoperation for GBM. Given the promising results of intracranial IL-2 treatment, novel means of delivery (including the use of biodegradable microspheres) have also been explored. The use of microspheres for delivery of IL-2 and the chemotherapeutic agent carmustine (BCNU) has demonstrated improved median survival when compared with controls in animal models.

The effectiveness of IL-4 for the treatment of intracranial tumors has also been explored in multiple studies [18-20]. The effectiveness of various cytokines (IL-4, IL-12, GM-CSF and IFN- α) has been compared using intracerebral or intradermal injections of transfected tumor cells, as prophylaxis or therapy against intracerebral glial tumors in an animal model. However, only intradermal IL-4-expressing cell vaccinations were shown to be effective at providing immunity against later challenges with intracranial tumors. Additionally, only IL-4vaccinated animals showed long-term survival benefits when used as therapy for existing tumors. These results corroborated data demonstrating the efficacy of subcutaneous tumor cell vaccines expressing IL-4 (with and without herpes simplex virus-thymidine kinase [HSVtk]/ganciclovir) as therapy or prophylaxis against intracranial glial tumors in animals. Recently, a single patient with recurrent right temporal GBM was vaccinated with autologous glioma cells admixed with transgene-derived IL-4-expressing fibroblasts. A transient response to the vaccine was suggested in this trial and the patient survived for 10 months after treatment. Clearly, further trials with peripheral IL-4 vaccines will be needed to better assess their effectiveness.

The subcutaneous immunization of rats with IL-12-secreting

glial tumors has been demonstrated to suppress the growth of identical cell type brain tumors [21–26]. The use of intracerebral cytokine delivery has also been explored by other means. Significantly prolonged survival times have been demonstrated in animals chal-

lenged intracranially with glioma cells after earlier intracranial paracrine delivery of IL-12 with transduced glial cells. Neural stem cells (NSC) that express IL-12 have also demonstrated effectiveness in prolonging survival time in mice and rats with existing tumors. Finally, systemic IL-12 has also demonstrated effectiveness in eliciting immunological responses against transplanted and endogenous murine CNS tumors.

Most recently, DC-based vaccines have received a significant amount of attention. The rationale for using DCs lies in their potency as APCs. With the advancement of techniques allowing for their isolation and propagation in vitro, these therapies have become more feasible in the treatment of gliomas. Each of these therapy regimens, however, requires patient-derived tumor antigens, due to the lack of a universal glioma-specific antigen. There are several reports of peripheral vaccinations with DCs loaded with glioma antigens (e.g., tumor cell extracts, lysates and RNA) or DCs fused with glioma cells (with or without recombinant IL-12) leading to the generation or augmentation of antitumor responses in the brains of animal models [27-30]. The promising results of glioma DC vaccines in animal models have led to several clinical trials in adults and children [31-36]. Initially, Phase I trials established the feasibility and safety of peptide-pulsed DCs vaccinations. Most recently, vaccination with tumor lysate-pulsed DCs was shown to elicit the generation of antigen-specific, cytotoxic T cells in patients

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with malignant gliomas. In this study, the median survival for patients with recurrent glioblastoma who were treated with the vaccine was extended to 133 weeks. Together, these results suggest that DC vaccinations are safe and can generate intracranial antitumor effects and T-cell infiltration.

Although most DC vaccines are administered peripherally, direct intracranial injection with DCs has also been explored in glioma [37–39]. Importantly, preliminary studies indicate that such direct intratumoral DC therapy appeared to be more effective than peripheral subcutaneous vaccination at generating tumor regression. This result was mediated by the migration of DCs to extracranial lymph nodes and resulted in systemic antitumor immunity against intracranial glioma cells. Furthermore, the activity of IL-12 or IL-18 appears to enhance the effectiveness of intratumoral injection of DCs. These results suggest that augmenting the local immune response with cytokines can further enhance the infiltration of T cells into the tumor parenchyma, thereby promoting the interaction of T cells with tumor-pulsed APCs.

Nevertheless, the early success of DCs as vehicles for future vaccine development has to be viewed in the context of potential obstacles. Namely, several problems remain to be solved in order to fully optimize this form of therapy. These include:

- Sources of DCs or precursor cells
- Methods of maturation
- Antigen-loading strategies
- Route of administration
- Role of administering additional cytokines and end points for clinical trials

To date, the major sources of DCs include direct isolation from peripheral blood or in vitro generation. A direct comparison of these DCs has not yet been made. Moreover, given the different immunological properties of immature versus mature DCs (antigen uptake and presentation, MHC expression, migratory capacity, expression of adhesion molecules or costimulatory activity), it will be important to determine which subset provides for optimal vaccine. Similarly, the elucidation of the best antigen source, (i.e., peptides, proteins, DNA, mRNA, cell lysates, apoptotic bodies and fusions) awaits further investigation. While most current trials employ subcutaneous or intratumoral immunization, the availability of other strategies, such as intravenous, intradermal or intranodal, intralymphatic, has not been compared, with or without the use of adjuvant cytokines. Finally, there are no established criteria for measuring the success of vaccine therapy, although an increase in overall survival is commonly used as a study endpoint. Perhaps a better determinant of success would be a delay in time to progression or recurrence, given that immunotherapeutic applications are most likely to be successful in clinical situations where the volume of residual disease is low.

Finally, the current literature provides ample evidence for successful use of viral vectors for gene therapy of cancer. For example, HSV has been used to deliver IL-4, IL-10 and IL-12

against experimental glioma [40-43]. The combination of HSV/IL-4 was shown to augment the oncolytic effect of HSV alone and improve the survival of mice with experimental brain tumors. Likewise, local expression of IL-12 via HSV was shown to increase T helper 1 response and significantly increase the survival of mice with intracranial tumors. The effects observed with HSV have been documented with adenoviral vectors for the delivery of IL-4, IL-12 and tumor necrosis factor (TNF)- α and vaccinia-mediated delivery of IL-2 and IL-12. In all instances, mice intracerebrally implanted with viral vectors producing IL-2, IL-4, IL-12 or TNF- α survived significantly longer than those implanted with noncytokine-secreting vectors. These results further confirm the use of local cytokine delivery against brain tumors and, when used in combination with oncolytic therapy, suggest a powerful means for targeted brain tumor therapy.

The development of novel biological therapies for primary brain cancer has been and continues to be intensely investigated. There is abundant evidence that tumor-specific immunity

References

- Parney IF, Farr-Jones MA, Kane K *et al.* Human autologous *in vitro* models of glioma immunogene therapy using B7-2, GM-CSF, and IL-12. *Can. J. Neurol. Sci.* 29(3), 267–275 (2002).
- 2 Parney IF, Petruk KC, Zhang C *et al.* Granulocyte-macrophage colonystimulating factor and B7-2 combination immunogene therapy in an allogeneic Hu-PBL-SCID/beige mouse-human glioblastoma multiforme model. *Hum. Gene Ther.* 8(9), 1073–1085 (1997).
- ³ Jean WC, Spellman SR, Wallenfriedman MA *et al.* Effects of combined granulocyte–macrophage colony-stimulating factor (GM-CSF), interleukin-2, and interleukin-12 based immunotherapy against intracranial glioma in the rat. *J. Neurooncol.* 66(1–2), 39–49 (2004).
- 4 Wallenfriedman MA, Conrad JA, DelaBarre L *et al.* Effects of continuous localized infusion of granulocyte-macrophage colonystimulating factor and inoculations of irradiated glioma cells on tumor regression. *J. Neurosurg.* 90(6), 1064–1071 (1999).
- 5 Herrlinger U, Kramm CM, Johnston KM et al. Vaccination for experimental gliomas using GM-CSF-transduced glioma cells. Cancer Gene Ther. 4(6), 345–352 (1997).
- 6 Lefranc F, Cool V, Velu T *et al.* Granulocyte macrophage-colony stimulating factor gene transfer to induce a protective anti-tumoral immune response against the 9L rat gliosarcoma model. *Int. J. Oncol.* 20(5), 1077–1085 (2002).

- 7 Sampson JH, Archer GE, Ashley DM et al. Subcutaneous vaccination with irradiated, cytokine-producing tumor cells stimulates CD8⁺ cell-mediated immunity against tumors located in the 'immunologically privileged' central nervous system. *Proc. Natl Acad. Sci. USA* 93(19), 10399–10404 (1996).
- 8 Wakimoto H, Abe J, Tsunoda R *et al.* Intensified antitumor immunity by a cancer vaccine that produces granulocyte-macrophage colonystimulating factor plus interleukin 4. *Cancer Res.* 56(8), 1828–1833 (1996).
- 9 Yu JS, Burwick JA, Dranoff G et al. Gene therapy for metastatic brain tumors by vaccination with granulocyte-macrophage colonystimulating factor-transduced tumor cells. *Hum. Gene Ther.* 8(9), 1065–1072 (1997).
- 10 Sobol RE, Fakhrai H, Shawler D *et al.* Interleukin-2 gene therapy in a patient with glioblastoma. *Gene Ther.* 2(2), 164–167 (1995).
- 11 Lichtor T, Glick RP. Cytokine immunogene therapy for treatment of brain tumors. *J. Neurooncol.* 65(3), 247–259 (2003).
- 12 Lichtor T, Glick RP, Tarlock K *et al.* Application of interleukin-2-secreting syngeneic/allogeneic fibroblasts in the treatment of primary and metastatic brain tumors. *Cancer Gene Ther.* 9(5), 464–469 (2002).
- 13 Glick RP, Lichtor T, Panchal R, Mahendra A, Cohen EP. Treatment with allogeneic interleukin-2 secreting

can be induced in both experimental models and patients. However, there is also increasing evidence that as the tumor grows, the ability of the immune system to overcome the cancer fades. On an intuitive level, this makes perfect sense. Historically, vaccines were developed to prevent disease. We are now proposing to cure cancer. As many immunologists probably recognize, the degree of T-cell priming as well as the number of antigen-specific T cells, is probably far below the level that is required to eliminate solid tumors. Targeted vaccines to low-burden disease as well as newer vaccine technologies administered alone or in combination with neoadjuvant disease will certainly optimize clinical results. The question is no longer where are we heading, rather, how will we get there? The important studies in the next few years will analyze immunological approaches to brain cancer not only via clinical response but also via the mechanism of action. Optimizing both arms will likely advance immunotherapy into clinical practice and make a significant impact on the lives of patients with malignant brain tumors.

fibroblasts protects against the development of malignant brain tumors. *J. Neurooncol.* 64(1–2), 139–146 (2003).

- 14 Ishikawa E, Tsuboi K, Takano S *et al.* Intratumoral injection of IL-2-activated NK cells enhances the antitumor effect of intradermally injected paraformaldehydefixed tumor vaccine in a rat intracranial brain tumor model. *Cancer Sci.* 95(1), 98–103 (2004).
- 15 Iwadate Y, Yamaura A, Sato Y *et al.* Induction of immunity in peripheral tissues combined with intracerebral transplantation of interleukin-2-producing cells eliminates established brain tumors. *Cancer Res.* 61(24), 8769–8774 (2001).
- 16 Dillman RO, Duma CM, Schiltz PM *et al.* Intracavitary placement of autologous lymphokine-activated killer (LAK) cells after resection of recurrent glioblastoma. *J. Immunother.* 27(5), 398–404 (2004).
- 17 Rhines LD, Sampath P, DiMeco F et al. Local immunotherapy with interleukin-2 delivered from biodegradable polymer microspheres combined with interstitial chemotherapy: a novel treatment for experimental malignant glioma. *Neurosurgery* 52(4), 872–879; (2003).
- 18 Okada H, Villa L, Attanucci J *et al.* Cytokine gene therapy of gliomas: effective induction of therapeutic immunity to intracranial tumors by peripheral immunization with interleukin-4 transduced glioma cells. *Gene Ther.* 8(15), 1157–1166 (2001).
- 19 Okada H, Giezeman-Smits KM, Tahara H et al. Effective cytokine gene therapy against an intracranial glioma using a

retrovirally transduced IL-4 plus HSVtk tumor vaccine. *Gene Ther.* 6(2), 219–226 (1999).

- 20 Okada H, Lieberman FS, Edington HD et al. Autologous glioma cell vaccine admixed with interleukin-4 gene transfected fibroblasts in the treatment of recurrent glioblastoma: preliminary observations in a patient with a favorable response to therapy. J. Neurooncol. 64(1–2), 13–20 (2003).
- 21 Iwadate Y, Namba H, Sakiyama S *et al.* Interleukin-12-mediated induction of systemic immunity in the periphery and recruitment of activated T cells into the brain produce limited antitumor effects compared with interleukin-2. *Int. J. Mol. Med.* 10(6), 741–747 (2002).
- 22 DiMeco F, Rhines LD, Hanes J et al. Paracrine delivery of IL-12 against intracranial 9L gliosarcoma in rats. J. Neurosurg. 92(3), 419–427 (2000).
- 23 Ehtesham M, Kabos P, Kabosova A *et al.* The use of interleukin 12-secreting neural stem cells for the treatment of intracranial glioma. *Cancer Res.* 62(20), 5657–5663 (2002).
- 24 Yang SY, Liu H, Zhang JN. Gene therapy of rat malignant gliomas using neural stem cells expressing IL-12. *DNA Cell Biol.* 23(6), 381–389 (2004).
- 25 Kishima H, Shimizu K, Miyao Y *et al.* Systemic interleukin 12 displays antitumour activity in the mouse central nervous system. *Br. J. Cancer* 78(4), 446–453 (1998).
- 26 Roy EJ, Gawlick U, Orr BA *et al.* IL-12 treatment of endogenously arising murine brain tumors. *J. Immunol.* 165(12), 7293–7299 (2000).
- 27 Insug O, Ku G, Ertl HC *et al.* A dendritic cell vaccine induces protective immunity to intracranial growth of glioma. *Anticancer Res.* 22(2A), 613–621 (2002).

- 28 Kikuchi T, Akasaki Y, Abe T *et al.* Vaccination of glioma patients with fusions of dendritic and glioma cells and recombinant human interleukin 12. *J. Immunother*, 27(6), 452–459 (2004).
- 29 Ni HT, Spellman SR, Jean WC *et al.* Immunization with dendritic cells pulsed with tumor extract increases survival of mice bearing intracranial gliomas. *J. Neurooncol.* 51(1), 1–9 (2001).
- ³⁰ Saito R, Mizuno M, Nakahara N *et al.* Vaccination with tumor cell lysate-pulsed dendritic cells augments the effect of IFN- β gene therapy for malignant glioma in an experimental mouse intracranial glioma. *Int. J. Cancer* 111(5), 777–782 (2004).
- 31 Caruso DA, Orme LM, Neale AM *et al.* Results of a Phase 1 study utilizing monocytederived dendritic cells pulsed with tumor RNA in children and young adults with brain cancer. *Neurooncology* 6(3), 236–246 (2004).
- 32 De Vleeschouwer S, Van Calenbergh F, Demaerel P *et al.* Transient local response and persistent tumor control in a child with recurrent malignant glioma: treatment with combination therapy including dendritic cell therapy. Case report. *J. Neurosurg. Spine* 100(5), 492–497 (2004).
- 33 Rutkowski S, De Vleeschouwer S, Kaempgen E *et al.* Surgery and adjuvant dendritic cell-based tumour vaccination for patients with relapsed malignant glioma, a feasibility study. *Br. J. Cancer* 91(9), 1656–1662 (2004).
- 34 Yamanaka R, Abe T, Yajima N *et al.* Vaccination of recurrent glioma patients with tumour lysate-pulsed dendritic cells elicits immune responses: results of a clinical Phase I/II trial. *Br. J. Cancer* 89(7), 1172–1179 (2003).
- 35 Yu JS, Liu G, Ying H *et al.* Vaccination with tumor lysate-pulsed dendritic cells elicits antigen-specific, cytotoxic T-cells in patients with malignant glioma. *Cancer Res* 64(14), 4973–4979 (2004).
- 36 Yu JS, Wheeler CJ, Zeltzer PM *et al.* Vaccination of malignant glioma patients with peptide-pulsed dendritic cells elicits systemic cytotoxicity and intracranial T-cell infiltration. *Cancer Res* 61(3), 842–847 (2001).

- 37 Ehtesham M, Kabos P, Gutierrez MA *et al.* Intratumoral dendritic cell vaccination elicits potent tumoricidal immunity against malignant glioma in rats. *J. Immunother.* 26(2), 107–116 (2003).
- 38 Kikuchi T, Akasaki Y, Abe T *et al.* Intratumoral injection of dendritic and irradiated glioma cells induces anti-tumor effects in a mouse brain tumor model. *Cancer Immunol. Immunother.* 51(8), 424–430 (2002).
- 39 Yamanaka R, Tsuchiya N, Yajima N et al. Induction of an antitumor immunological response by an intratumoral injection of dendritic cells pulsed with genetically engineered Semliki Forest virus to produce interleukin-18 combined with the systemic administration of interleukin-12. J. Neurosurg. 99(4), 746–753 (2003).
- 40 Andreansky S, He B, van Cott J *et al.* Treatment of intracranial gliomas in immunocompetent mice using herpes simplex viruses that express murine interleukins. *Gene Ther.* 5(1), 121–130 (1998).
- 41 Miyatake S, Martuza RL, Rabkin SD. Defective herpes simplex virus vectors expressing thymidine kinase for the treatment of malignant glioma. *Cancer Gene Ther.* 4(4), 222–228 (1997).
- 42 Parker JN, Gillespie GY, Love CE *et al.* Engineered herpes simplex virus expressing IL-12 in the treatment of experimental murine brain tumors. *Proc. Natl Acad. Sci. USA* 97(5), 2208–2213 (2000).
- 43 Hellums EK, Markert JM, Parker JN *et al.* Increased efficacy of an interleukin-12secreting herpes simplex virus in a syngeneic intracranial murine glioma model. *Neurooncology* 7(3), 213–224 (2005).

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