



## Effect of Chronic Metabolic Acidosis on Renal Growth and Renal Sodium Handling in Uninephrectomized Rats

L. F. Menegon, J. F. Figueiredo & J. A. R. Gontijo

To cite this article: L. F. Menegon, J. F. Figueiredo & J. A. R. Gontijo (1999) Effect of Chronic Metabolic Acidosis on Renal Growth and Renal Sodium Handling in Uninephrectomized Rats, Renal Failure, 21:1, 13-22, DOI: [10.3109/08860229909066966](https://doi.org/10.3109/08860229909066966)

To link to this article: <https://doi.org/10.3109/08860229909066966>



Published online: 07 Jul 2009.



Submit your article to this journal [↗](#)



Article views: 158



View related articles [↗](#)

## LABORATORY STUDY

# Effect of Chronic Metabolic Acidosis on Renal Growth and Renal Sodium Handling in Uninephrectomized Rats

---

L.F. Menegon,<sup>1</sup> J.F. Figueiredo,<sup>2</sup> and  
J.A.R. Gontijo<sup>1</sup>

<sup>1</sup>*Disciplinas de Medicina Interna e Nefrologia*

<sup>2</sup>*Laboratórios de Conservação de Órgãos e Balanço Hidro-Salino*

*Núcleo do Medicina e Cirurgia Experimental*

*Departamento de Clínica Médica*

*Faculdade de Ciências Médicas*

*Universidade Estadual de Campinas*

*Campinas, SP*

*Brasil*

## ABSTRACT

*Paucity studies have indicated that a systemic metabolic acidosis cause a decrease in salt and water reabsorption in the kidney. The following study was undertaken on male Wistar-Hannover rats (200–250 g) to investigate the effects of a chronic, NH<sub>4</sub>Cl-induced metabolic acidosis on the renal handling of Na<sup>+</sup> in sham-operated and uninephrectomized rats, by lithium clearance. The present study shows that chronic acidosis (blood pH, 7.16 ± 0.13) caused a sustained increase in renal fractional Na<sup>+</sup> excretion (267.9 ± 36.4%), accompanied by a rise in the fractional proximal (113.3 ± 3.6%) and post-proximal (179.7 ± 20.2%) Na<sup>+</sup> and fractional K<sup>+</sup> (163.4 ± 5.6%) excretions when compared to pair-fed rats. These differences occurred in spite of an unchanged creatinine clearance and Na<sup>+</sup> filtered load. On the other hand, a body growth impairment was observed in the acidotic (control, 258 ± 3.7 g versus acidotic, 232 ± 4.6 g) and pair-fed rats (225 ± 3.6 g), whereas there was significant enhance in the kidney weights in acidotic rats (1.73 ± 0.05 g) com-*

*Address correspondence to:* J.A.R. Gontijo, Departamento de Clínica Médica, Faculdade de Ciências Médicas, Universidade Estadual de Campinas, 13083-100 Campinas, SP, Brasil. Fax: 55-19-239-4107.

pared to other experimental groups (control,  $1.46 \pm 0.05$  g; pair-fed,  $1.4 \pm 0.05$  g). The renal growth indexes after metabolic acidosis  $\text{NH}_4\text{Cl}$ -induced did not show statistical difference at 1.5, 3.0 and 12 hours after uninephrectomy when were compared with pair-fed groups. However, from the fifth to tenth day after unilateral nephrectomy the renal growth index of acidotic group was significantly greater than pair-fed groups. Unilateral nephrectomy in acidotic animals caused a striking additional but transient increase in fractional renal sodium ( $\text{FENa}^+$ ) and potassium ( $\text{FEK}^+$ ) excretion from 1.5 to 3 hours post-surgery mainly associated with an enhanced post-proximal sodium excretion when compared to pair-fed uninephrectomized rats. By the fifth postoperative day the all functional values returned to baseline levels. This altered renal  $\text{Na}^+$  handling and  $\text{K}^+$  excretion may result from a reciprocal relationship between tubular metabolic pathway stimuli and ion transport. Further studies are required to investigate the acidosis involvement on functional kidney response.

**Key words:** Renal function; Renal sodium excretion; Lithium clearance; Metabolic acidosis.

## INTRODUCTION

There is a surprising lack of experimental data on the actual mechanisms of the metabolic acidosis-induced disturbance in sodium handling in the renal tubules. Paucity studies have indicated that a systemic metabolic acidosis causes a decrease in salt and water reabsorption in the kidney (1,2). Ammonium chloride-induced chronic metabolic acidosis has been shown to cause a decrease in hydro-saline reabsorption in dog kidney (3,4). Using free water clearance technique (5) as an index of proximal tubular function, the effect of ammonium chloride acidosis on renal ion balance appeared to be localized to the proximal tubule segments in the dog. On the other hand, during sustained metabolic acidosis in man and other animals show a progressive increase in capacity to excrete ammonia in the urine (1,4,6). This process may continue until practically the entire load of acid is excreted as the ammonium salt.

Chronic ammonium chloride induced metabolic acidosis has been found to be an inducing factor for renal growth in rats. The hypertrophic mechanism involved in this process is not entirely known and could be due to increased proteic synthesis or due reduced proteic catabolism in the tubule renal cells (7, 13). Nephronal hypertrophy, characterized by an increase in cell protein content and cell size, is predominantly accounted for by an elevated renal tubule mass (7). This hypertrophy occurs in several disorders such as diabetic nephropathy, the remnant kidney, and renal insufficiency (7).

After a reduction in the renal mass, the remaining kidney undergoes hypertrophy with an associated increment in functional capacity (7). In the remaining kidney, there is an increased glomerular filtration rate; enhanced reabsorption of sodium, bicarbonate and tubular fluid; and cell hypertrophy. The increase in transport capacity is seen in various nephron segments but primarily occurs in the proximal tubule (7,8).

Renal lithium clearance has been used as a noninvasive method to estimate the output of sodium and fluid by the proximal renal tubules, thus allowing the study of various factors which influence sodium handling in different tubule segments (9, 10).

The following study was undertaken to further investigate the effects of a chronic,  $\text{NH}_4\text{Cl}$ -induced metabolic acidosis on the renal growth and the tubule sodium handling, by lithium clearance, in sham-operated and uninephrectomized rats.

## MATERIAL AND METHODS

The experiments were performed on sixty, male Wistar-Hannover rats (200-250 g) allowed free access to water and normal rat chow. The general guidelines established by the Declaration of Helsinki (1964) for laboratory animals were followed throughout the study.

### Experimental Design

Metabolic acidosis was produced by substituting the drinking water for 0.25 M solution of  $\text{NH}_4\text{Cl}$  (Merck). All animals drank *ad libitum* but records of volume were kept, and only those rats which had drunk nearly equivalent amount of water or  $\text{NH}_4\text{Cl}$  were selected for use.  $\text{NH}_4\text{Cl}$ -treated rats were maintained on their respective regimens for 10 days. To simulate the hypophagia induced by  $\text{NH}_4\text{Cl}$  metabolic acidosis, the renal function was also performed in a subgroup of animals (pair-fed) submitted to food-restricted pattern similar to the average food intake observed in the experimental acidotic rats. On the 11th day the renal function of rats having both kidneys (sham-operated) and uninephrectomized rats was studied at intervals of 1.5 h, 3 h and 12 h after unilateral nephrectomy and then again after 5 and 10 days.

Fourteen hours before the renal test, 60mmol  $\text{LiCl}$ /100 g body weight was given by gavage. The rats were subsequently housed individually in metabolic cages with free access to tap water but no food. The experiment was performed at the same time in each group of control, pair-fed and acidotic animals. At 8:00 a.m., each animal received a load of tap water by gavage (5% of body weight), followed by a second load of the same volume one hour later. Twenty minutes after the second load, spontaneously voided urine was collected over a 2 h period. The voided urine passed through the funnel in the bottom of the cage into a graduated centrifuge tube. At the end of the experiment, blood samples were drawn by cardiac puncture and the kidney was immediately removed, decapsulated and weighed.

### Unilateral Nephrectomy

Ethyl ether was used to anesthetize the animals and a 1 cm incision was made immediately below the last left rib. The kidney was carefully removed, drained and weighed on a precise balance. The adrenal was kept in place intact. After certifying the absence of excessive bleeding, the incision was sutured.

### Biochemical Analysis

Plasma and urine sodium, potassium and lithium concentrations were measured by

flame photometry, while the creatinine concentrations were determined spectrophotometrically by the alkaline picrate method (11).

### Statistics and Calculations

The results are reported as means  $\pm$  SEM per 100 g body weight. Renal clearances (C) was calculated by a standard formula ( $C = UV/P$ ) using the plasma creatinine and lithium levels for each period. Creatinine clearance was used to estimate glomerular filtration rate (GFR) and lithium clearance ( $CLi^+$ ) was used to assess proximal tubule output. Fractional sodium ( $FENa^+$ ) and potassium ( $FEK^+$ ) excretion was calculated as  $CNa^+/CCr$  and  $CK^+/CCr$  respectively, where  $CNa^+$  and  $CK^+$  is ion clearance and  $CCr$  is creatinine clearance. The fractional proximal ( $PFENa^+$ ) and post-proximal ( $PPFENa^+$ ) sodium excretion were calculated as  $CLi/CCr \times 100$  and  $CNa^+/CLi^+ \times 100$ , respectively (10). Changes in fractional excretion were estimated using the control and pair-fed values. The renal growth index was calculated as the percentage variant of renal weight of the same rat multiplied by 100. Statistical analysis of the data was performed using one-way analysis of variance for repeated measurements. When the results were significant, the Bonferroni's contrast test was used to determine the extent of the differences. A  $p$  value  $< 0.05$  was considered to indicate significance.

## RESULTS

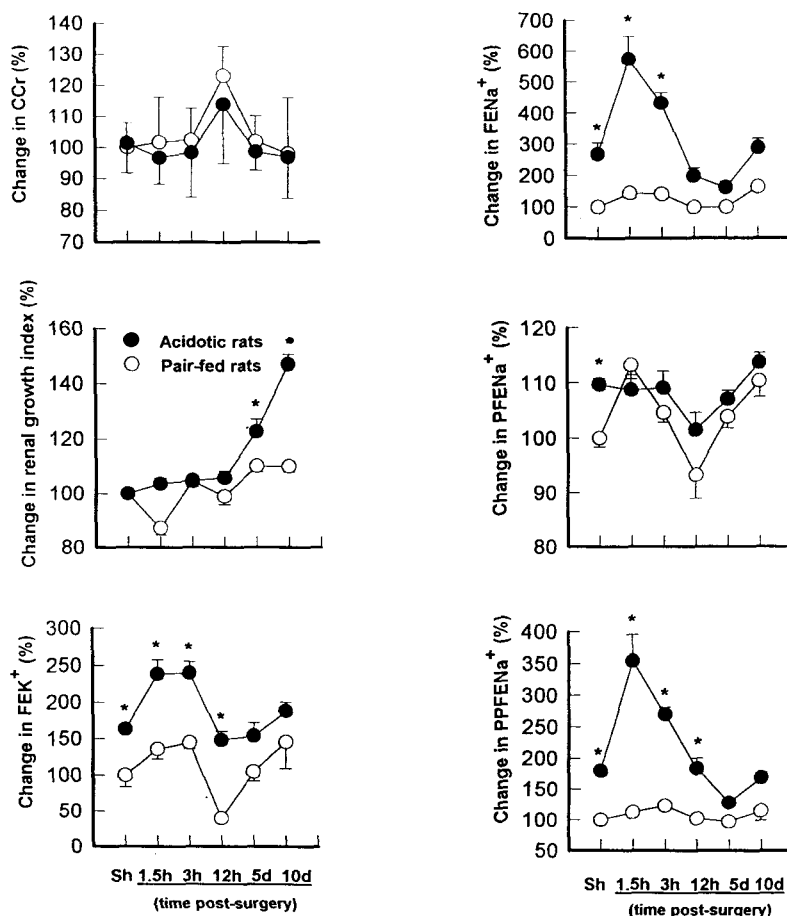
Groups of ten animals (acidotic control (sham-operated); acidotic uninephrectomized and pair-fed groups) were studied. Metabolic acidosis was confirmed by a blood pH of  $7.16 \pm 0.13$ . All rats survived and were clinically healthy up to tenth day of the study. As show in Table 1, body growth impairment was observed in the acidotic sham-operated rats after ten days of  $NH_4Cl$  or pair-fed intake. In contrast, the renal growth index was significantly less for the pair-fed compared to acidotic sham-operated group. There was also a significant difference between serum potassium, but not in sodium and lithium levels in acidotic sham-operated compared to pair-fed rats (Table 1) The renal growth indexes after metabolic acidosis  $NH_4Cl$ -induced did not shown statistical difference at 1.5, 3.0 and 12 hours after uninephrectomy when were compared with pair-fed groups. However, from the fifth to tenth day after unilateral nephrectomy the renal growth index of acidotic group was significantly greater than pair-fed groups (Figure 1). The data from the renal function studies in the experimental groups ten days after metabolic acidosis  $NH_4Cl$ -induced is shown in Figure 1. Metabolic acidosis caused a sustained increase in basal renal fractional sodium and potassium excretion, accompanied by a rise in the fractional proximal and post-proximal sodium excretion when sham-operated were compared to pair-fed rats (Figure 1). These differences occurred in spite of an unchanged creatinine clearance (Figure 1) and sodium filtered load. The urinary flow rates throughout the renal tubule sodium handling studies were not significantly different between unilateral nephrectomized rats and sham-operated animals. Unilateral nephrectomy in acidotic animals caused a striking additional but transient increase in fractional renal sodium ( $FENa^+$ ) and potassium ( $FEK^+$ ) excretion from 1.5 to 3 hours post-surgery. By the 1.5 to 3 hours post-uninephrectomy the fractional urinary sodium and potassium excretion in acidotic rats was associated with an enhanced post-proximal

Table 1

*Effect of Metabolic Acidosis Induced by  $\text{NH}_4\text{Cl}$  Administration ( $\text{NH}_4$ ) in Body Weight, Renal Weight and Serum Sodium, Potassium and Lithium Levels Compared to Sham-Operated (Sham) and Pair-Fed (PF) Rats. Data Are Reported as Means  $\pm$  SEM for 10 Rats Per Group.*

Groups	Body Weight (g)	Renal Weight (g)	Liquid Intake (mL)	$\text{Na}^+$ (mM)	$\text{K}^+$ (mM)	$\text{Li}^+$ (mM)
Sham	$258 \pm 3.7$	$1.46 \pm 0.05$	$34.9 \pm 0.9$	$143 \pm 0.9$	$3.7 \pm 0.1$	$100 \pm 5.0$
PF	$225 \pm 3.6$	$1.40 \pm 0.05$	$34.8 \pm 0.5$	$142 \pm 1.2$	$3.5 \pm 0.1$	$113 \pm 6.0$
$\text{NH}_4$	$232 \pm 4.3^a$	$1.73 \pm 0.05^b$	$35.7 \pm 0.7$	$143 \pm 0.4$	$4.4 \pm 0.1^{ab}$	$109 \pm 8.0$

<sup>a,b</sup>  $p < 0.01$  compared to Co or PF respectively (ANOVA and Bonferroni's contrast test).



**Figure 1.** Effect of metabolic acidosis in uninephrectomized rats on renal creatinine clearance (CCr), renal growth index, fractional sodium (FENa) and potassium (FEK) excretion, and fractional proximal (PFENa) and post-proximal (PPFENa) tubule sodium excretion compared to acidotic sham-operated (Sh) and uninephrectomized pair-fed (PF) rats. The data are reported as percent of sham-operated or pair-fed values and represented as the mean  $\pm$  SEM for 10 rats per group. \*  $P < 0.05$  compared to the control or pair-fed animals (ANOVA and Bonferroni contrast test).

sodium excretion when compared to pair-fed uninephrectomized rats (Figure 1). At this time, the increased urinary excretion of ions occurred in spite of an unchanged creatinine clearance. However, twelve hours after uninephrectomy there was a statistically significant increase of creatinine clearance to a similar extent in acidotic and pair-fed rats. By the fifth postoperative day the all-functional values returned to baseline levels. The high urinary sodium excretion in acidotic sham-operated animals was not enhanced by unilateral nephrectomy. On the other hand, at 1.5 to 3 postoperative hours the fractional proximal sodium reabsorption in uninephrectomized pair-fed group was significantly depressed (Figure 1).

## DISCUSSION

These findings are in agreement with previous studies (2,3,5), which have demonstrated decreased sodium transport in the presence of reduced blood pH. Absolute sodium excretion and fractional excretion of sodium were increased in sham-operated acidotic animals, despite similarities in the filtered load of sodium with pair-fed rats. Sartorius, Roemmoelt and Pitts (12), showed the humans, administered an oral ammonium chloride load, demonstrate a natriuresis in the early phases of the induced acidosis. Studies in rats and dogs undergoing a metabolic acidosis indicate that there is a decrease in renal tubular reabsorption of salt and water (2-4), and micropuncture studies localize the site of depression of sodium reabsorption to the proximal tubule (5,6). The relationship between sodium reabsorption and blood bicarbonate levels has been the subject of study (13). This study demonstrated that isotonic sodium and water reabsorption in the rat proximal tubules was correlated to the bicarbonate concentration in the peritubular capillaries. A depression of pericapillary bicarbonate level was correlated to a reduction in the proximal tubular reabsorption of salt and fluid. This observation may explain at least in part, our lithium clearance data in sham-operated rats, undergoing a metabolic acidosis, indicating that there is a decrease in tubular sodium reabsorption in the proximal tubule segments (2,5). The present data shows also a proximal sodium rejection followed by an enhanced distal fractional sodium excretion in the absence of change in creatinine clearance. In our study, the metabolic acidosis in sham-operated rats induced by  $\text{NH}_4\text{Cl}$  feeding caused an absolute increase in kidney weight. The data are consistent with *in vitro* studies of proximal tubule cells (14), where cells hypertrophy were accompanied by a marked decline in protein breakdown but not in cells protein synthesis after application of  $\text{NH}_4\text{Cl}$ . Also, the  $\text{Na}^+/\text{H}^+$  exchanger,  $\text{H}^+/\text{K}^+$  ATPase and  $\text{H}^+$ -ATPase stimulation has been implicated as an important initiating signal for subsequent events leading to cell proliferation (21-24). Fine et al. (20), demonstrated that in renal proximal tubular cells *in vitro*, activation of the  $\text{Na}^+/\text{H}^+$  antiporter correlated better with cell hypertrophy than with proliferation. In contrast with our observations, acidification *in vitro* has failed to produce cells hypertrophy when not produced by ammonium chloride (15,16). It has become apparent only when metabolic acidosis cause an effect of increased ammonia production rather than of the acidosis per se (17).

In agreement with several previous studies (8,18,19), we found that fractional proximal reabsorption was reduced acutely after uninephrectomy in pair-fed rats. The cause of this glomerulo-tubular imbalance remains unknown, but it appears to be short-lived, within 5 days fractional proximal reabsorption had increased so that fractional reabsorption had returned to a value similar to that in basal sham-operated animals. It seems likely that this increased rate of ion reabsorption is the *in vivo* parallel of the well-documented changes in ultrastructure and function in proximal tubular cells, including increased activity of the  $\text{Na}^+/\text{H}^+$  antiport and the  $\text{Na}^+/\text{K}^+$  pump, which have been described *in vitro* (20-24). The increase in transport capacity is seen in various nephron segments but primarily occurs in the proximal tubule (7,20). The activity of  $\text{Na}^+/\text{H}^+$  exchanger in the remaining kidney was found to be increased following 24 h after uninephrectomy in rats (23,24). Authors have demonstrated that antiport activity measured in cortical brush-border membrane vesicles rapidly increases within the 30 minutes after contralateral uninephrectomy (24). However, the signal that triggered antiport activation in the remaining kidney still also unknown. In the present study, the accentuated



fall in the proximal sodium reabsorption after 10-day treatment with ammonium chloride may have blunted in acidotic rats the natriuretic response to uninephrectomy observed in pair-fed animals. These results could be explained by the maximal  $\text{Na}^+/\text{H}^+$  exchanger expression or activity after acidotic stimuli. The present study also emphasizes the importance of poor nutrition as an inhibitor of linear growth. Our data show a significant inhibition of renal growth in uninephrectomized pair-fed as compared to uninephrectomized acidotic rats (Figure 1). This corroborates earlier data by Philipps et al (25) which showed that calorie limitation in neonatal rats reduced IGF-I and II in serum, liver and brain.

Previous studies have been shown that alterations in acid-base balance modify renal gluconeogenesis. Metabolic acidosis stimulates it in a variety of preparations by increase of phosphoenolpyruvate carboxykinase mRNA cloning and enzyme activity (26,27). There are many indications that gluconeogenesis and the reabsorption of  $\text{Na}^+$  are reciprocally related. Investigation has demonstrated that American opossum kidney (OK cells), respond to the stimulus of acidosis with increased glutamine metabolism and ammonium formation (28). Previous studies using OK cells have demonstrated that acidosis decrease the activity of  $\text{Na}^+/\text{H}^+$  exchanger thereby increasing intracellular  $\text{H}^+$  (29). Isolated proximal tubules studies showed that enhanced glutamine metabolism and ammonia production are linked to increased gluconeogenesis (30). On the other hand, maneuvers that inhibit of  $\text{Na}^+/\text{K}^+$  ATPase and thereby sodium tubule transport by the kidney stimulate gluconeogenesis (31,32). Thus, as metabolic acidosis may result in a lower filtered load of bicarbonate and consequently less bicarbonate is reabsorbed; and a decreased  $\text{Na}^+/\text{H}^+$  antiporter activity associated with stimulated gluconeogenesis. These features take together, it should lead to less dissipation of the electrochemical gradient for  $\text{Na}^+$  resulting in a decreased reabsorption of  $\text{Na}^+$  so as observed in present study. Additionally, the present findings suggest that in energy-requiring processes, renal growth, sodium transport and gluconeogenesis, may compete for energy availability in the nephron tubule, and could explain the striking natriuresis observed in ours study. Our results indicate that a transient increase in ion delivery out of the proximal tubule is a constant feature of the response of the remaining kidney to uninephrectomy in the rat, but that the underlying changes in proximal tubular function vary with the time elapsed and systemic acid-base status. Therefore, an increased sodium reabsorption in the proximal and distal nephrons can compensate partially the early, altered renal sodium handling in acidotic rats. Under our experimental conditions instead of previous studies (4,33), the fractional potassium excretion was also increased during acidosis. Many factors have been proposed as being important influences on the renal excretion of potassium including blood pH, potential across luminal membrane, sodium delivery to the distal tubule and urinary flow rate (34). Finally, the kaliuresis of uninephrectomized rats can be explained as a consequence of a markedly increased rate of potassium secretion into the accessible region of the distal tubule, mainly in acidotic rats.

While the present study provides fresh insights into the response of the renal  $\text{Na}^+$  excretion and chronic metabolic acidosis, it should be emphasized that it was conducted on whole-kidney function in metabolic cages using unanesthetized, unrestrained rats. In conclusion, the present study demonstrated that chronic metabolic acidosis in rats induces a sustained increase of renal weight and ion excretion. This altered renal  $\text{Na}^+$  handling and  $\text{K}^+$  excretion may result from a reciprocal relationship between tubular metabolic pathway stimuli and ion transport. Further studies are required to investigate the acidosis involvement on functional kidney response.

## REFERENCES

1. Lemmon EJ, Piering WF: A comparison of the effects of glucose ingestion and  $\text{NH}_4\text{Cl}$  acidosis on urinary calcium and magnesium excretion in man. *Journal of Clinical Investigation* 49:1458-1465, 1970.
2. Levine DZ, Nash L: Effect of chronic  $\text{NH}_4\text{Cl}$  acidosis on proximal tubular  $\text{H}_2\text{O}$  and  $\text{HCO}_3$  reabsorption. *American Journal of Physiology* 225: 380-384, 1973.
3. Safirstein R, Glassman VP, DiScala VA: Effects of  $\text{NH}_4\text{Cl}$  induced metabolic acidosis on salt and water reabsorption in dog kidney. *American Journal of Physiology* 225: 805-809, 1973.
4. Glassman VP, Safirstein R, DiScala VA: Effects of metabolic acidosis on proximal tubule ion reabsorption in dog kidney. *American Journal of Physiology* 227:759-765, 1974.
5. Stein RM, Abramson RG, Kahn T, Levitt MF: Effect of hypotonic saline loading in hydrated dog: evidence for a saline-induced limit on distal tubular sodium transport. *Journal of Clinical Investigation* 46:1205-1214, 1967.
6. Stein JH, Rector, Jr FC, Seldin DW: The effect of acute metabolic acidosis on proximal tubular sodium reabsorption in the rat. *Journal of Clinical Investigation* 47: R277, 1968.
7. Fine L: The biology of renal hypertrophy. *Kidney International* 29:619-634, 1986.
8. Shirley DG, Walter SJ: Acute and chronic changes in renal function following unilateral nephrectomy. *Kidney Int* 40:62-68, 1991.
9. Thomsen K, Shirley DG: The validity of lithium clearance as an index of sodium and water delivery from the proximal tubules. *Nephron* 77:125-138, 1997.
10. Quadros MR, Gontijo JAR, Figueiredo JF: Renal tubular sodium handling determined by lithium clearance in partially hepatectomized rats. *Brazilian Journal of Medical and Biological Research* 29: 1077-1083, 1996.
11. Brod J, Sirota JH: The renal clearance of endogenous creatinine in man. *Journal Clinical Investigation* 27: 645-651, 1948.
12. Sartorius OW, Roemmelt JC, Pitts RF: The renal regulation of acid-base balance in man. IV. The nature of the renal compensations in ammonium chloride acidosis. *Journal of Clinical Investigation* 28:423-439, 1949.
13. Herbert CS, Martinez-Maldonado M, Suki WN: Relation of bicarbonate to sodium reabsorption in dog kidney. *American Journal of Physiology* 222:1014-1020, 1972.
14. Ling H, Vamvakas S, Gekle M, Schaefer L, Teschner M, Shaefer RM, Heidland A: Role of lysosomal cathepsin activities in cell hypertrophy induced by  $\text{NH}_4\text{Cl}$  in cultured renal proximal tubule cells. *Journal of American Society of Nephrology* 7:73-80, 1996.
15. Nash KA, Hostetter MK, Hostetter TH: Increased ammoniogenesis as a determinant of progressive renal injury. *American Journal of Kidney Disease* 17: 654-657, 1991.
16. Jurkowitz CT, England BK, Kurtz I: Influence of ammonia and pH on protein and amino acids metabolism in LLC-PK1 cells. *Kidney International* 42:595-601, 1992.
17. Kurtz I: Role of ammonia in the induction of renal hypertrophy. *American Journal of Kidney Disease* 17: 650-653, 1991.
18. Dirks JH, Wong NLM: Acute functional adaptation to nephron loss: micropuncture studies. *Yale J Biol Med* 51: 255-263, 1978.
19. Diezi J, Michoud-Hausel P, Nicolas-Buxcel N: Studies on possible mechanisms of early functional compensatory adaptation in remaining kidney. *Yale J Biol Med* 51:265-270, 1978.
20. Fine LG, Norman J: Cellular events in renal hypertrophy. *Ann Rev Physiol* 51:19-32, 1989.
21. Elam-ong S, Kurtzman NA, Sabatini S: Renal ATPases twenty-four hours after uninephrectomy: The role of IGF-1. *Miner Electrolyte Metab* 22: 234-241, 1996.
22. Mackovic-Basic M, Fan R, Kurtz I: Denervation inhibits early increase in  $\text{Na}^+\text{-H}^+$  exchange after uninephrectomy but does not suppress hypertrophy. *Am J Physiol* 263:F328-F334, 1992.
23. Harris RC, Seifter JL, Brenner BM: Adaptation of  $\text{Na}^+\text{-K}^+$  exchange in renal microvillus membrane vesicles. Role of dietary protein and uninephrectomy. *J Clin Invest* 74: 1979-1987, 1984.
24. Salihagic A, Mackovic M, Banfic H, Sabalic I: Short-term and long-term stimulation of  $\text{Na}^+\text{-H}^+$  exchange in cortical brush-border membranes during compensatory growth of the rat kidney. *Pfluegers Arch* 413: 190-196, 1988.
25. Philipps A, Drakenberg K, Percon B, Sjogren B, Eklof AC, Hall K, Sara V: The effects of altered nutritional status upon insulin-like growth factors and their binding proteins in neonatal rats. *Pediatr Res* 26:128-134, 1989.
26. Parry DM, Brosnan JT: Renal phosphoenolpyruvate carboxykinase during perturbation of acid-base homeostasis in rats. Immunocytochemical studies. *Canadian Journal of Biochemistry* 58:1298-1301, 1980.
27. Ilyedjian PB, Jacob MM: Glucocorticoid-dependent induction of mRNA coding for phosphoenolpyruvate carboxykinase (GTP) in rat kidney. Its inhibition by cycloheximide. *European Journal of Biochemistry* 111:89-98, 1980.

28. Nissim I, States B, Nissim I, Lin Z-P, Yudkoff M: Hormonal regulation of glutamine metabolism by OK cells. *Kidney International* 47: 96–105, 1995.
29. Miller RT, Pollack AS: Modification of the internal pH sensitivity of Na/H antiporter by parathyroid hormone in cultured renal cell lines. *Journal of Biological Chemistry* 262: 9115–9120, 1987.
30. Nissim I, Nissim I, Yudkoff M: Adaptation of renal tricarboxylic acid cycle metabolism to various acid-base states. *Mineral and Electrolyte Metabolism* 17:21–31, 1991.
31. Friedrichs D, Schoner W: Stimulation of renal gluconeogenesis by inhibition of the sodium pump. *Biochemistry Biophysical Acta* 304:142–160, 1973.
32. McGeoch J, Falconer-Smith J, Ledingham J, Ross BD: Inhibition of active Na<sub>2</sub>K ATPase by myeloma protein. *Lancet* ii:17–18, 1978.
33. Malnic G, DeMello Aires M, Giebisch G: Potassium transport across renal distal tubules during acid-base disturbances. *American Journal of Physiology* 221:1192–1208, 1971.
34. Wright FS, Giebisch G: Regulation of potassium excretion. In: *The Kidney Physiology and Pathophysiology*. Raven Press Ltd, New York, 1992, pp 2209–2247.