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SHORT COMMUNICATION

Using C₆₀ fullerenes for photodynamic inactivation of mosquito iridescent viruses

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Abstract

This article describes the photodynamic inactivation of mosquito iridescent virus (MIV) *Aedes flavescens* in the presence of water-soluble C₆₀ fullerenes. It has been observed that the photodynamic inactivation of MIV for about 1 h reduces the infectious titre of the virus in large wax-moth larvae *Galleria mellonella* to 4.5 lg ID₅₀/mL. The influence of the C₆₀ concentration on its anti-viral activity was tested in the concentration range from 1 to 0.001 mg/mL. It has been found that C₆₀ is able to inactivate the iridovirus even in low concentrations. Consequently, the findings of this work suggest that photoexcited C₆₀ fullerenes can be successfully used for the inactivation of iridoviruses in biological systems.

Keywords: Water-soluble C60 fullerenes, mosquito iridescent virus, photodynamic inactivation

Introduction

Fullerenes belong to the third natural allotropic form of carbon. C₆₀ fullerene is the most studied member of fullerene family. It is a highly stable icosahedral molecule (with size of ~0.7 nm) consisting of 60 carbon atoms¹. In recent years, much attention was paid to a comprehensive investigation of C₆₀ influence in biological systems *in vitro* and *in vivo*^{2,3}. It was shown that the addition of fullerenes in some gel-like medicinal forms may significantly increase their wound healing properties⁴. Fullerenes and their derivatives are able to penetrate through cell membranes⁵ and they have strong anti-oxidant⁶, anti-viral^{3,7,8} as well as anti-microbial^{9–11} properties. It is also interesting that under irradiation, C₆₀ can be activated and reactive oxygen species are generated, which can cause destruction of nucleic acids, proteins and lipids^{3,12,13}. There are several reports in literature regarding studies on photodynamic inactivation of viruses from *Orthomyxoviridae*, *Rhabdoviridae*, *Togaviridae* and *Leviviridae* families. It has been reported that the viruses lose their infectivity by more than 7 lg ID₅₀ during a few hours of photoinactivation with C₆₀

fullerenes^{14–16}. A general concept of applying fullerenes in the photodynamic therapy is discussed more detail in paper¹⁷.

Iridoviruses are large DNA viruses (with sizes of ~180–200 nm) that can be replicated in the cytoplasm of infected cells. Iridoviruses have been found to infect invertebrates and poikilothermic vertebrates, including amphibians, reptiles and fishes¹⁸. Iridoviruses that infect mosquitoes and midges belong to the genus *Chloriridovirus*¹⁹. Mosquito iridescent viruses (MIVs) are widely distributed in natural reservoirs and play role as regulators of the insect quantity.

Iridoviruses not only infect mosquitoes and other insects but also cause mass mortality of fish that leads to significant economic losses in industrial aquaculture. Consequently, the isolation of these emerging iridoviruses is closely associated with an increase of the world aquaculture production rates²⁰.

Thus, the purpose of this work was to study the effect of photoactivated C₆₀ on infectious titre of iridoviruses. For this study, virions of MIV, isolated from the mosquito larvae *Aedes flavescens* in natural reservoirs of the Kyiv region (Ukraine) were used.

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Materials and methods

Highly stable water colloid suspension of C₆₀ fullerenes (maximum concentration 1 mg/mL) was prepared by transfer of C₆₀ (purity >99.5%) from toluene to water using ultrasound sonication²¹.

It is important to note that the considered water solution of C₆₀ fullerenes does not show a cytotoxic effect at concentrations below 1 mg/mL²².

For the characterisation of C₆₀ films deposited from water solutions (C₆₀ concentration in water was 1 mg/mL) on Au(111) surface, scanning tunnelling microscopy (STM; NT-MDT Moscow, Russia) images using a low-current scanning head were taken. The Pt-Ir (80:20) tip was mechanically cut from a wire (0.25 mm in diameter). Typical imaging conditions were in a range from 0.01 to 0.1 nA and from 0.1 to 0.8 V. Reconstructed Au(111) substrates of 150-nm thickness were prepared by vacuum deposition onto a freshly cleaved mica surfaces heated at ~600 K followed by a careful annealing in a gas flame (propane). During ~10 min after flaming, the substrate revealed reconstruction lines in air. Exposition in air during 1-h lifted reconstruction. A droplet of the water solution of C₆₀ fullerenes was deposited on these gold substrates.

MIV *Ae. flavescens* was propagated in multi-host wax-moth larvae *G. mellonella*. The wax-moth larvae were infected by intraperitoneal injection and incubated at room temperature (20–22°C).

Fourteen days after injection of the MIV *Ae. flavescens*, it was purified from the infected wax-moth larvae *G. mellonella* by the differential centrifugation method. Briefly, after homogenisation of infected larvae *G. mellonella*, cell debris was separated by centrifugation at 3000g for 5 min at 10°C. The pellet was discarded and the supernatant centrifuged in a ultracentrifuge (Beckman L5-50B in a rotor SW-40) for 40 min at 70,500g at 4°C. A characteristic blue pellet confirmed the presence of the mosquito iridescent virions. The virus pellet was suspended in TNE (50-mM Tris-HCl, 150-mM NaCl, 1-mM disodium ethylene diaminetetraacetic acid [EDTA], pH 7.5) and centrifuged at 1100g for 5 min at 10°C. The suspension was layered onto a 10–50% (w/w) linear sucrose gradient in TNE. After centrifugation at 70,500g for 40 min at 4°C, one visible band near the bottom of the tube was collected, diluted in fresh TNE and centrifuged at 70,500g for 40 min at 4°C. The virus pellet was placed into TNE at a final protein concentration of 1.7 mg/mL. The infectious titre of MIV *Ae. flavescens* in the wax-moth larvae *G. mellonella* was determined by the Reed and Munich method²³. The infectious titre of virus in larvae *G. mellonella* was 11.2 lg ID₅₀/mL.

For the photodynamic inactivation of MIV, a 400-W halogen lamp (intensity of light radiation was 10²⁰ photons/m²s) was used.

For the experiments, various solutions with constant concentration of virus (0.17 mg/mL) and different concentrations of C₆₀ (1, 0.1, 0.01 and 0.001 mg/mL) were used. The photodynamic inactivation of MIV was

performed at 400–850-nm wavelengths in a glass tube. The distance between the source of light and virus was 10 cm. The exposure time was 2.5, 10, 30 and 60 min. The MIV without C₆₀ fullerenes and MIV with C₆₀ fullerenes but without photodynamic inactivation were used as controls.

After exposure, the material from all variants was diluted in the sterile distilled water in an interval of 10⁻¹ to 10⁻¹². The wax-moth larvae *G. mellonella* were infected by the material from every dilution. The infected larvae were incubated at temperature 20–22°C. Fourteen days after injection, the titers of MIV were determined by the methods of blue pellets.

Results and discussion

The STM images of a submonolayer C₆₀ film deposited from water solution on Au(111) surface are shown in Figure 1. An almost random arrangement of C₆₀ clusters (with sizes up to ~2.8 nm) (Figure 1A) can be seen. It is well known that C₆₀ molecules form highly ordered hexagonal structures on reconstructed Au(111) in vacuum²⁴. The absence of such ordering in our case can be attributed to the degradation in water solution. However, nucleation of islands occurs along the preferential direction <112> (see Figure 1A). Despite of the high mobility of C₆₀ molecules on Au(111) at room temperature, we were able to observe single C₆₀ molecules (Figure 1B). The immobilisation of the C₆₀ fullerene is due to the lifting of the reconstruction by water.

The above-obtained results are in a good agreement with previously predicted theoretical calculations^{25,26}, which show that the water colloid solution of C₆₀ contains single C₆₀ molecules, their clusters and crystalline phase (with sizes of ~0.7–4 nm in dependence of C₆₀ concentration in water) in the hydrated state. Analysis of the UV/VIS spectroscopic data²¹ showed that C₆₀ fullerenes in water are characterised by two intense broad absorption bands with maximums at 265 and 345 nm and also by narrow spectral lines at 450 and 622 nm.

The most interesting and important finding of this study is possibility for the photodynamic inactivation of MIV *Ae. flavescens* by using an aqueous solution of C₆₀ (concentration 1 mg/mL). It has been found that after a treatment of about 2.5 min, the infectious titre of virus can be decreased by 2 lg ID₅₀/mL units (Figure 2). In the control group, the infectious titre of MIV was 11.2 lg ID₅₀/mL. The infectious titre of MIV in the *G. mellonella* after handling for about 2.5 min was decreased to 9.0 lg ID₅₀/mL. Further handling for 10 and 30 min leads to decrease of the infectious titre of the virus by 2.93 and 3.88 lg ID₅₀/mL units, respectively. Interestingly, the indexes of infectious titre of MIV after the photodynamic inactivation for 1 h were not significantly different from those obtained from photodynamic inactivation for 30 min. Infectious titre of MIV *Ae. flavescens* after the photodynamic inactivation during 1 h was 7.15 lg ID₅₀/mL.

Our results showed that after photodynamic inactivation for 30 min, C₆₀ in a concentration of 0.1 mg/mL

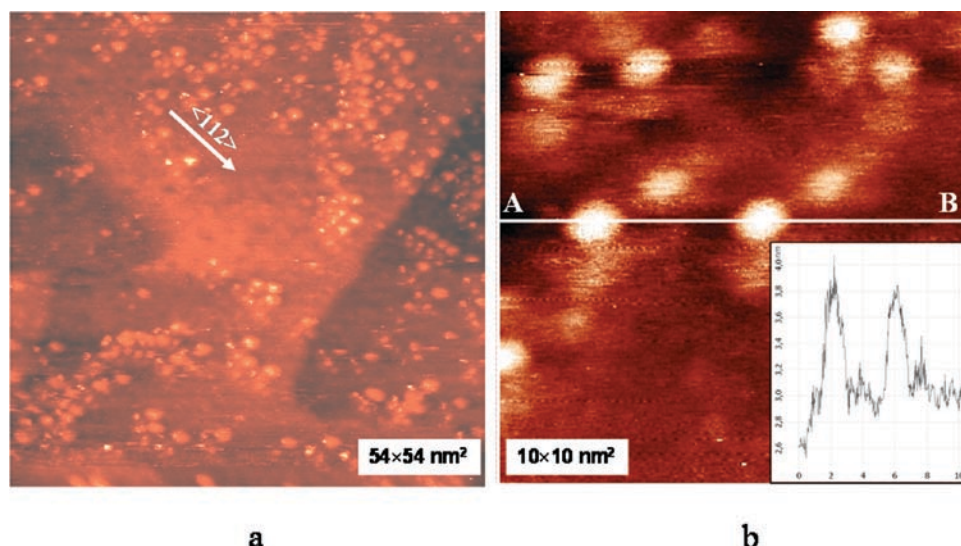


Figure 1. (A) Large-scale scanning tunnelling microscopy image of submonolayer C₆₀ fullerene film deposited from water solution (C₆₀ fullerene concentration in water was 1 mg/mL) on Au(111) surface. Some C₆₀ fullerene clusters are aligned along preferential direction. (B) Single C₆₀ fullerenes. Lateral size is increased because of shape of the tip. Inset: Cross-section along line AB. Scanning parameters: It=40 pA, Ut=0.7 V.

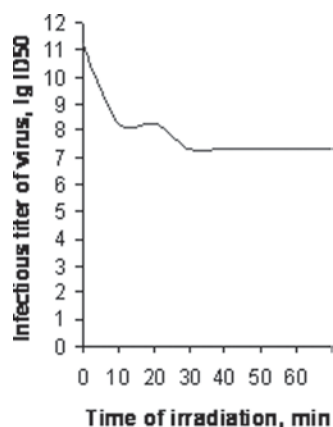


Figure 2. Photodynamic inactivation of mosquito iridescent virus *Ae. flavescens* by C₆₀ fullerenes (maximum concentration 1 mg/mL).

reduced the infectious titre of MIV *Ae. flavescens* by 4.0 lg ID₅₀/mL units. In a concentration of 0.01 mg/mL, C₆₀ reduced the infectious titre of the MIV by 4.5 lg ID₅₀/mL units. The use of C₆₀ in a concentration of 0.001 mg/mL actually did not influence the infectious titre of MIV *Ae. flavescens*. The infectious titre of MIV *Ae. flavescens* was 10.7 lg ID₅₀/mL, that is only 0.5 lg ID₅₀/mL less than in the control group (Table 1). The presence of C₆₀ in a viral suspension, but without the photodynamic inactivation, as well as irradiation of viral suspension in the absence of C₆₀ did not influence the infectious titre of MIV.

It is important to note that the effectiveness of reducing the infectious titre of MIV on the C₆₀ concentration and exposure time significantly related to fullerene aggregation in water²⁶: formation of fullerene clusters (aggregates) with single molecules leads to a significant restructuring of their electronic structure, which determines the mechanism of their photoactivation¹⁷.

Table 1. Photodynamic inactivation of mosquito iridoviruses *Ae. flavescens* by C₆₀.

Time of irradiation, min	30				
Concentration of C ₆₀ fullerenes, mg/mL	1	0.1	0.01	0.001	Control
Infectious titre of virus, lg ID ₅₀ /mL	7.32	7.2	6.7	10.7	11.2

Conclusion

The photodynamic inactivation of viruses by using C₆₀ and their derivatives can be considered as a promising treatment, and thus, it can be successfully applied in medicine. Recently, a lot of works involved in investigation of anti-viral and anti-bacterial properties of C₆₀ were reported in the literature^{9,27}. In spite of the fact that the most of the investigations were involved in interactions of fullerenes with either the human immunodeficiency virus²⁸ or hepatitis C virus²⁹, in this work we studied for first time the interaction of C₆₀ and MIV *Ae. flavescens*. The obtained results demonstrated the anti-viral properties of C₆₀ fullerenes during photoactivation. Namely, during the photodynamic inactivation the C₆₀ fullerenes (30 min) in a most efficient concentration of 0.01 mg/mL interact with the virions of MIV *Ae. flavescens* causing a reduce of the infectious titre of virus by 4.5 lg ID₅₀/mL units.

Furthermore, the effect of the concentration of C₆₀ fullerenes on the infectious titre of virus was also investigated. In photodynamic inactivation, C₆₀ fullerenes were found to be active against virions of MIV *Ae. flavescens* even at very low concentration (in the concentration range from 0.1 to 0.01 mg/mL). Our results are in agreement with previous published reports^{27,30}.

Declaration of interest

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References

- Kroto H, Heath J, O'Brien S, Curl R, Smalley R. C₆₀: Buckminsterfullerene. *Nature* 1985;318:162-163.
- Bakry R, Vallant RM, Najam-ul-Haq M, Rainer M, Szabo Z, Huck CW et al. Medicinal applications of fullerenes. *Int J Nanomedicine* 2007;2:639-649.
- Cataldo F, Da Ros T. (Eds.). Medicinal chemistry and pharmacological potential of fullerenes and carbon nanotubes. Series. Carbon Materials: Chemistry and Physics. 2008. Springer, Netherlands.
- Da Ros T, Spalluto G, Prato M. Biological Applications of Fullerene Derivatives: A Brief Overview. *Croat Chem Acta* 2001;74:743-755.
- Foley S, Crowley C, Smaih M, Bonfils C, Erlanger BF, Seta P et al. Cellular localisation of a water-soluble fullerene derivative. *Biochem Biophys Res Commun* 2002;294:116-119.
- Prylutska SV, Grynyuk II, Matyshevska OP, Prylutsky YuI, Ritter U, Scharff P. Anti-oxidant Properties of C₆₀ Fullerenes *in vitro*. Fullerenes, Nanotubes, and Carbon Nanostruct 2008;16:698-705.
- Sirotkin AK, Zarubaev VV, Poznyakova LN, Dumpis MA, Muravieva TD., Krisko TK, Belousova IM, Kiselev OI, Piotrovsky LB. Pristine Fullerene C₆₀: Different Water Soluble Forms-Different Mechanisms of Biological Action. Fullerenes, Nanotubes, and Carbon Nanostruct 2006;14:327-333.
- Ji H, Yang Z, Jiang W, Geng C, Gong M, Xiao H et al. Antiviral activity of nano carbon fullerene liposomes against influenza virus *in vitro*. *J Huazhong Univ Sci Technol Med Sci* 2008;28:243-246.
- Tsao N, Luh TY, Chou CK, Chang TY, Wu JJ, Liu CC et al. *In vitro* action of carboxyfullerene. *J Antimicrob Chemother* 2002;49:641-649.
- Li Q, Mahendra S, Lyon DY, Brunet L, Liga MV, Li D et al. Antimicrobial nanomaterials for water disinfection and microbial control: potential applications and implications. *Water Res* 2008;42:4591-4602.
- Spesia MB, Milanese ME, Durantini EN. Synthesis, properties and photodynamic inactivation of *Escherichia coli* by novel cationic fullerene C₆₀ derivatives. *Eur J Med Chem* 2008;43:853-861.
- Bosi S, Da Ros T, Spalluto G, Prato M. Fullerene derivatives: an attractive tool for biological applications. *Eur J Med Chem* 2003;38:913-923.
- Burlaka AP, Sidorik EP, Prylutska SV, Matyshevska OP, Golub AA, Prylutsky YuI, Scharff P. Catalytic system of the reactive oxygen species on the C₆₀ fullerene basis. *Exp Oncol* 2004;26:326-327.
- Käsermann F, Kempf C. Photodynamic inactivation of enveloped viruses by buckminsterfullerene. *Antiviral Res* 1997;34:65-70.
- Zarubaev VV, Belousova IM, Kiselev OI, Piotrovsky LB, Anfimov PM, Krisko TC, Muraviova TD, Rylkov VV, Starodubzev AM, Sirotkin AC. Photodynamic inactivation of influenza virus with fullerene C₆₀ suspension in allantoic fluid. *Photodiagnosis and Photodynamic Therapy* 2007;4:31-35.
- Badireddy AR, Hotze EM, Chellam S, Alvarez P, Wiesner MR. Inactivation of bacteriophages via photosensitization of fullerol nanoparticles. *Environ Sci Technol* 2007;41:6627-6632.
- Mroz P, Tegos GP, Gali H, Wharton T, Sarna T, Hamblin MR. Photodynamic therapy with fullerenes. *Photochem Photobiol Sci* 2007;6:1139-1149.
- Buchatsky LP. Viral infections of marine and freshwater animals. Kiev: Noosphera 1994 (in Russian).
- Chinchar VG, Essbauer S, He JG, Hyatt A, Miyazaki T, Seligy V, Williams T. Eighth report of the international committee on taxonomy of viruses. San Diego: Elsevier, Academic Press. 2005. pp. 150-162.
- Chinchar VG, Essbauer S, He JG, Hyatt AD, Miyazaki T, Seligy V, Williams T. Family Iridoviridae. In: Fauquet CM, Mayo MA, Maniloff J, Desselberger U, Ball LA. (Eds.). Virus taxonomy: classification and nomenclature of viruses: Eighth report of the International Committee on the taxonomy of viruses. Elsevier, Amsterdam, Netherlands. 2005. pp. 145-162.
- Williams T, Barbosa-Solomieu V, Chinchar VG. A decade of advances in iridovirus research. *Adv Virus Res* 2005;65:173-248.
- Scharff P, Risch K, Carta-Abelmann L, Dmytruk IM, Bilyi MM, Golub OA, Khavryuchenko AV, Buzaneva EV, Aksenov VL, Avdeev MV, Prylutsky YuI, Durov SS. Structure of C₆₀ fullerene in water: Spectroscopic data. *Carbon* 2004;42:1203-1206.
- Prylutska SV, Matyshevska OP, Golub AA, Prylutsky YuI, Potebnya GP, Ritter U, Scharff P. Study of C₆₀ fullerenes and C₆₀-containing composites cytotoxicity *in vitro*. *Mater Sci Engineer C* 2007;27:1121-1124.
- Reed L, Muench H. A simple method of estimating fifty percent endpoints. *Am J Hy* 1938;27:493-497.
- Altman EI, Colton RJ. Determination of the orientation of C₆₀ adsorbed on Au(111) and Ag(111). *Phys Rev, B Condens Matter* 1993;48:18244-18249.
- Prilutski YuI, Durov SS, Yashchuk VN, Ogul'chansky TYu, Pogorelov VE, Astashkin YuA, Buzaneva EV, Kirghizov Yu D, Andrievsky GV, Scharff P. Theoretical predictions and experimental studies of self-organized C₆₀ nanoparticles in water solution and on the support. *Eur Phys J D* 1999;9:341-343.
- Bulavin L, Adamenko I, Prylutsky Yu, Durov S, Graja A, Bogucki A, Scharff P. Structure of fullerene C₆₀ in aqueous solution. *Phys Chem Chem Phys* 2000;2:1627-1629.
- Zarubaev VV, Belousova I, Rylkov V, Slita A, Sirotkin A, Anfimov P, Muraviova T, Starodubtsev A. Photodynamic inactivation of enveloped viruses by fullerene: Study of efficacy and safety, medicinal chemistry and pharmacological potential of fullerenes and carbon nanotubes. *Carbon Mat Chem Phys* 2008;1:107-121.
- Marchesan S, Da Ros T, Spalluto G, Balzarini J, Prato M. Anti-HIV properties of cationic fullerene derivatives. *Bioorg Med Chem Lett* 2005;15:3615-3618.
- Mashino T, Shimotohno K, Ikegami N, Nishikawa D, Okuda K, Takahashi K et al. Human immunodeficiency virus-reverse transcriptase inhibition and hepatitis C virus RNA-dependent RNA polymerase inhibition activities of fullerene derivatives. *Bioorg Med Chem Lett* 2005;15:1107-1109.
- Kumar A, Menon SK. Fullerene derivatized s-triazine analogues as antimicrobial agents. *Eur J Med Chem* 2009;44:2178-2183.