



## Evidence for complement activation in the amniotic fluid of women with spontaneous preterm labor and intra-amniotic infection

Eleazar Soto, Roberto Romero, Karina Richani, Bo H. Yoon, Tinnakorn Chaiworapongsa, Edi Vaisbuch, Pooja Mittal, Offer Erez, Francesca Gotsch, Moshe Mazor & Juan P. Kusanovic

**To cite this article:** Eleazar Soto, Roberto Romero, Karina Richani, Bo H. Yoon, Tinnakorn Chaiworapongsa, Edi Vaisbuch, Pooja Mittal, Offer Erez, Francesca Gotsch, Moshe Mazor & Juan P. Kusanovic (2009) Evidence for complement activation in the amniotic fluid of women with spontaneous preterm labor and intra-amniotic infection, *The Journal of Maternal-Fetal & Neonatal Medicine*, 22:11, 983-992, DOI: [10.3109/14767050902994747](https://doi.org/10.3109/14767050902994747)

**To link to this article:** <https://doi.org/10.3109/14767050902994747>



Published online: 17 Mar 2010.



Submit your article to this journal [↗](#)



Article views: 861



View related articles [↗](#)



Citing articles: 9 View citing articles [↗](#)

## Evidence for complement activation in the amniotic fluid of women with spontaneous preterm labor and intra-amniotic infection

ELEAZAR SOTO<sup>1,2</sup>, ROBERTO ROMERO<sup>1,2,3</sup>, KARINA RICHANI<sup>1</sup>, BO H. YOON<sup>4</sup>, TINNAKORN CHAIWORAPONGSA<sup>1,2</sup>, EDI VAISBUCH<sup>1,2</sup>, POOJA MITTAL<sup>1,2</sup>, OFFER EREZ<sup>1,2</sup>, FRANCESCA GOTSCH<sup>1</sup>, MOSHE MAZOR<sup>5</sup>, & JUAN P. KUSANOVIC<sup>1,2</sup>

<sup>1</sup>Perinatology Research Branch, NICHD/NIH/DHHS, Hutzel Women's Hospital, Bethesda, Maryland and Detroit, Michigan, USA, <sup>2</sup>Department of Obstetrics and Gynecology, Wayne State University School of Medicine, Detroit, Michigan, USA, <sup>3</sup>Center for Molecular Medicine and Genetics, Wayne State University, Detroit, Michigan, USA, <sup>4</sup>Department of Obstetrics and Gynecology, Seoul National University, Seoul, Korea, and <sup>5</sup>Department of Obstetrics and Gynecology, Ben Gurion University of the Negev, Beer-Sheva, Israel

(Received 18 February 2009; revised 30 March 2009; accepted 9 April 2009)

### Abstract

**Objective.** The complement system plays an important role in host defense against infection. Concentrations of complement split products or anaphylatoxins (C3a, C4a, and C5a) in biological fluids are considered to reflect complement activation. The purpose of this study was to determine if term and preterm parturition are associated with evidence of complement activation in the amniotic fluid.

**Study design.** Amniotic fluid (AF) samples were collected from 270 women in the following groups: (1) normal pregnant women in midtrimester ( $n=70$ ), (2) term not in labor ( $n=23$ ), (3) term in labor ( $n=48$ ), and (4) preterm labor (PTL) ( $n=129$ ). PTL was categorized into: (a) PTL without microbial invasion of the amniotic cavity (MIAC) who delivered at term ( $n=42$ ), (b) PTL who delivered preterm without MIAC ( $n=57$ ), and (c) PTL with MIAC ( $n=30$ ). C5a, C4a, and C3a concentrations in amniotic fluid were determined by ELISA. Nonparametric tests were used for statistical analysis.

**Results.** (1) The median AF C5a concentration was higher in women at term than that of those in the midtrimester ( $p=0.02$ ); (2) Spontaneous labor at term was not associated with changes in AF concentrations of anaphylatoxins C3a, C4a, and C5a (all  $p > 0.05$ ); (3) Among patients with PTL who delivered preterm, those with MIAC had higher AF C4a and C5a concentrations than those without infection ( $p < 0.01$ ); and (4) AF C3a, C4a, and C5a concentrations were higher in patients with PTL with MIAC than in those with PTL without MIAC who delivered at term.

**Conclusion.** Patients with spontaneous preterm labor and intact membranes with microbial invasion of the amniotic cavity had higher median amniotic fluid concentration of complement split products C3a, C4a, and C5a than patients without intra-amniotic infection. These findings suggest that preterm labor in the context of infection is associated with activation of the complement system.

**Keywords:** C5a, C4a, C3a, anaphylatoxins, pregnancy, MIAC, chorioamnionitis, intra-amniotic inflammation, prematurity, complement

### Introduction

The immune system is composed by an innate and adaptive limb [1]. The innate limb is nonspecific, acts immediately and lacks immunological memory. A key component of innate immunity is the complement system [2,3], which is also involved in the regulation

of the adaptive immune response [4]. The complement system is composed by a group of plasma proteins with catalytic properties that react in a sequential manner, yielding active biological mediators and lytic components to clear microorganisms and 'nonself' cells [2,3,5]. Activation of the complement system through 'the classical', 'the alternative',

and 'the mannose-binding lectin' pathways [3] leads to the generation of complement split products C3a, C4a, and C5a [6]. These bioactive fragments, known as anaphylatoxins, can induce smooth muscle contraction [7–9], enhance vascular permeability [7,9,10], and attract white blood cells [11–13]. In addition to their role in host defense, uncontrolled or excessive production of anaphylatoxins have been implicated in the pathogenesis of inflammatory diseases including sepsis [14–17], asthma [18,19], rheumatoid arthritis [20], acute respiratory distress syndrome [21], ischemic/hypoxic injury [22,23], systemic lupus erythematosus [24], and pregnancy loss [25–28].

Preterm parturition is one of the leading causes of perinatal mortality and long-term neurologic handicap [29]. Intrauterine infection is a frequent and important mechanism of disease in preterm birth [30–37]. Infection triggers an inflammatory response in maternal and fetal tissues mediated by the production of proinflammatory cytokines and chemokines [37,38]. The maternal plasma concentrations of the complement split products C5a and C3a are higher in women with spontaneous preterm labor/delivery with microbial invasion of the amniotic cavity (MIAC) than in those who delivered preterm without MIAC [39]. These findings suggest that there is a systemic maternal immune response to either microbial products or proinflammatory mediators located in the uterine cavity. However, there is paucity of information regarding the changes in amniotic fluid anaphylatoxins during microbial invasion of the amniotic cavity. The purpose of this study was to determine whether term and preterm spontaneous labor and/or MIAC are associated with evidence of complement activation in the amniotic fluid.

## Material and methods

### *Study population*

A cross-sectional study was conducted by searching our clinical database and bank of biological samples, including 270 pregnant women classified into the following groups: (1) women in the midtrimester of pregnancy (14–18 weeks) who underwent amniocentesis for genetic indications and delivered a normal neonate at term ( $n=70$ ); (2) normal pregnant women at term ( $\geq 37$  weeks) not in labor ( $n=23$ ); (3) women with spontaneous labor at term ( $n=48$ ); and (4) women with spontaneous preterm labor and intact membranes (PTL,  $n=129$ ). Women with PTL were classified into: (a) PTL without MIAC who delivered at term ( $n=42$ ); (b) PTL without MIAC who delivered preterm ( $n=57$ ); and (c) PTL with MIAC who delivered preterm ( $n=30$ ).

All women provided written informed consent prior to the collection of amniotic fluid samples. The utilization of samples for research purposes was approved by the Institutional Review Boards of both Wayne State University and by the Eunice Kennedy Shriver National Institute of Child Health and Human Development, NIH, DHHS. Many of these samples have been used in the previous studies of inflammatory mediators, growth factors, and other biological markers of disease.

### *Clinical definitions*

Patients were considered to have a normal pregnancy outcome if they did not have any obstetrical, medical, or surgical complication of pregnancy and delivered a term neonate ( $\geq 37$  weeks) of appropriate birth weight for the gestational age [40]. Spontaneous preterm labor was defined by the presence of regular uterine contractions occurring at a frequency of at least two contractions every 10 min associated with cervical changes that required hospitalization before 37 weeks of gestation. Preterm delivery was defined as delivery before 37 weeks of gestation. Microbial invasion of the amniotic cavity was defined as a positive amniotic fluid culture for microorganisms.

Amniotic fluid samples were obtained from transabdominal amniocentesis performed for genetic indication, evaluation of microbial status of the amniotic cavity and/or assessment of fetal lung maturity in patients approaching term. Women at term in labor consisted of women who were admitted for suspected preterm labor because of uncertain dates and had an amniocentesis for the assessment of fetal lung maturity. The criteria for considering that these patients were at term in labor was derived retrospectively, if the following criteria were met: (1) spontaneous labor; (2) delivery within 24 h from amniocentesis; (3) analysis of amniotic fluid consistent with maturity; (4) birth weight  $> 2500$  g; (5) absence of respiratory distress syndrome or other complications of prematurity; and (6) physical examination of the newborn by pediatricians consistent with a term neonate. Immediately upon retrieval, amniotic fluid was transported to the laboratory in a capped plastic sterile syringe and cultured for aerobic/anaerobic bacteria and genital mycoplasmas (*Ureaplasma urealyticum* and *Mycoplasma hominis*), except in the midtrimester group. White blood cells count, glucose concentration, and Gram-stain were also performed shortly after collection except in the midtrimester group. The results of these tests were used for subsequent clinical management. Amniotic fluid not required for clinical purposes was centrifuged for 10 minutes at  $4^{\circ}\text{C}$  and stored at  $-70^{\circ}\text{C}$  until analysis.

### Complement C3a, C4a, and C5a immunoassays

The characteristics of the assays using this study have been previously described in publications by our group [39,41]. The calculated inter- and intra-assay coefficients of variation for C3a, C4a, and C5a immunoassays in our laboratory were 4.7, 6.4, and 4.1%, and 4.9, 5.8, and 2.5%, respectively. The sensitivity was 0.13 ng/ml for C3a assay, 0.27 ng/ml for C4a assay, and 0.06 ng/ml for C5a assay.

### Statistical analysis

The Shapiro–Wilk test was used to test for normal distribution of the data. Kruskal–Wallis with *post hoc* Mann–Whitney *U* tests was performed when indicated to determine the difference of the median among groups, and Bonferroni correction was applied to adjust for multiple comparisons.  $\chi^2$  test was used for comparison of proportions. The statistical package used was SPSS 12 (SPSS; Chicago, IL). A probability value of  $<0.05$  was considered significant.

### Results

Tables I and II display the demographic and clinical characteristics of pregnant women in the

midtrimester and term groups, and of those with spontaneous preterm labor, respectively. There were no significant differences in the maternal age, nulliparity gestational age at delivery, gestational age at amniocentesis, and birth weight between women at term in labor and those not in labor (Table I).

The gestational age at amniocentesis and delivery, as well as the birth weight, were significantly higher among women who had an episode of preterm labor and delivered at term when compared with those who delivered preterm with or without MIAC (Table II). Among patients with spontaneous preterm labor and delivery without MIAC, the gestational age at delivery, and birth weight were significantly higher than that of those who delivered preterm with MIAC (Table II).

Complement split products C3a, C4a, and C5a were detected in all amniotic fluid samples. Women at term not in labor had a significantly higher median C5a amniotic fluid concentration than those in the midtrimester (see Figure 1C). In contrast, no differences were observed in the median amniotic fluid concentrations of C3a and C4a (see Figures 1A and 1B, respectively).

Among normal pregnancies at term, there was no difference in the median amniotic fluid concentration of C3a, C4a, and C5a between women not in labor and those in labor (see Figure 2).

Table I. Demographic and clinical characteristics of women who underwent amniocentesis at midtrimester and term gestation not in labor and in spontaneous labor.

	Midtrimester, <i>n</i> = 70	Term; no labor, <i>n</i> = 23	Term; in labor, <i>n</i> = 48
Maternal age (years)	37 (35–38)*	27 (21–32)	23 (20–27.5)
Nulliparity	12 (17.1)**	5 (23.8)†	23 (47.9)
Gestational age at amniocentesis (weeks)	16 (16–17)*	39.7 (38.7–40)	39.1 (38–40.2)
Gestational age at delivery (weeks)	39.5 (38–40)	39.5 (38.5–40)	39.1 (38–40.1)
Birth weight (g)	3345 (3103–3626.5)‡	3430 (3130–3790)†	3265 (3092–3695)

Values are expressed as median (interquartile range) or number (percent).

\**p* < 0.05 compared with term not in labor and in labor.

\*\**p* < 0.05 compared with term in labor.

†*n* = 21.

‡*n* = 69.

Table II. Demographic and clinical characteristics of women with spontaneous preterm labor and intact membranes.

	Preterm labor; No MIAC; Term delivery, <i>n</i> = 42	Preterm labor; No MIAC; Preterm delivery, <i>n</i> = 57	Preterm labor; MIAC; Preterm delivery, <i>n</i> = 30
Maternal age (years)	21.5 (19–27)	23 (20–27)	25 (20–29.2)
Nulliparity	10 (23.8)	21 (36.8)	14 (46.7)**
Gestational age at amniocentesis (weeks)	30.6 (28.7–32.2)	27.2 (25–29.6)*	25.9 (23.6–29.2)**
Gestational age at delivery (weeks)	39 (37.6–40.1)	30.5 (26.1–33.4)*	26.1 (24–30.2)**†
Birth weight (g)	3035 (2773–3296)	1300 (850–1958)*	844 (515–1395)**†

MIAC, microbial invasion of the amniotic cavity.

Values are expressed as median (interquartile range) or number (percent).

\**p* < 0.05 compared with preterm labor no MIAC with term delivery.

\*\**p* < 0.05 compared with preterm labor no MIAC with term delivery.

†*p* < 0.05 compared with preterm labor no MIAC with preterm delivery.

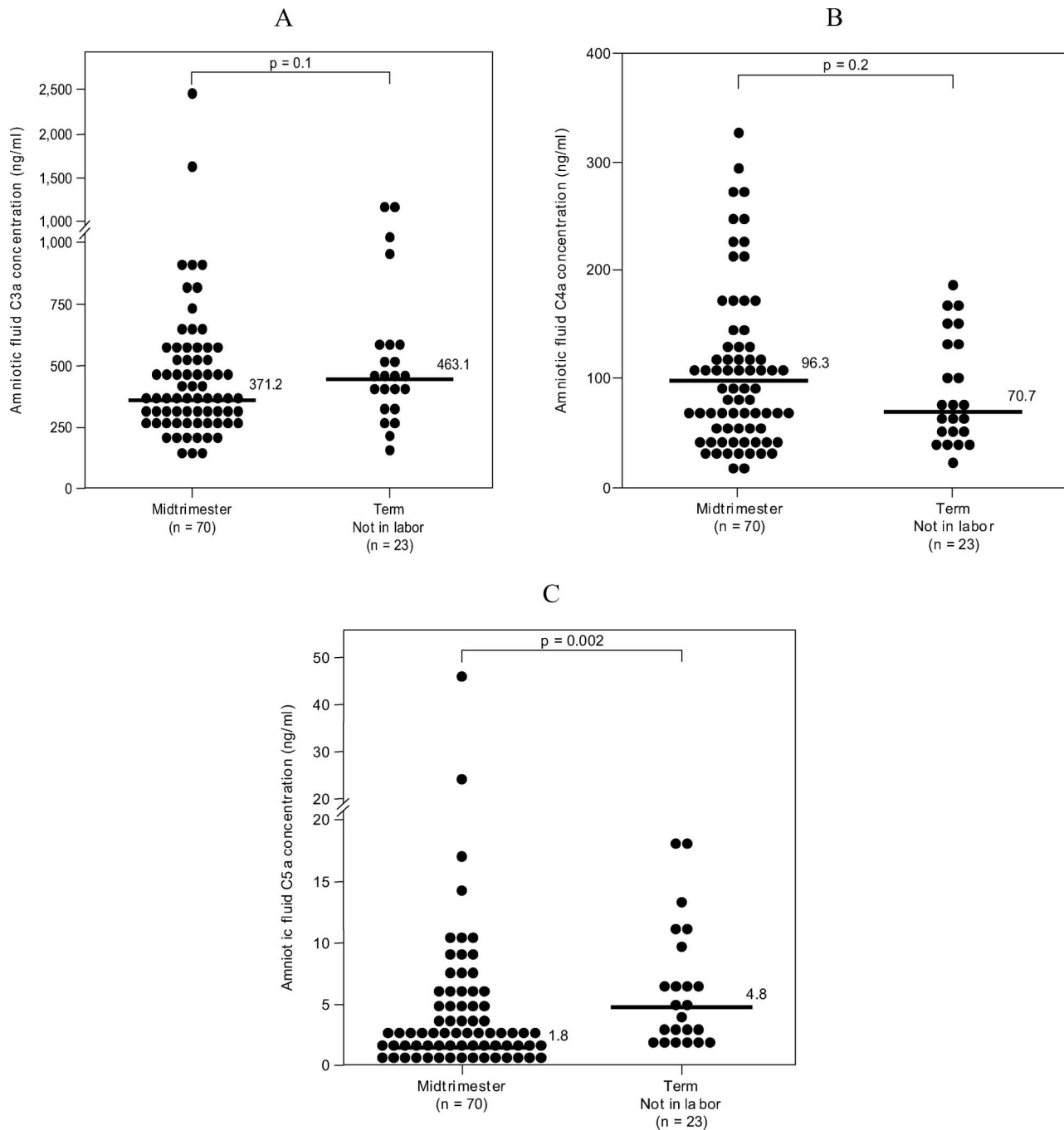


Figure 1. Amniotic fluid anaphylatoxins concentrations from women in the midtrimester (14–18 weeks of gestation) who delivered a normal neonate at term and those at term not in labor. A and B: There were no differences in the median amniotic fluid concentrations of C3a and C4a between pregnant women in the midtrimester and those at term not in labor [C3a: median 371.2 ng/ml, (range 130.4–2468.2) *vs.* median 463.1 ng/ml, (range 161.9–1131.3);  $p = 0.1$ ] and [C4a: median 96.3 ng/ml, (range 13.8–326.2) *vs.* median 70.7 ng/ml, (range 21.4–184.4);  $p = 0.2$ ]; C: In contrast, the median amniotic fluid concentration of C5a was significantly higher in women at term not in labor than in those in the midtrimester [median 4.8 ng/ml, (range 1.7–18.1) *vs.* median 1.8 ng/ml, (range 0.07–46.3);  $p = 0.002$ ].

Among patients with spontaneous preterm labor, those with MIAC had a significantly higher median amniotic fluid concentration of C3a, C4a, and C5a than those with preterm labor without MIAC who delivered at term (all  $p < 0.05$ ) (Figures 3A–3C). Similarly, women with spontaneous preterm labor and MIAC had a significantly higher median amniotic

fluid concentration of C4a and C5a, but not of C3a, than those with spontaneous preterm labor without MIAC who delivered preterm (C4a and C5a:  $p < 0.05$ ); (C3a:  $p > 0.05$ ) (Figures 3A–3C). There were no differences in the median C3a, C4a, and C5a amniotic fluid concentrations between women with spontaneous preterm labor without MIAC who

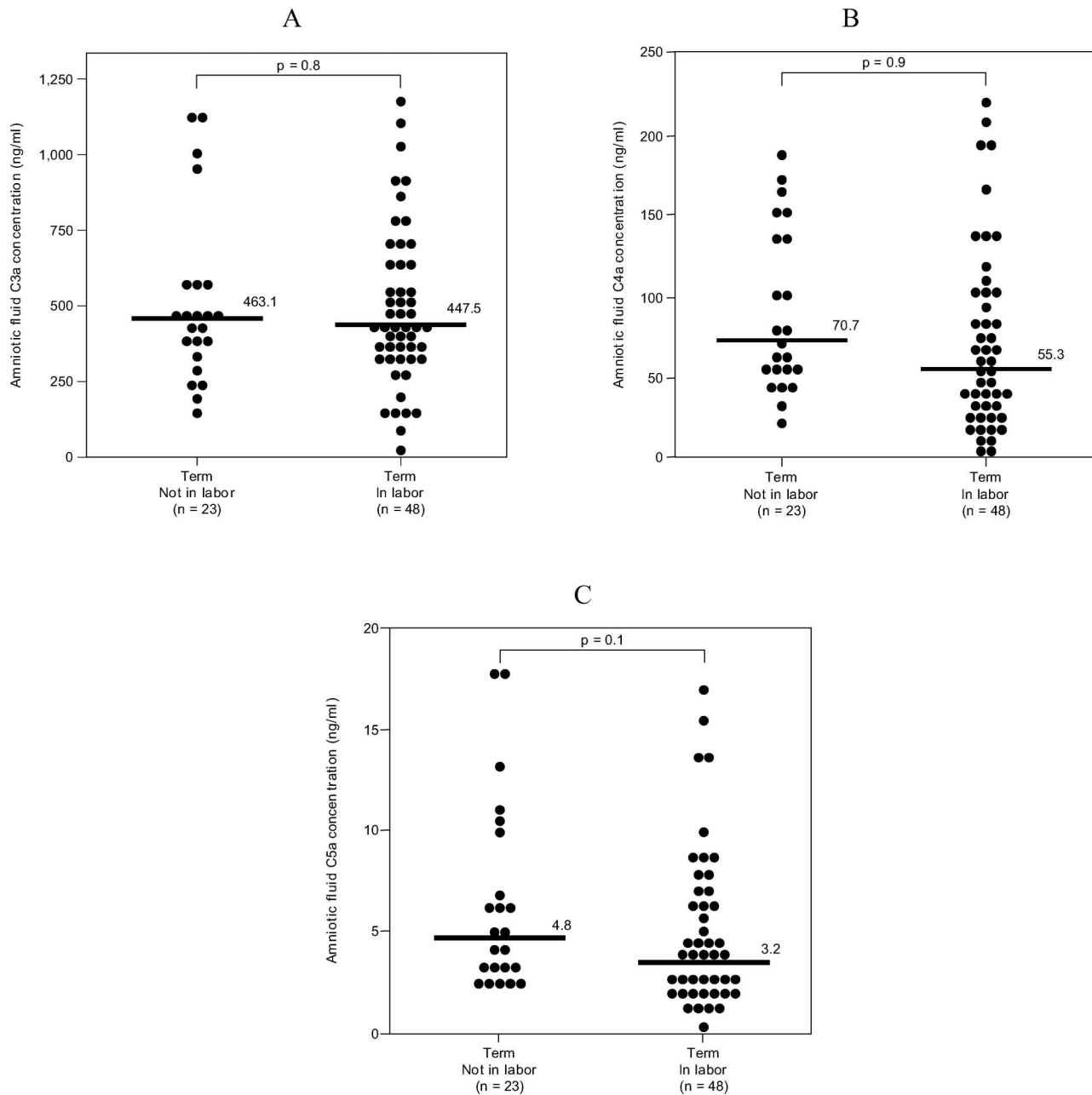


Figure 2. Amniotic fluid anaphylatoxins concentrations of normal pregnant women at term. A, B, C: There were no significant differences in the median amniotic fluid concentrations of C3a, C4a, and C5a between women at term in labor and those not in labor [C3a: median 463.1 ng/ml (range 161.9–1131.3) *vs.* median 447.5 ng/ml (range 12.3–1196.3;  $p=0.8$ ); [C4a: median 70.7 ng/ml (range 21.4–184.4) *vs.* median 55.3 ng/ml (range 0.6–217.1;  $p=0.9$ ); and [C5a: median 4.8 ng/ml (range 1.7–18.1) *vs.* median 3.2 ng/ml (range 0.07–17.1);  $p=0.1$ ].

delivered preterm and those who had an episode of spontaneous preterm labor and delivered at term (all  $p$ -values  $> 0.05$ ) (Figures 3A–3C).

## Discussion

### Principal findings of the study

(1) Among patients with spontaneous preterm labor, those with microbial invasion of the amniotic cavity have significantly higher median amniotic fluid concentrations of C3a, C4a, and C5a than those

with a negative amniotic fluid culture; (2) the amniotic fluid concentration of C5a, but not that of C3a and C4a, increases with advancing gestational age; and (3) spontaneous labor at term is not associated with changes in amniotic fluid C3a, C4a, and C5a concentrations.

### What are anaphylatoxins C3a, C4a, and C5a?

These complement split products are known as anaphylatoxins because of their properties to induce edema, increase vascular permeability [10], and to



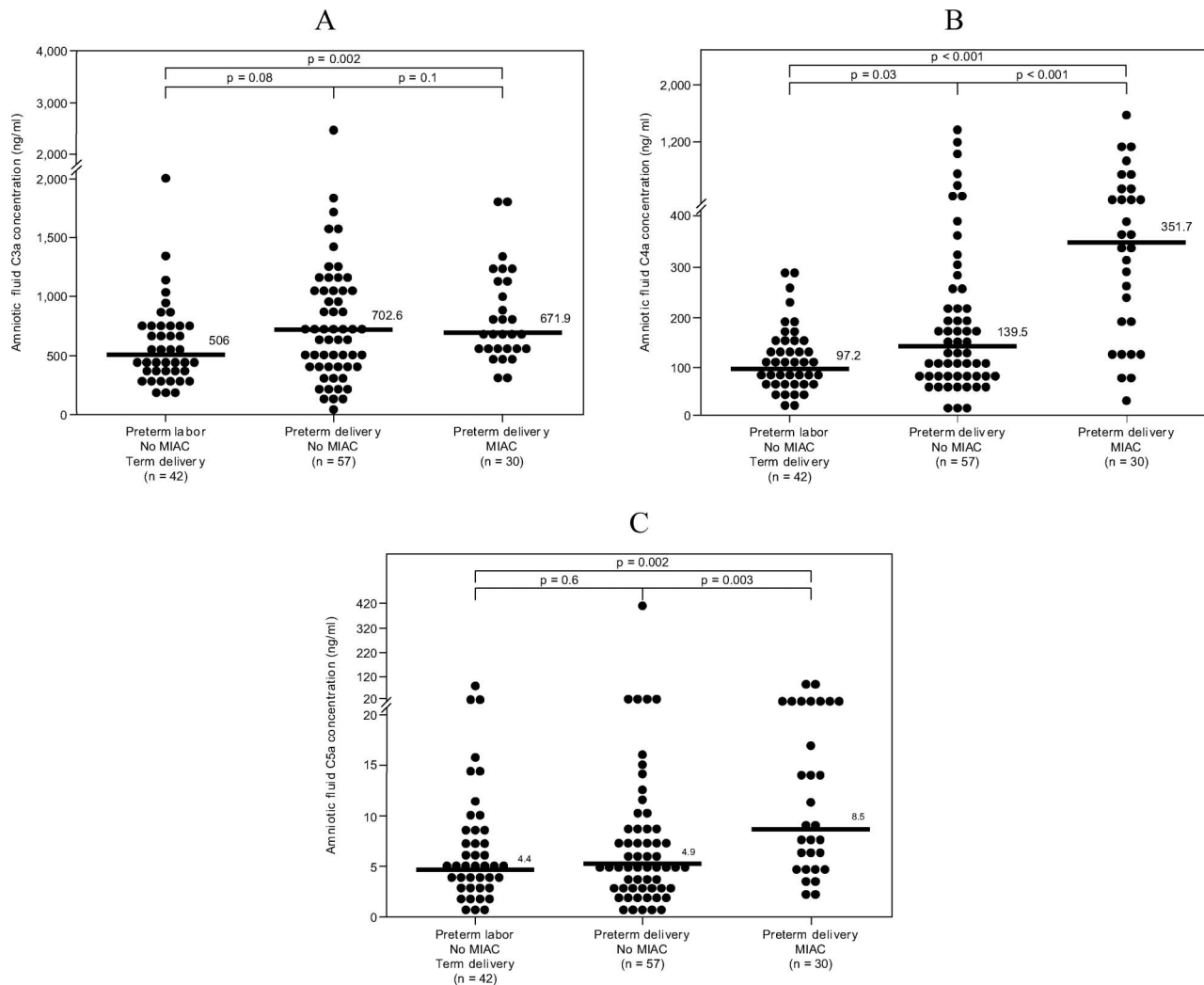


Figure 3. Amniotic fluid anaphylatoxins concentrations in patients with preterm labor. A: Patients with preterm delivery and MIAC had a median amniotic fluid C3a concentration higher than those who in preterm labor and delivered at term [median: 671.9 ng/ml (range 312.8–3552.9) vs. median: 506 ng/ml (range 184.5–1992.8)]. There was no difference in the median amniotic fluid C3a concentration between patients in preterm labor who delivered at term and those who delivered preterm without MIAC. Similarly, there was no difference in the median amniotic fluid C3a concentration between patients with preterm delivery and MIAC and those without MIAC. B: Patients with preterm delivery and MIAC had a higher median amniotic fluid C4a concentration than those who had a preterm delivery without MIAC [median: 351.7 ng/ml (range 24.8–1640.4) vs. median: 139.5 ng/ml (range 0.6–1377.4)]. Similarly, patients with preterm delivery and MIAC had a higher median amniotic fluid C4a concentration than those who had preterm labor a delivered at term [median: 351.7 ng/ml (range 24.8–1640.4) vs. median: 97.2 ng/ml (range 130.4–2468.2)]. In contrast, there was no difference between the median amniotic fluid C4a concentration of women who had a preterm delivery without MIAC and those who had preterm labor and delivery at term. C: Patients with preterm delivery and MIAC had a higher median amniotic fluid C5a concentration than those who had a preterm delivery without MIAC [median: 8.5 ng/ml (range 2.2–83.2) vs. median: 4.9 ng/ml (range 0.07–400)]. Similarly, patients with preterm delivery and MIAC had a higher median amniotic fluid C5a concentration than those who had preterm labor a delivered at term [median: 8.5 ng/ml (range 2.2–83.2) vs. median: 4.4 ng/ml (range 0.5–83.5)]. In contrast, there was no difference between the median amniotic fluid C5a concentration in women who had a preterm delivery without MIAC and those who had preterm labor and delivery at term.

stimulate smooth muscle contractions [7]. C3a biological effects are predominantly on mast cells and eosinophils, and include: (1) chemotaxis [12,13]; (2) granule release [12,42]; (3) expression and shedding of adhesion molecules [43]; (4) increased oxidative burst in neutrophils and eosinophils [44,45]; and (5) immunomodulation of IL-1, IL-6, and TNF- $\alpha$  [46,47]. C4a is the weakest of all anaphylatoxins inducing vascular permeability and

smooth muscle contraction [9]. Nonetheless, C4a may modulate the inflammatory response because it can inhibit monocytes chemotaxis [48]. The biological activities of C5a depend on the target cell including: (1) chemotaxis [11–13] and degranulation of inflammatory cells [12,42,49]; (2) enhancement of the respiratory burst and consequent generation of reactive oxygen species in leukocytes [50–52]; and (3) delayed neutrophil apoptosis [53]. Other

important functions of C5a are the induction and/or release of inflammatory cytokines such as IL-1 [54–56], IL-6 [57–59], IL-8 [60], and TNF- $\alpha$  [55,56] from neutrophils, mononuclear and endothelial cells.

#### *The complement system in the fetus and amniotic fluid*

Complement proteins has been detected in cord blood [61–67], placenta [68–70], and chorioamniotic membranes [70–72]. Interestingly, the presence of complement proteins in amniotic fluid has been previously reported [64,73–75]. Stabile et al. [64] used rocket immunoelectrophoresis and detected complement factors C3, C4, C5, Factor B, H, and I in amniotic fluid and cord blood of normal pregnancies between 15 and 28 weeks of gestation. The authors reported that the concentrations of these complement proteins were 10 times higher in cord blood than in amniotic fluid, and that the concentrations of some of these proteins (C3 and Factor B) increased with gestational age in amniotic fluid. Similarly, Sharma et al. [73] reported that amniotic fluid concentration of C3 increased from the first to the second trimester, but not in the third trimester.

C3a, C4a, and C5a were detected in all amniotic fluid samples included in this study, suggesting that these anaphylatoxins are physiologic constituents of the amniotic fluid. Haeger et al. [76] were the first to describe the presence of complement anaphylatoxins C3a and C5a in amniotic fluid collected in patients with preeclampsia and in those with uncomplicated pregnancies. The concentration of C3a and C5a did not differ between the two groups. Interestingly, the authors performed an *in vitro* study incubating amniotic fluid with fresh plasma, and a dose-dependent release of C3a and C5a in the plasma was noted. The authors concluded that amniotic fluid can activate the complement cascade. Our results indicate that the amniotic fluid concentration of C5a changes during gestation, because women at term not in labor had a higher median amniotic fluid concentration of C5a than those in the midtrimester. In contrast, no changes were observed in the amniotic fluid concentrations of C3a and C4a (direct split product of C3 and C4, respectively) with advancing gestational age.

#### *Amniotic fluid complement split products and term gestation*

In this study, labor at term was not associated with changes in the amniotic fluid concentration of anaphylatoxins. This is not consistent with the conventional view that spontaneous labor at term is an inflammatory process [77,78] characterized by increased maternal neutrophil count [79] and increased maternal serum/plasma concentrations of

proinflammatory cytokines (IL-1 $\beta$ , IL-6, IL-8, TNF  $\alpha$ ) [77,80–82] and chemokines (growth-related oncogene  $\alpha$ , granulocyte colony stimulating factor, granulocyte macrophage colony stimulating factor, neutrophil attractant/activating peptide-1, etc.) [77,83]. However, it is possible that the low-grade inflammatory state involved in the process of labor may not require the activation of the complement system. Alternatively, if the fetus is the main source of the complement system in the amniotic fluid, it is possible that inflammation during normal labor at term may not be enough to activate the complement system in the amniotic cavity.

#### *Complement and preterm delivery*

The observation that MIAC is associated with complement activation in patients with preterm labor, as indicated by elevated amniotic fluid concentrations of C3a, C4a, and C5a, is novel. Elimian et al. [74] reported that among patients with preterm labor and intact membranes, those with positive amniotic fluid cultures had a higher amniotic fluid concentration of C3 (total protein) than those with negative amniotic fluid cultures. Moreover, the authors reported that the amniotic fluid concentration of C3 had similar diagnostic performance as other markers of intra-amniotic infection [74], such as amniotic fluid white blood cell count [84], glucose concentration [85], Gram stain [86], and LDH [87].

In a previous study, the maternal serum complement hemolytic activity (CH50) in women with preterm labor was similar between those who delivered preterm and those who delivered at term [75]. However, the CH50 assay is an insensitive marker of complement activation [88]. Recently, Lynch et al. [89] proposed that complement activation in early pregnancy may lead to preterm delivery. In a prospective study, the authors determined that elevated plasma concentrations of factor Bb (primarily part of the alternative pathway) was predictive of delivery at less than 34 weeks of gestation. Indeed, women with factor Bb plasma concentrations in the top quartile prior to 20 weeks of gestation were 4.7 times more likely to have a spontaneous preterm delivery before 34 weeks compared with women who had factor Bb plasma concentrations in the lower three quartiles (95% confidence interval 1.5–14). Our group reported that patients with intra-amniotic infection/inflammation (regardless of the membranes status) had higher median amniotic fluid fragment Bb concentrations than that of those without IAI (in press) [90]. In another study, maternal plasma C5a concentrations were higher in patients with spontaneous preterm labor with MIAC than that of those with preterm labor without MIAC delivering



preterm or at term [39]. This finding suggests that the maternal immune system is responding to either microbial products or proinflammatory mediators produced in the amniotic cavity, and that such response can be detected in the maternal compartment.

In conclusion, this study demonstrates that patients with spontaneous preterm labor and intact membranes with microbial invasion of the amniotic cavity had increased amniotic fluid concentrations of complement split products C3a, C4a, and C5a.

## Acknowledgements

This research was supported (in part) by the Perinatology Research Branch, Division of Intramural Research, Eunice Kennedy Shriver National Institute of Child Health and Human Development, NIH, DHHS.

## References

- Medzhitov R, Janeway C Jr. Innate immunity. *N Engl J Med* 2000;343:338–344.
- Gasque P. Complement: a unique innate immune sensor for danger signals. *Mol Immunol* 2004;41:1089–1098.
- Walport MJ. Complement. First of two parts. *N Engl J Med* 2001;344:1058–1066.
- Carroll MC. The complement system in regulation of adaptive immunity. *Nat Immunol* 2004;5:981–986.
- Medzhitov R, Janeway CA Jr. Decoding the patterns of self and nonself by the innate immune system. *Science* 2002;296:298–300.
- Ember JA, Jagels MA, Hugli TE. Characterization of complement anaphylatoxins and their biological responses. In: Volanakis JE, Frank MM, editors. *The Human Complement System in Health and Disease*. New York: Marcel Dekker; 1998. pp 241–284.
- Cochrane CG, Muller-Eberhard HJ. The derivation of two distinct anaphylatoxin activities from the third and fifth components of human complement. *J Exp Med* 1968;127:371–386.
- Dias DS, Lepow IH. Complement as a mediator of inflammation. II. Biological properties of anaphylatoxin prepared with purified components of human complement. *J Exp Med* 1967;125:921–946.
- Gorski JP, Hugli TE, Muller-Eberhard HJ. C4a: the third anaphylatoxin of the human complement system. *Proc Natl Acad Sci USA* 1979;76:5299–5302.
- Schumacher WA, Fantone JC, Kunkel SE, Webb RC, Lucchesi BR. The anaphylatoxins C3a and C5a are vasodilators in the canine coronary vasculature in vitro and in vivo. *Agents Actions* 1991;34:345–349.
- Shin HS, Snyderman R, Friedman E, Mellors A, Mayer MM. Chemotactic and anaphylatoxic fragment cleaved from the fifth component of guinea pig complement. *Science* 1968;162:361–363.
- Daffern PJ, Pfeifer PH, Ember JA, Hugli TE. C3a is a chemotaxin for human eosinophils but not for neutrophils. I. C3a stimulation of neutrophils is secondary to eosinophil activation. *J Exp Med* 1995;181:2119–2127.
- Hartmann K, Henz BM, Kruger-Krasagakes S, Kohl J, Burger R, Guhl S, Haase I, Lippert U, Zuberbier T. C3a and C5a stimulate chemotaxis of human mast cells. *Blood* 1997;89:2863–2870.
- Hack CE, Nuijens JH, Felt-Bersma RJ, Schreuder WO, Eerenberg-Belmer AJ, Paardekooper J, Bronsveld W, Thijs LG. Elevated plasma levels of the anaphylatoxins C3a and C4a are associated with a fatal outcome in sepsis. *Am J Med* 1989;86:20–26.
- Nakae H, Endo S, Inada K, Yoshida M. Chronological changes in the complement system in sepsis. *Surg Today* 1996;26:225–229.
- Czermak BJ, Sarma V, Pierson CL, Warner RL, Huber-Lang M, Bless NM, Schmal H, Friedl HP, Ward PA. Protective effects of C5a blockade in sepsis. *Nat Med* 1999;5:788–792.
- Ward PA. The dark side of C5a in sepsis. *Nat Rev Immunol* 2004;4:133–142.
- Humbles AA, Lu B, Nilsson CA, Lilly C, Israel E, Fujiwara Y, Gerard NP, Gerard C. A role for the C3a anaphylatoxin receptor in the effector phase of asthma. *Nature* 2000;406:998–1001.
- Nakano Y, Morita S, Kawamoto A, Suda T, Chida K, Nakamura H. Elevated complement C3a in plasma from patients with severe acute asthma. *J Allergy Clin Immunol* 2003;112:525–530.
- Moxley G, Ruddy S. Elevated plasma C3 anaphylatoxin levels in rheumatoid arthritis patients. *Arthritis Rheum* 1987;30:1097–1104.
- Robbins RA, Russ WD, Rasmussen JK, Clayton MM. Activation of the complement system in the adult respiratory distress syndrome. *Am Rev Respir Dis* 1987;135:651–658.
- Riedemann NC, Ward PA. Complement in ischemia reperfusion injury. *Am J Pathol* 2003;162:363–367.
- Arumugam TV, Shiels IA, Woodruff TM, Granger DN, Taylor SM. The role of the complement system in ischemia-reperfusion injury. *Shock* 2004;21:401–409.
- Hopkins P, Belmont HM, Buyon J, Philips M, Weissmann G, Abramson SB. Increased levels of plasma anaphylatoxins in systemic lupus erythematosus predict flares of the disease and may elicit vascular injury in lupus cerebritis. *Arthritis Rheum* 1988;31:632–641.
- Girardi G, Berman J, Redecha P, Spruce L, Thurman JM, Kraus D, Hollmann TJ, Casali P, Carroll MC, Wetsel RA, et al. Complement C5a receptors and neutrophils mediate fetal injury in the antiphospholipid syndrome. *J Clin Invest* 2003;112:1644–1654.
- Holers VM, Girardi G, Mo L, Guthridge JM, Molina H, Pierangeli SS, Espinola R, Xiaowei LE, Mao D, Vialpando CG, et al. Complement C3 activation is required for antiphospholipid antibody-induced fetal loss. *J Exp Med* 2002;195:211–220.
- Salmon JE, Girardi G, Holers VM. Complement activation as a mediator of antiphospholipid antibody induced pregnancy loss and thrombosis. *Ann Rheum Dis* 2002;61 (Suppl. 2):ii46–ii50.
- Xu C, Mao D, Holers VM, Palanca B, Cheng AM, Molina H. A critical role for murine complement regulator crry in fetomaternal tolerance. *Science* 2000;287:498–501.
- Stoll BJ, Hansen NI, Adams-Chapman I, Fanaroff AA, Hintz SR, Vohr B, Higgins RD. Neurodevelopmental and growth impairment among extremely low-birth-weight infants with neonatal infection. *JAMA* 2004;292:2357–2365.
- Minkoff H. Prematurity: infection as an etiologic factor. *Obstet Gynecol* 1983;62:137–144.
- Romero R, Mazar M. Infection and preterm labor. *Clin Obstet Gynecol* 1988;31:553–584.
- Romero R, Mazar M, Wu YK, Sirtori M, Oyarzun E, Mitchell MD, Hobbins JC. Infection in the pathogenesis of preterm labor. *Semin Perinatol* 1988;12:262–279.
- McGregor JA, French JI, Lawellin D, Todd JK. Preterm birth and infection: pathogenic possibilities. *Am J Reprod Immunol Microbiol* 1988;16:123–132.

34. Romero R, Sirtori M, Oyarzun E, Avila C, Mazor M, Callahan R, Sabo V, Athanassiadis AP, Hobbins JC. Infection and labor. V. Prevalence, microbiology, and clinical significance of intraamniotic infection in women with preterm labor and intact membranes. *Am J Obstet Gynecol* 1989;161:817–824.
35. Goldenberg RL, Culhane JF. Infection as a cause of preterm birth. *Clin Perinatol* 2003;30:677–700.
36. Boggess KA. Pathophysiology of preterm birth: emerging concepts of maternal infection. *Clin Perinatol* 2005;32:561–569.
37. Romero R, Espinoza J, Kusanovic J, Gotsch F, Hassan S, Erez O, Chaiworapongsa T, Mazor M. The preterm parturition syndrome. *BJOG* 2006;113 (Suppl. 3):17–42.
38. Romero R, Espinoza J, Goncalves LF, Kusanovic JP, Friel L, Hassan S. The role of inflammation and infection in preterm birth. *Semin Reprod Med* 2007;25:21–39.
39. Soto E, Romero R, Richani K, Espinoza J, Nien JK, Chaiworapongsa T, Santolaya-Forgas J, Edwin SS, Mazor M. Anaphylatoxins in preterm and term labor. *J Perinat Med* 2005;33:306–313.
40. Alexander GR, Himes JH, Kaufman RB, Mor J, Kogan M. A United States national reference for fetal growth. *Obstet Gynecol* 1996;87:163–168.
41. Soto E, Richani K, Romero R, Espinoza J, Chaiworapongsa T, Nien JK, Edwin S, Kim YM, Hong JS, Goncalves L, et al. Increased concentration of the complement split product C5a in acute pyelonephritis during pregnancy. *J Matern Fetal Neonatal Med* 2005;17:247–252.
42. Takafuji S, Tadokoro K, Ito K, Dahinden CA. Degranulation from human eosinophils stimulated with C3a and C5a. *Int Arch Allergy Immunol* 1994;104 (Suppl. 1):27–29.
43. Jagels MA, Daffern PJ, Hugli TE. C3a and C5a enhance granulocyte adhesion to endothelial and epithelial cell monolayers: epithelial and endothelial priming is required for C3a-induced eosinophil adhesion. *Immunopharmacology* 2000;46:209–222.
44. Elsner J, Oppermann M, Czech W, Kapp A. C3a activates the respiratory burst in human polymorphonuclear neutrophilic leukocytes via pertussis toxin-sensitive G-proteins. *Blood* 1994;83:3324–3331.
45. Elsner J, Oppermann M, Czech W, Dobos G, Schopf E, Norgauer J, Kapp A. C3a activates reactive oxygen radical species production and intracellular calcium transients in human eosinophils. *Eur J Immunol* 1994;24:518–522.
46. Takabayashi T, Vannier E, Burke JF, Tompkins RG, Gelfand JA, Clark BD. Both C3a and C3a(desArg) regulate interleukin-6 synthesis in human peripheral blood mononuclear cells. *J Infect Dis* 1998;177:1622–1628.
47. Takabayashi T, Vannier E, Clark BD, Margolis NH, Dinarello CA, Burke JF, Gelfand JA. A new biologic role for C3a and C3a desArg: regulation of TNF- $\alpha$  and IL-1  $\beta$  synthesis. *J Immunol* 1996;156:3455–3460.
48. Tsuruta T, Yamamoto T, Matsubara S, Nagasawa S, Tanase S, Tanaka J, Takagi K, Kambara T. Novel function of C4a anaphylatoxin. Release from monocytes of protein which inhibits monocyte chemotaxis. *Am J Pathol* 1993;142:1848–1857.
49. Haeger M, Unander M, Norder-Hansson B, Tylman M, Bengtsson A. Complement, neutrophil, and macrophage activation in women with severe preeclampsia and the syndrome of hemolysis, elevated liver enzymes, and low platelet count. *Obstet Gynecol* 1992;79:19–26.
50. Wymann MP, Kernen P, Deranleau DA, Baggiolini M. Respiratory burst oscillations in human neutrophils and their correlation with fluctuations in apparent cell shape. *J Biol Chem* 1989;264:15829–15834.
51. Goldstein IM, Roos D, Kaplan HB, Weissmann G. Complement and immunoglobulins stimulate superoxide production by human leukocytes independently of phagocytosis. *J Clin Invest* 1975;56:1155–1163.
52. Ehrengreuber MU, Geiser T, Deranleau DA. Activation of human neutrophils by C3a and C5a. Comparison of the effects on shape changes, chemotaxis, secretion, and respiratory burst. *FEBS Lett* 1994;346:181–184.
53. Perianayagam MC, Balakrishnan VS, King AJ, Pereira BJ, Jaber BL. C5a delays apoptosis of human neutrophils by a phosphatidylinositol 3-kinase-signaling pathway. *Kidney Int* 2002;61:456–463.
54. Okusawa S, Dinarello CA, Yancey KB, Endres S, Lawley TJ, Frank MM, Burke JF, Gelfand JA. C5a induction of human interleukin 1. Synergistic effect with endotoxin or interferon-gamma. *J Immunol* 1987;139:2635–2640.
55. Okusawa S, Yancey KB, van der Meer JW, Endres S, Lonnemann G, Hefter K, Frank MM, Burke JF, Dinarello CA, Gelfand JA. C5a stimulates secretion of tumor necrosis factor from human mononuclear cells in vitro. Comparison with secretion of interleukin 1  $\beta$  and interleukin 1  $\alpha$ . *J Exp Med* 1988;168:443–448.
56. Schindler R, Gelfand JA, Dinarello CA. Recombinant C5a stimulates transcription rather than translation of interleukin-1 (IL-1) and tumor necrosis factor: translational signal provided by lipopolysaccharide or IL-1 itself. *Blood* 1990;76:1631–1638.
57. Riedemann NC, Guo RF, Hollmann TJ, Gao H, Neff TA, Reuben JS, Speyer CL, Sarma JV, Wetsel RA, Zetoune FS, et al. Regulatory role of C5a in LPS-induced IL-6 production by neutrophils during sepsis. *FASEB J* 2004;18:370–372.
58. Scholz W, McClurg MR, Cardenas GJ, Smith M, Noonan DJ, Hugli TE, Morgan EL. C5a-mediated release of interleukin 6 by human monocytes. *Clin Immunol Immunopathol* 1990;57:297–307.
59. Albrecht EA, Chinnaiyan AM, Varambally S, Kumar-Sinha C, Barrette TR, Sarma JV, Ward PA. C5a-induced gene expression in human umbilical vein endothelial cells. *Am J Pathol* 2004;164:849–859.
60. Ember JA, Sanderson SD, Hugli TE, Morgan EL. Induction of interleukin-8 synthesis from monocytes by human C5a anaphylatoxin. *Am J Pathol* 1994;144:393–403.
61. Fireman P, Zuchowski DA, Taylor PM. Development of human complement system. *J Immunol* 1969;103:25–31.
62. Ballow M, Fang F, Good RA, Day NK. Developmental aspects of complement components in the newborn. The presence of complement components and C3 proactivator (properdin factor B) in human colostrum. *Clin Exp Immunol* 1974;18:257–266.
63. Miyano A, Nakayama M, Fujita T, Kitajima H, Imai S, Shimizu A. Complement activation in fetuses: assessment by the levels of complement components and split products in cord blood. *Diagn Clin Immunol* 1987;5:86–90.
64. Stabile I, Nicolaides KH, Bach A, Teisner B, Rodeck C, Westergaard JG, Grudzinskas JG. Complement factors in fetal and maternal blood and amniotic fluid during the second trimester of normal pregnancy. *Br J Obstet Gynaecol* 1988;95:281–285.
65. Zilow G, Zilow EP, Burger R, Linderkamp O. Complement activation in newborn infants with early onset infection. *Pediatr Res* 1993;34:199–203.
66. Enskog A, Bengtsson A, Bengtson JP, Heideman M, Andreasson S, Larsson L. Complement anaphylatoxin C3a and C5a formation in premature children with respiratory distress. *Eur J Pediatr* 1996;155:41–45.
67. Sonntag J, Brandenburg U, Polzehl D, Strauss E, Vogel M, Dudenhausen JW, Obladen M. Complement system in healthy term newborns: reference values in umbilical cord blood. *Pediatr Dev Pathol* 1998;1:131–135.

68. Kohler PF. Maturation of the human complement system. I. Onset time and sites of fetal C1q, C4, C3, and C5 synthesis. *J Clin Invest* 1973;52:671–677.
69. Goldberg M, Luknar-Gabor N, Keidar R, Katz Y. Synthesis of complement proteins in the human chorion is differentially regulated by cytokines. *Mol Immunol* 2007;44:1737–1742.
70. Holmes CH, Simpson KL. Complement and pregnancy: new insights into the immunobiology of the fetomaternal relationship. *Baillieres Clin Obstet Gynaecol* 1992;6:439–460.
71. Vanderpuye OA, Labarrere CA, McIntyre JA. Expression of CD59, a human complement system regulatory protein, in extraembryonic membranes. *Int Arch Allergy Immunol* 1993;101:376–384.
72. Richani K, Soto E, Romero R, Han Y, Pineles B, Kim YM, Cushenberry E, Yoon BH, Kusanovic J, Kim CJ. Decreased mRNA expression of complement regulatory proteins in chorioamnionitis. *Am J Obstet Gynecol* 2006;195 (Suppl.):S71.
73. Sharma A, Prabhakar P, Sharma DP, Jayasinghe RG. Immunoglobulin and C3 levels in normal human amniotic fluid. *West Indian Med J* 1983;32:140–146.
74. Elimian A, Figueroa R, Canterino J, Verma U, Aguero-Rosenfeld M, Tejani N. Amniotic fluid complement C3 as a marker of intra-amniotic infection. *Obstet Gynecol* 1998;92: 72–76.
75. Huffaker J, Witkin SS, Cutler L, Druzyn ML, Ledger WJ. Total complement activity in maternal sera, amniotic fluids and cord sera in women with premature labor, premature rupture of membranes or chorioamnionitis. *Surg Gynecol Obstet* 1989;168:397–401.
76. Haeger M, Bengtson A, Karlsson K, Heideman M. Complement activation and anaphylatoxin (C3a and C5a) formation in preeclampsia and by amniotic fluid. *Obstet Gynecol* 1989;73:551–556.
77. Keelan JA, Blumenstein M, Helliwell RJ, Sato TA, Marvin KW, Mitchell MD. Cytokines, prostaglandins and parturition – a review. *Placenta* 2003;24 (Suppl. A):S33–S46.
78. Romero R, Espinoza J, Goncalves LF, Kusanovic JP, Friel LA, Nien JK. Inflammation in preterm and term labour and delivery. *Semin Fetal Neonatal Med* 2006;11:317–326.
79. Siegel I, Gleicher N. Peripheral white blood cell alterations in early labor. *Diagn Gynecol Obstet* 1981;3:123–126.
80. Romero R, Brody DT, Oyarzun E, Mazor M, Wu YK, Hobbins JC, Durum SK. Infection and labor. III. Interleukin-1: a signal for the onset of parturition. *Am J Obstet Gynecol* 1989;160: 1117–1123.
81. Hebisch G, Neumaier-Wagner PM, Huch R, von Mandach U. Maternal serum interleukin-1  $\beta$ , -6 and -8 levels and potential determinants in pregnancy and peripartum. *J Perinat Med* 2004;32:475–480.
82. Buonocore G, De Filippo M, Gioia D, Picciolini E, Luzzi E, Bocci V, Bracci R. Maternal and neonatal plasma cytokine levels in relation to mode of delivery. *Biol Neonate* 1995;68:104–110.
83. Haddad R, Tromp G, Kuivaniemi H, Chaiworapongsa T, Kim YM, Mazor M, Romero R. Human spontaneous labor without histologic chorioamnionitis is characterized by an acute inflammation gene expression signature. *Am J Obstet Gynecol* 2006;195:394–324.
84. Romero R, Quintero R, Nores J, Avila C, Mazor M, Hanaoka S, Hagay Z, Merchant L, Hobbins JC. Amniotic fluid white blood cell count: a rapid and simple test to diagnose microbial invasion of the amniotic cavity and predict preterm delivery. *Am J Obstet Gynecol* 1991;165:821–830.
85. Romero R, Jimenez C, Lohda AK, Nores J, Hanaoka S, Avila C, Callahan R, Mazor M, Hobbins JC, Diamond MP. Amniotic fluid glucose concentration: a rapid and simple method for the detection of intraamniotic infection in preterm labor. *Am J Obstet Gynecol* 1990;163:968–974.
86. Romero R, Emamian M, Quintero R, Wan M, Hobbins JC, Mazor M, Edberg S. The value and limitations of the Gram stain examination in the diagnosis of intraamniotic infection. *Am J Obstet Gynecol* 1988;159:114–119.
87. Bobitt JR, Ledger WJ. Amniotic fluid analysis. Its role in maternal neonatal infection. *Obstet Gynecol* 1978;51:56–62.
88. Ahmed AE, Peter JB. Clinical utility of complement assessment. *Clin Diagn Lab Immunol* 1995;2:509–517.
89. Lynch AM, Gibbs RS, Murphy JR, Byers T, Neville MC, Giclas PC, Salmon JE, Van Hecke TM, Holers VM. Complement activation fragment Bb in early pregnancy and spontaneous preterm birth. *Am J Obstet Gynecol* 2008;199:354–358.
90. Vaisbuch E, Romero R, Erez O, Mazaki-Tovi S, Kusanovic JP, Soto E, Gotsch F, Dong Z, Chaiworapongsa T, Kim SW, et al. Fragment Bb in amniotic fluid: evidence for complement activation by the alternative pathway in women with intra-amniotic infection/inflammation. *J Matern Fetal Neonatal Med*, in press.