

Nanotoxicology



ISSN: 1743-5390 (Print) 1743-5404 (Online) Journal homepage: informahealthcare.com/journals/inan20

In vitro models of the human placental barrier – In regione caecorum rex est luscus

Sara Correia Carreira, Laura Walker, Kai Paul & Margaret Saunders

To cite this article: Sara Correia Carreira, Laura Walker, Kai Paul & Margaret Saunders (2015) In vitro models of the human placental barrier - In regione caecorum rex est luscus, Nanotoxicology, 9:sup1, 135-136, DOI: 10.3109/17435390.2013.869628

To link to this article: <u>https://doi.org/10.3109/17435390.2013.869628</u>



Published online: 29 Apr 2015.



Submit your article to this journal 🗹





View related articles 🗹



則 View Crossmark data 🗹

Nanotoxicology

Nanotoxicology, 2015; 9(S1): 135–136 © 2015 Informa UK Ltd. DOI: 10.3109/17435390.2013.869628

LETTER TO THE EDITOR

In vitro models of the human placental barrier – *In regione caecorum rex est luscus*

Sara Correia Carreira¹, Laura Walker², Kai Paul³, and Margaret Saunders⁴

¹Bristol Centre for Functional Nanomaterials, University of Bristol, Bristol, UK, ²School of Clinical Sciences, Bristol Heart Institute, University of Bristol, Bristol, UK, ³Nano Safety Research Group, School of Life Sciences, Heriot-Watt University, Edinburgh, UK, and ⁴Department of Medical Physics & Bioengineering, Bioengineering, Innovation & Research Hub (BIRCH), St Michael's Hospital, University Hospitals Bristol NHS Foundation Trust, Bristol, UK

The development of engineered nanomaterials has been on a steep rise in the last 10 years, and many of these new materials have found their way into consumer products (PEN, 2013). Given the overwhelming number of newly developed nanomaterials, the cost of rigorous toxicological assessment of all of them is estimated to be enormous (Choi et al., 2009). Therefore, resourceful testing strategies are essential to overcome the associated cost of toxicity testing while at the same time achieving the highest possible degree of safety. In this endeavor, high throughput screening tools are paramount for rapid preliminary assessment of a variety of nanomaterials. Despite being a relatively simple model of the human placental barrier, the easily maintainable choriocarcinoma derived cell line BeWo has been invaluable as a screening tool for a variety of xenobiotics (Levkovitz et al., 2013; Prouillac & Lecoeur, 2010; Saunders, 2009). Despite the fact that the cell line is malignant in origin, it has been demonstrated that the BeWo cell model has many similarities to normal trophoblast cells, which are of relevance to both toxicity and transport assessment. These similarities include placental hormone secretion (Sastry, 1996), close cell apposition and apical microvillous projections (Bode et al., 2006), and the ability to form a confluent monolayer with functional polarity (Bode et al., 2006; Cariappa et al., 2003). BeWo cells have also been shown to differentially express enzymes and transporters such as cytochrome P450 and Pgp with responses similar to trophoblasts in vivo (Jin & Audus, 2005; Thadani et al., 2004; Vähäkangas & Myllynen, 2006).

With the rapid development of nanotechnology there is a need for a model that can be used in preliminary toxicological screening for assessing safety of nanoparticle exposure during pregnancy. It has been estimated that a successful placental perfusion experiment is achieved every two weeks (Günday & Schneider, 2012), which is an unacceptable cost if this model alone was to be employed for nanotoxicological assessment. This model also requires large amounts of nanoparticles because of the circulation volume required and it is dependent upon the provision of placentae that are in good condition and provided in a timely manner. Using the BeWo model, areas of particular concern can rapidly be identified and subsequently tested in more sophisticated models which are less easily accessible and therefore more costly, both in terms of time and resources. For example, using the BeWo model we recently found that the toxicological profile of Fe_3O_4 nanoparticles (NPs) with different surface coatings resulted in a more complex dose and timedependent response than exposure to different sizes of silica NPs (Correia Carreira et al., 2015). This finding allows us to allocate resources for further testing more efficiently.

informa

healthcare

The transport of a variety of chemicals studied both in the placental perfusion and the BeWo model have been found to be in good agreement (Correia Carreira et al., 2011; Li et al., 2013; Poulsen et al., 2009) which is a very promising result. Due to their colloidal nature, however, nanoparticles may sediment and therefore lead to more discrepancies in both models. Further work in developing and refining the BeWo model is undoubtedly required to adapt to the assessment of particulate as opposed to soluble agents. Nevertheless, using the BeWo model as a rapid screening tool proceeding *ex vivo* placental perfusion studies allows refinement of experimental conditions, such as determining appropriate concentration ranges. This enables the more scarce experimental resources such as intact perfused placentae to then be used to maximum advantage.

Placental perfusion and placental explant models are almost entirely restricted to term placenta, unless placentae from pregnancy terminations can be obtained which brings more complex ethical considerations into play). These models are therefore limited to assessing nanoparticle transport in late pregnancy but no conclusions can be drawn for the early, most sensitive stages of fetal development. The early placenta possesses multiple trophoblast layers whereas third trimester placenta bear a single trophoblast layer. Primary trophoblast cells can be isolated from human placental tissue but contamination with other cell types requires good separation and purification procedures and the formation of an intact polarized monolayer has proven difficult to achieve for transport studies (Vähäkangas & Myllynen, 2006). A multilayered trophoblast can be simulated using BeWo cells, and this model has in fact shown that nanoparticle toxicity was more pronounced in BeWo multilayersa simple model of early placentae compared to a BeWo monolayer (Sood et al., 2011). Thanks to the flexibility of the BeWo model, it is possible to make a discovery like this which can be subsequently verified using a more sophisticated and

Correspondence: Dr Margaret Saunders, Department of Medical Physics and Bioengineering, BIRCH, St Michael's Hospital, UH Bristol NHS Foundation Trust, Southwell St, Bristol BS2 8EG, UK. Tel: +44 117 342 5685. E-mail: M.Saunders@bristol.ac.uk

hard-to-come-by model such as an early trimester placental explants (Miller et al., 2005). Issues such as sedimentation of particles can be addressed through the use of appropriate equipment to maintain particles in suspension.

By no means is the BeWo model to replace more complex models such as ex vivo placental perfusion or placental explants culture. No single model is going to provide us with all the answers to what the biological outcomes will be of gestational exposure to nanoparticles. Ultimately, we will need to work with a combination of models, including assessment of NP toxicity in the most appropriate in vivo model, to advance the field and to provide us with the answers that are needed. However, given the high cost and limited availability of such complex models on the one hand, and the overwhelming number of nanomaterials that require safety assessment on the other, alternative and less costly models are needed which can provide us with a rapid throughput to be able to test a number of variables in a short period of time under replicable conditions. As put by the Renaissance humanist Erasmus of Rotterdam: "In regione caecorum rex est luscus" (In the land of the blind, the one-eyed man is king). The "oneeyed view" on placental transport and toxicity we gain from the BeWo model enables us to begin to tackle the enormous task of rigorous nanoparticle safety assessment in pregnancy. This would hardly be possible if we were solely relying on the "two-eyed view" we may obtain from more complex models.

Declaration of interest

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of this article.

References

- Bode CJ, Jin H, Rytting E, Silverstein PS, Young AM, Audus KL. 2006. In vitro models for studying trophoblast transcellular transport. Methods Mol Med 122:225–39.
- Cariappa R, Heath-Monnig E, Smith CH. 2003. Isoforms of amino acid transporters in placental syncytiotrophoblast: plasma membrane localization and potential role in maternal/fetal transport. Placenta 24: 713–26.
- Choi J-Y, Ramachandran G, Kandlikar M. 2009. The impact of toxicity testing costs on nanomaterial regulation. Environ Sci Technol 43: 3030–4.

- Correia Carreira S, Cartwright L, Mathiesen L, Knudsen LE, Saunders M. 2011. Studying placental transfer of highly purified non-dioxin-like PCBs in two models of the placental barrier. Placenta 32:283–91.
- Correia Carreira S, Walker L, Paul K, Saunders M. 2015. The toxicity, transport and uptake of nanoparticles in the in vitro BeWo b30 placental cell barrier model used within NanoTEST. Nanotoxicology 9(S1):66–78.
- Günday N, Schneider M. 2012. Conference scene: nanomedicine and nanotoxicology: future prospects and the need for translational factors for the combination of both. Nanomedicine (London, England) 7: 811–14.
- Jin H, Audus KL. 2005. Effect of bisphenol A on drug efflux in BeWo, a human trophoblast-like cell line. Placenta 26:S96–103.
- Levkovitz R, Zaretsky U, Gordon Z, Jaffa AJ, Elad D. 2013. In vitro simulation of placental transport: Part I. Biological model of the placental barrier. Placenta 34:699–707.
- Li H, van Ravenzwaay B, Rietjens IM, Louisse J. 2013. Assessment of an in vitro transport model using BeWo b30 cells to predict placental transfer of compounds. Arch Toxicol 87:1661–9.
- Miller RK, Genbacev O, Turner MA, Aplin JD, Caniggia I, Huppertz B. 2005. Human placental explants in culture: approaches and assessments. Placenta 26:439–48.
- PEN. 2013. The Project on Emerging Nanotechnologies. Consumer Products Inventory. Available at: http://www.nanotechproject.org/cpi/. Last accessed on Nov 11, 2013.
- Poulsen MS, Rytting E, Mose T, Knudsen LE. 2009. Modeling placental transport: correlation of in vitro BeWo cell permeability and ex vivo human placental perfusion. Toxicol In Vitro 23:1380–6.
- Prouillac C, Lecoeur S. 2010. The role of the placenta in fetal exposure to xenobiotics: importance of membrane transporters and human models for transfer studies. Drug Metab Dispos 38:1623–35.
- Sastry BVR. 1996. Techniques: cultured tissues and cells to study placental function. In: Sastry BVR, ed. Placental Pharmacology. London: CRC Press, 47–66.
- Saunders M. 2009. Transplacental transport of nanomaterials. Wiley Interdiscip Rev Nanomed Nanobiotechnol 1:671–84.
- Sood A, Salih S, Roh D, Lacharme-Lora L, Parry M, Hardiman B, et al. 2011. Signalling of DNA damage and cytokines across cell barriers exposed to nanoparticles depends on barrier thickness. Nat Nanotechnol 6:824–33.
- Thadani PV, Strauss III JF, Dey SK, Anderson VM, Audus KL, Coats KS, et al. 2004. National Institute on Drug Abuse Conference report on placental proteins, drug transport, and fetal development. Am J Obstet Gynecol 191:1858–62.
- Vähäkangas K, Myllynen P. 2006. Experimental methods to study human transplacental exposure to genotoxic agents. Mutat Res 608:129–35.